**Project Title** 

Field grown horticultural crops. A costed study in the use of selected green manures/biofumigants to control selected nematode pests and pythia and their influence on soil nutrition status.

Project number:

FV 273

Project leader:

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Report:

Final report, March 2006

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Location of project:

CSL and two field sites on commercial

farms in Norfolk.

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Biofumigant, biocidal crop, *Brassica* juncea, cultivations, fallow, free-living nematodes, green manure, mustard.

pythia, soil nutrition.

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The results and conclusions in this report are based on two trials conducted over a one-year period. The conditions under which the experiments were carried out and the results have been reported in detail and with accuracy. However, because of the biological nature of the work it must be borne in mind that different circumstances and conditions could produce different results. Therefore, care must be taken with interpretation of the results, especially if they are used as the basis for commercial product recommendations.

## **AUTHENTICATION**

We declare that this work was done under our supervision according to the procedures described herein and that the report represents a true and accurate record of the results obtained.

[Name]

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# **Grower Summary**

#### Headlines

- Caliente Brand Mustard 119 replaced Nemat *Eruca sativa* as a biocidal crop in the trial.
- The Mustards increased the levels of plant-parasitic nematodes during the growing period and but reduced them after incorporation.
- However, there was no overall significant reduction of plant-parasitic nematodes by either Mustard 99 or Mustard 119 (each different blends of Brassica juncea) compared to fallow plots.
- There was no overall significant reduction of pythia by the biocidal crops compared to fallow plots.
- Overall, few significant differences were found between biocide crop areas and uncultivated fallow for K, Mg, P or pH during the project.
- The biocide crops appear to capture N and will thus serve to assist in its management and delay leaching.
- Both Caliente Brand Mustards 99 and 119 produced total ground cover canopy in three weeks and good weed suppression was observed, for the entire nine week life of the crops.
- The effect of the biocidal crops was not the same at each site, supporting
  evidence that the ability of such crops to reduce pest pressure and improve soil
  quality will vary with factors such as soil type, management techniques and
  weather conditions.
- Long-term studies of the regular use of biocidal crops are necessary to assess their full potential.

## Background and expected deliverables

Certain pesticides such as Temik are often used prophylactically to control plant-parasitic nematodes, but whether such use is warranted has been the subject of recent HDC research (HDC Reports FV232, FV249) and debate (HDC Report FV 278). Whilst this product is due to be revoked on 31 December 2007, alternative chemical products may be used.

Intensive root crop rotations on light/medium soil types with irrigation are leading to higher levels of soil-borne fungal pathogens and novel methods of control are being sought.

Assured Produce Schemes are encouraging the use of sustainable pest management practices, such as the inclusion of poor or non-host crops (being investigated in The Netherlands (Korthals *et al.*, 2004) or green manures or biofumigants (collectively called 'biocidal crops) which are now being marketed in the UK. Generally speaking, green manures may serve several functions including improvement of soil conditions, whilst biofumigants have properties that are used specifically for the control of pathogens, although they may also, incidentally, improve soil structure. Both crops have potential as weed suppressants. However, there is a need for an independent assessment of the benefits and disadvantages of such crops, especially for the control of pests and diseases. Such work is being carried out worldwide, but there has been little investigation in UK conditions.

This project sought to investigate and quantify the effectiveness of two biocidal crops being sold as green manures and biofumigants, namely Caliente Brand Mustards 99 and 119 (hereafter called Mustard 99 and Mustard 119), in controlling plant-parasitic nematodes and pythia as well as their influence on the nutritional status of the soil. Mustard 99 is *Brassica juncea* ISCI 99, and is particularly high in glucosinolates. The particular blend of glucosinolates and enzymes will, in theory, affect the amount of isothiocyanates that are produced when the crop is chopped and incorporated and hence the level of kill of plant-parasitic nematodes and other pathogens. Mustard 119 is a blend of *Brassica juncea* and *Sinapsis alba*, with ISCI 20 being the predominant species in the blend, and was said to be the best all-round variety.

In such a short investigation the results can only suggest the consequences of using such crops and their role in developing an integrated crop management system that would offer a more sustainable option for the future control of pathogens in vegetable crops.

## Summary of the project and main conclusions

- There were higher levels of plant-parasitic nematodes in the Mustard areas
  than the fallow areas by pre-incorporation (crop at full crop canopy, four
  weeks from first flower), despite the latter supporting a range of weeds that
  could act as nematode hosts
- Also at pre-incorporation, there was a significant increase in the number of stubby-root nematodes in Mustard 99 compared to the fallow plots, but this effect was not detected with the Mustard 119 crop.
- Whilst the biofumigant effect of the incorporated biocidal crops significantly reduced their higher levels of nematodes, it was not sufficient to reduce them below the levels found in the fallow plots six weeks post-incorporation, resulting in no differences between the treatments for total numbers of nematodes. The reduction in total nematode numbers was more noticeable at Knights than at Elveden.
- Overall there was no statistically significant difference between treatments for total pythia counts.
- The cultivation action on incorporation had itself no significant effect on nematode levels, although there was a small and just significant effect of cultivation on pythia in the Mustard 99 areas but not in the Mustard 119 areas.
- Nutrient values of soil from the cropping area were compared with soil from the uncultivated fallow plots for all sampling dates. Nitrogen was applied to all

plots after drilling, making the most important comparison for N that relating to variations between pre-incorporation and six weeks post-incorporation.

- At pre-drilling there were no significant differences in nutritional values across the plots or through the profiles, except for K in the Mustard 119 area, where the mean level was higher in the uncultivated fallow.
- The levels of P, K, Mg or pH did not change between pre-drilling and preincorporation.
- Between pre-incorporation and six weeks after incorporation K levels in Mustard 99 plots increased by 7.8% compared with a 6.3% drop for uncultivated fallow. This represents a capture of K during cropping then subsequent release after the crop was incorporated. However, such an effect did not occur with Mustard 119. There were no statistically significant differences in pH with either Mustard 99 or Mustard 119.
- Over the whole sampling period, however, the only changes were a slight decrease in pH in the uncultivated fallow compared with Mustard 99 whilst in Mustard 119 there was an increase of P and K.
- Between pre-drilling and pre-incorporation N values at both depths varied. The change was greatest in uncultivated fallow, where the levels of N were higher. This higher level of N, in the uncultivated fallow, when compared with the cropped areas, reflects the biocidal crops ability to capture N.
- The difference in levels of N between the two profiles (0-30cm and 30-60cm) was the same in both the biocidal crops and the uncultivated fallow at predrilling and six weeks post-incorporation. At pre-incorporation, there were no differences with Mustard 99 but with Mustard 119 there was a slight difference caused by just significantly higher N in the cropped area compared to the fallow.

- Between pre-incorporation and six weeks after incorporation there was a significant increase of N in the shallow profile with both biocidal crops compared with a small loss in uncultivated fallow. In the deeper profile there was a greater difference between the increase of N in Mustard 99 and the small loss with the associated uncultivated fallow but with Mustard 119 there was no significant difference. Comparing real values of N, variations were found between sites; at Elveden there was a mean increased availability behind Mustards of 80kg N/Ha (indicating effective trapping and then release). The associated fallow showed a mean decrease or 61kgN/Ha (indicating loss by leaching). For the same period at Knights there was a mean increase of 35kgN/Ha behind Mustards (again showing these crops' ability to trap and release N). However, the fallow here also registered a small increased availability of 6kgN/Ha. These results suggest that environmental or site variations are important. There is a need for extending the sampling period post-incorporation to gain better understanding of the period of release (mineralisation) of N from green manures.
- Both Caliente Brand Mustards 99 and 119 produced total ground cover canopy in three weeks, resulting in good weed suppression compared to the fallow areas. This effect was also noticeable six weeks after incorporation.
- The results supported previous suggestions that individual biocide 'brands'
  may differ in their particular effects, each 'brand' being a blend of different
  cultivars in order to maximize particular effects.
- A range of factors, such as differences in management techniques and environmental factors, probably contributed to site differences in nematode control and N levels seen in this project. Soil type varied; both sites are glacial deposits, with fields at Elveden being a deep loamy sand over chalk but in places over deep sand and occasionally small areas of clay, whilst Knights Top Battle is a sandy loam (but not so deep) over chalk with pockets of deep sand. Thus the latter site has potentially the most moisture retentive soil. Such

factors will affect the ability of biocidal crops to reduce pest pressure or improve soil quality.

- Overwintering a biocidal crop, or drilling and incorporating it close to drilling may provide benefits not seen here, providing cress tests indicate no phytotoxic effects are likely.
- Such a relatively small investigation can only give an indication of the effects
  of the selected biocidal crops on soil quality and pests and diseases of specific
  interest to carrot growers. Further work is needed to clarify the effects of
  biocidal crops in a sustainable rotation, and their contribution to a long-term
  improvement in both soil condition and pest and disease control.

#### Financial benefits

A relatively small investigation such as this will only provide an indication of the potential of the selected biocidal crops. However, the findings could result in more productive use of Defra grants. The costs associated with the use of mustard crops are set out below, but a full assessment of benefits is not possible as sampling did not continue to eventual cropping of the field sites.

Table 1. The cost of establishing and management of Mustard crops to include incorporation. Expressed as cost per Ha for commercial areas at contract/farmer rates (associated costs from A.B.C. Farm Machinery Costs Book).

Variables:	Comments	Cost
Seed	ISCI 99 @ 10Kg/Ha x	£88.00
	£8.80	
	199 @ 15Kg/Ha x	£90.00
	£6.00	
Fertilizer:	Nitrogen @140KgN/Ha	£72.20
	x £0.53	
Total Variables:		£72.20
Operations:		
Plough/Press		£40.00/33.00
Combi Drill		£37.25/29.50
Fertilizer Spreading		£ 6.00/ 5.00
Roll	,	£14.00/10.00
Irrigate 30mm x £2.2		£66.00
Mulch		£16.00/12.00
Disc/Press		£31.00/26.50
Total cost		£371.45/343.20

The most likely cost to change is irrigation; the 30mm applied is based on experience from the project with most applied at time of incorporation.

Biocidal crops offer savings as part of a soil management/rotation enhancing policy. Cultivations of fallow land x 3 @ £16.60/12.90 =£49.80/38.70

Fertilizer input to the following crop will also be reduced. A small increase in organic matter will help soil management (a requirement of Single Farm Payment) with possible reductions in levels of erosion, both water and wind.

#### Action points for growers

If growers are considering incorporating biocidal crops into a rotation they should note that

- Significant benefits of biocidal crops compared with uncultivated fallow for nematode and pythia control have not been seen in this project
- Biocidal crops may assist in the retention of N for the benefit of subsequent crops, thus may have a place in rotation
- Biocidal crops may offer a sustainable method of weed control between crops
- Overwintering a biocidal crop, or drilling and incorporating it close to drilling may provide benefits not seen here, providing cress tests indicate no phytotoxic effects are likely.

#### Acknowledgements

We are grateful for the assistance of the following companies for the use of their sites and assistance with ground work:

Elveden Farms Ltd, Estate Office, Elveden, Thetford, Norfolk, IP24 3TQ

Knights Farms Ltd, The Estate Office, Lower Farm, Narborough, Kings Lynn, Norfolk, PE32 1JB

We are also grateful to Tom Will and Emily Heading (Vegetable Consultancy Services), James Quill and all staff involved with extraction of nematodes and *Pythium* from soils (Central Science Laboratory) for their invaluable assistance with this project.

Lastly, for supplying the seed free of charge, our thanks go to Plant Solutions Limited, Pyports, Downside Bridge Road, Cobham, Surrey KT11 3EH

#### **Science Section**

#### Introduction

Certain pesticides such as Temik are often used prophylactically to control plant-parasitic nematodes, but whether such use is warranted has been the subject of recent HDC research (HDC Reports 232, 249) and debate (HDC Report FV 278). Whilst Temik is due to be revoked on 31 December 2007, alternative chemical products may be used.

A literature review prepared for the HDC in conjunction with the proposal for this project (Hockland, 2005; Appendix 1) concluded that chemicals derived from biofumigant crops offered an additional sustainable control tool to reduce pathogens and weeds. Intensive root crop rotations on light/medium soil types with irrigation are leading to higher levels of soil-borne fungal pathogens and thus novel methods of control are also being sought for diseases.

A range of biocidal crops is being marketed in the UK, but none offer blanket control of all pathogens and the effectiveness of many biocidal crops is difficult to predict. There has been no scientific or costed assessment of the field use of biofumigants in horticultural crops in the UK, thus lending support to this investigation. Concurrently, new formulations of biocidal plant products are being developed (such as a pelleted form), making their use more versatile, so growers also need to consider if they need the potential advantages of growing such crops as a green manure for improving soil structure or as a crop cover for weed control.

This project sought to investigate and quantify the effectiveness of two biocidal crops being sold as green manures and biofumigants, namely Caliente Brand Mustards 99 and 119 (hereafter called Mustard 99 and Mustard 119), in controlling plant-parasitic nematodes and pythia as well as their influence on the nutritional status of the soil. Mustard 99 is *Brassica juncea* ISCI 99, and is particularly high in glucosinolates. The particular blend of glucosinolates and enzymes will, in theory, affect the amount of isothiocyanates that are produced when the crop is chopped and incorporated and hence the level of kill of plant-parasitic nematodes and other pathogens. Mustard 119

is a blend of *Brassica juncea* and *Sinapsis alba*, with ISCI 20 being the predominant species in the blend, and was said to be the best all-round variety (Anon, 2004).

In such a short investigation the results can only suggest the consequences of using such crops and their role in developing an integrated crop management system that would offer a more sustainable option for the future control of pathogens in vegetable crops.

#### Materials and Methods

On 7 April 2005 soil sampling took place in Norfolk to determine those fields likely to offer the best combination of plant-parasitic nematodes and pythia for assessment purposes. The trials eventually took place on two sites, namely Elveden Waterloo and Knights Top Battle, and the sequence of events is set out in Table 2. On each site the trial followed a fully randomized replicated plot design, using four replicates of each treatment, which were 'Mustard 99', 'Mustard 119', and fallow, either with cultivation only at incorporation or with no cultivation at all. Soil samples for nematode, pythia and nutritional analysis were taken pre-drilling in June, pre-incorporation of the biocidal crops in August and six weeks post-incorporation in September.

Table 2. Sequence of events at Elveden Waterloo and Knights Top Battle, Norfolk

Date	Task	Site
week beginning 23.05.05	Sites ploughed and pressed	.
24/25.05.05	Sites marked out	
01.06.05	First full plot sampling	Elveden
	·	(dry)
02.06.05		Knights
		(wet)
03.06.05	Drilling and rolling	Both
		(good
		moisture)
Pre- drilling	N application 140Kg N both sites	Knights
Post-drilling	·	Elveden
Immediately post drilling -		both sites
heavy rain		
09/10.06.05	Crop emergence	Both
w.c. 20.06.05	Crop 3 / 4 TL	Both
w.c. 27.06.05	Crop total ground cover	Both

Date	Task	Site
w.c. 04.07.05	Crop stem extension	Both
w.c. 11.07.05	Crop flowering commenced	Both
15.07.05	High level caterpillar & some aphid - Sprayed Hallmark	Knights
30.07.05	Irrigated 25mm	Elveden
03.08.05	Pre-incorporation plot sampling	Elveden
04.08.05		Knights
03.08.05	Crop chopped & incorporated; control plots requiring cultivation also done	Elveden
05.08.05	Crop chopped & incorporated; control plots requiring cultivation also done	Knights
06/07.08.05	Irrigated 20mm	Knights
06.09.05	Weed population & growth survey	Both
14.09.05	Final plot sampling (6 weeks post incorp)	Elveden
15.09.05		Knights

#### Soil sampling and site selection

Soil samples were taken using a 'cheese corer' type auger, taking at least 40 cores up to a depth of 30cm along a W-path across a field or plot. For Nitrogen samples a deeper core of soil was taken and a surface to 30cm section was separated from the 30-60cm section to provide two samples for analysis.

Extraction and recording of plant-parasitic nematode species
Soil samples were processed using the Whitehead Tray method and the numbers and genera
of the free-living plant-parasitic species were recorded for each plot.

#### Isolation of pythia from soil

A sub-sample of the soils used for the determination of nematodes was used for the detection of *Pythium* species.

A 30g sub-sample was weighed from each main soil sample into 300 mL glass bottles and 250 mL of de-ionised water added. The soil/water solution was shaken and left to soak for one hour. Following the period of soaking the solution was shaken for a further two minutes using a Stuart flask shaker set at maximum.

All soil solutions were then diluted 1:10 using sterile de-ionised water and 0.1 mL of the neat and diluted soil solutions spread plated onto separate plates of the *Pythium/Phytophthora* selective agar PARP (Jeffers and Martin, 1986; Cornmeal Agar (CMA)

17 g/L, Pimaricin 5 mg, sodium ampicillin 250 mg, Rifampicin 10 mg dissolved in 1ml dimethyl sulfoxide (DMSO), pentachloronitrobenzene (PCNB) 100 mg. All amendments were either suspended or dissolved in 10 ml of sterile distilled water and added to CMA after it had been autoclaved and cooled to 50°C in a water bath).

Plates were incubated at 20°C and after a period of 36-48 hours the number of colonies growing counted. Colonies were transferred onto Potato Dextrose Agar (PDA) and incubated at 20°C for 3 days. To allow differentiation between the colonies they were then sub-cultured onto CMA. Both CMA and PDA plates were returned to the 20°C incubator for a further 7 days. Colonies were assessed microscopically for production of spores and associated structures, in addition growth pattern and growth rate were recorded. For each group of 'presumptive pythia' the number of colonies were counted and the number of colony forming unit per gram of soil calculated.

To try and encourage spore production a number of the colonies isolated were also grown on pieces of grass (sterilised by boiling for 5 min) floating in a 50:50 mix of pond and de-ionised water.

#### Nutritional analysis

The basis of the analysis was as practiced by ASA, i.e. the Analysis of Agricultural Material (ADAS Reference Book 427). All soil samples were stored and transported in insulated containers.

#### Statistical analysis

Statistical analysis was by Analysis of Variance (anova) with contrasts, using Genstat 8. There were three treatments (Biocide, Cultivation, No Cultivation) with four replicates of each treatment at each of the two sites. There were two complete experiments conducted with each biocide crop at the same time, one with Mustard 99 and one with Mustard 119. In the analysis the sites were blocks and the two contrasts were Biocide versus Cultivation and Biocide versus No cultivation. The pythia and nematode counts for stunt and stubby nematodes were transformed to logarithms for the analyses (log(n+1) for (since these counts contained zeroes), which successfully

normalised the data and stabilised the variances, thus meeting the requirements of anova. The initial values for the soil nutritional data in June were compared by anova (Genstat) to see if they differed between the Crop and No Cultivation plots. The difference between N at 30 and 60cm was compared between treatments at each time point. Further comparisons of the difference between the August and September soil analyses was by anova with Mustards 99 and 119 as the two treatments and the two sites as blocks. The Mustard 99 and Mustard 119 trials were analysed separately.

#### **Results and Discussion**

Results of soil samples taken for site selection are set out in Appendix 2. The Contract required that the results were reviewed in early autumn to assess whether the sampling regime should be repeated in the spring, before cropping. As the results from the nematode sampling inferred the biocidal crops had had no significant effect on nematode numbers and virtually no effect on levels of pythia, it was decided not to proceed with further sampling.

#### Results of nematode extractions

Details of the numbers found of each nematode group are set out in Appendix 3A, 3B and 3C.

## Results of pythia isolations

The medium used was selective for both *Pythium* and *Phytophthora* species and the growth patterns of all the colonies isolated were consistent with these groups. The colonies isolated were labelled 'presumptive pythia', however some of these may have been *Phytophthora* species. Only group P2 of the 'presumptive pythia' was positively identified to species, this was *Pythium ultimum* (a well known pathogenic species).

## Effects of Biocides on Nematodes and pythia

The results of the sampling for total nematode numbers, total pythia, root-lesion and stubby-root nematodes are represented in Figs 1-8.

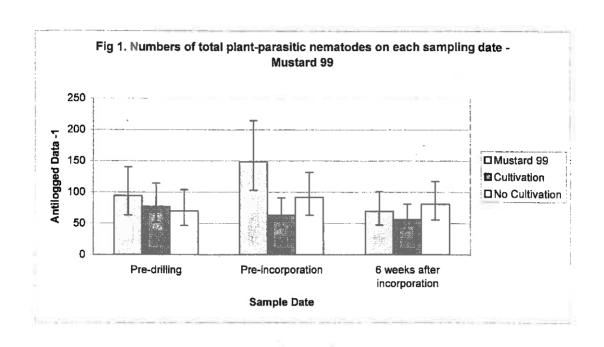
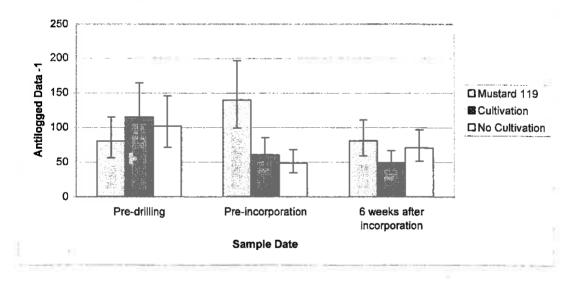


Fig 2. Numbers of total plant-parasitic nematodes on each sampling date - Mustard 119



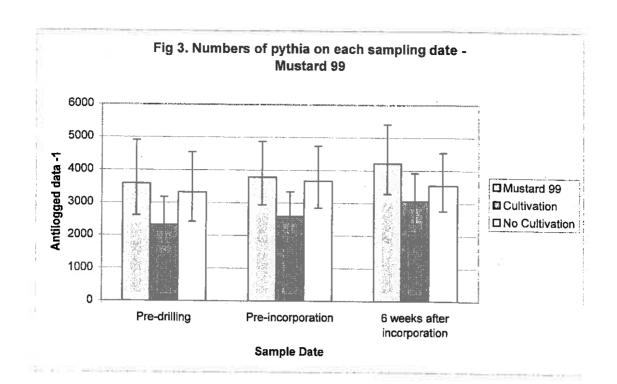
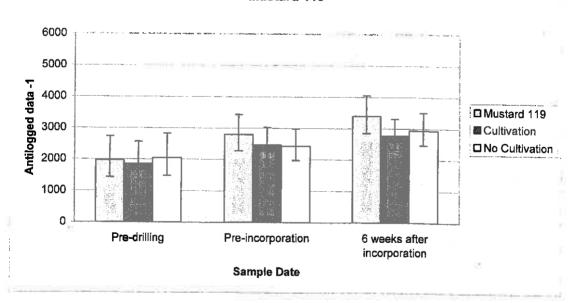


Fig 4. Numbers of pythia on each sampling date-Mustard 119



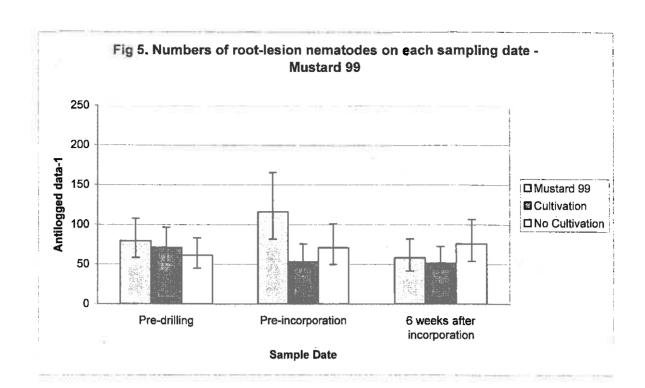
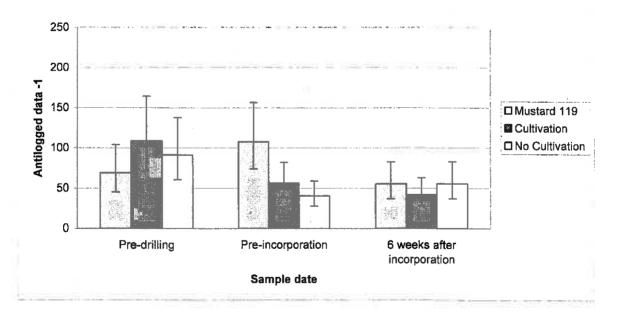


Fig 6. Numbers of root-lesion nematodes on each sampling date - Mustard 119



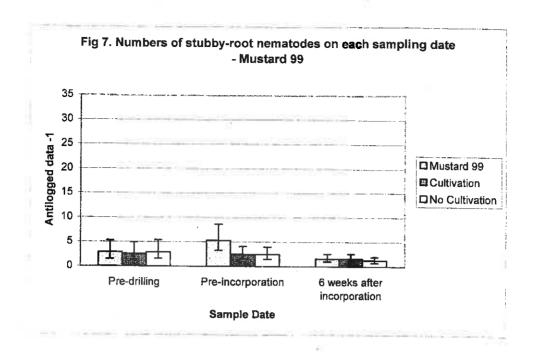
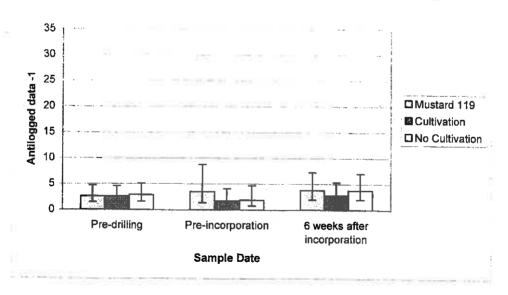


Fig 8. Numbers of stubby-root nematodes on each sampling date - Mustard 119



Pre-drilling soil samples produced no statistically significant differences between the three treatments (biocide, fallow with cultivation and fallow uncultivated) for either total nematodes (Figs 1-2.), total pythia (Figs 3-4.), root-lesion nematodes (Figs 5-6.), stubby nematodes (Figs 7-8.) or stunt nematodes, for either Mustard 99 or Mustard

119 (Appendix 4, Tables 1a and 1b). This was an ideal result on which to base subsequent sampling results.

Soil samples taken just before incorporation of the biocidal crops showed a significant difference between the biocide crop areas and fallow for total nematodes and root-lesion nematodes. This was due to there being significantly more nematodes in the Mustard 99 and Mustard 119 plots compared with the respective fallow (cultivated) plots and in the case of Mustard 119, in the fallow (uncultivated) plots. This was despite the fallow plots having a general cover of weeds that could also act as hosts for nematodes. There was a significant increase in the number of stubby-root nematodes in the Mustard 99 compared with the fallow plots, but this effect was not detected with Mustard 119 (see Appendix 4, Tables 1a and 1b).

Between pre-incorporation and six weeks post-incorporation there was a marked decrease in total nematode numbers with both biocide crops but not in the fallow plots (Appendix 4, Table 1c, Diff 2:3). Despite this, at six weeks post-incorporation there were no differences between the treatments for total numbers of nematodes or pythia. Changes in nematode numbers between pre-drilling and six weeks post-incorporation were compared between the treatments (referred to as Diff 1:3 in Appendix 4, Tables 1a and 1b). No significant differences were detected for any of the nematodes or pythia for either biocide crop (Appendix 4, Tables 1a and 1b). Thus despite the significant increase in nematode numbers in the biocide crop areas during cropping and the significant reduction of these levels after incorporation, there was no difference between nematode numbers in the biocide crop areas or the fallow areas at the end of the investigation.

Analysis of the reduction effect of the Mustards for the two sites showed it present at the Knights site but not at Elveden, thus illustrating a site effect which has been reported previously (HDC report FV 249). Differences in management techniques and environmental factors such as soil type and weather, probably contributed to site differences seen in this project. Soil type varied; both sites are glacial deposits, with fields at Elveden being a deep loamy sand over chalk and in places over deep sand and occasionally small areas of clay, whilst Knights Top Battle is a sandy loam (but not so

deep) over chalk with pockets of deep sand. Thus the latter site has potentially the most moisture retentive soil, with consequences for nematode and pathogen activity.

Overall, the level of pythia did not differ significantly between treatments over the life of the project, and there were no differences between treatments six weeks post-incorporation.

An analysis of the differences between the cultivated fallow and the uncultivated fallow produced no differences in nematode numbers; there was a small and just significant effect of cultivation on pythia in the Mustard 99 part of the trial but not in the Mustard 119 area. Cultivation may have an effect on weed control which may result in differences in soil moisture levels (undisturbed fallow land may have higher soil moisture levels than cultivated land).

This investigation only recorded information over a short period, but has shown that biocidal crops, before they are incorporated, may increase nematode numbers before they reduce them, with apparently no overall short-term benefit on control. In the USA, researchers have found a similar increase in free-living nematode species (Dale Gies, personal communication). Where they have seen meaningful reductions with Caliente Mustards for root-knot nematodes (*Meloidogyne* spp.), a group not recorded in this project and only of local importance in the UK.

It is possible that the Mustards were better hosts than the general weed hosts present. The variation observed in numbers of nematodes in the fallow plots may be due to a complex interaction of factors, such as the development (or not) of weed cover, with its provision of roots for nematode feeding and shade for the soil surface, and the effects of cultivation or not, which besides having a direct affect on the nematodes would also affect the moisture levels in the soil.

The two Mustard crops were each a different blend of *Brassica* plants, with different levels of glucosinolates and enzymes which were said to result in different benefits. However, the control of pathogens was disappointing overall in the period offered by this study but the reasons for this are not clear. Specific benefits promoted for selected

crops may, in practice, be difficult to achieve, given the known variation with different soil types, management techniques and weather conditions. Incorporation was done as advised by the suppliers and efforts were made to provide the essential soil moisture to facilitate the release of isothiocyanates and produce an effective seal.

#### Soil nutrients

Nutrient values of soil from the cropping area were compared with soil from the uncultivated fallow plots for all sampling dates. Nitrogen (N) was applied to all plots after drilling so the most important comparison for N relates to variation between pre-incorporation and six weeks post-incorporation.

The initial values at pre-drilling were compared by anova (Genstat) to see if they differed between the biocide crops and uncultivated fallow plots (Appendix 5, Table 3). There were no statistically significant differences, including for N at both 30cm and 60cm depths, except for K in the Mustard 119 area, where the mean level was higher in the uncultivated fallow.

There were no significant differences in P, K, Mg or pH in the biocide areas or the uncultivated fallow when sampled at pre-incorporation or six weeks post-incorporation (Appendix 5). The only difference recorded for N was at pre-incorporation, caused by a just significantly higher N in the Mustard 119 area compared to the fallow. It was expected that the crops would facilitate the capture of N but this was not always the case here, and this may have been affected by environmental or site factors.

Despite there being no obvious differences between treatments at each sampling date there were some differences in nutrient levels (including N) between sampling dates and these were examined in more detail.

The levels of P, K or Mg, or pH did not change between pre-drilling and preincorporation for either the biocide crops or the fallow areas but overall levels of N did change during this period at both 30cm and 60cm depths, with the change being greater in uncultivated fallow soil (with higher levels of N) than in the biocide crops soil, reflecting the ability of the plants to capture N and therefore assist with its management and delay or reduce leaching (Appendix 5, Table 6). However, this effect was less pronounced at Knights, indicating an effect of environmental or other site factors (Figs 9-16).

Between pre-drilling and six weeks post incorporation the only change with Mustard 99 was a slight decrease in pH in the uncultivated fallow compared with the biocide. For the same period, the effect of Mustard 119 was an increase in P and K compared to uncultivated fallow plots.

Statistical analyses were performed for both the change in soil nutrients (including N) between pre-incorporation and six weeks post-incorporation and the proportionate change (Appendix 5). Examination of the residuals in the analyses of proportionate values showed no sign of non-normality and hence transformation was not necessary. There was a significant change in K with Mustard 99 (7.8% increase) and uncultivated fallow (6.3% drop). This probably represents a capture of K during cropping then a subsequent release after the crop was incorporated. However, such an effect did not occur with Mustard 119. There were no statistically significant differences in Mg or P but with Mustard 119 the percentage change in P (%23P) differed but not the actual change (diff23P). There were no statistically significant differences in pH with either Mustard 99 or Mustard 119.

For the same period, between pre-incorporation and six weeks after incorporation, there was a significant increase in N at 30cm depth with both Mustard 99 and Mustard 119 compared with a small loss in the fallow. This suggests that the incorporated crops helped to retain N whereas in the fallow areas N was leached out.

For N at 60cm depth there was a statistically significant difference with a large increase for Mustard 99 and a small loss in the fallow. This difference was more pronounced than at 30cm depth, probably because there had been a continual loss of N from the fallow areas but in the cropped areas six weeks after incorporation the N had probably only just started to leach from the surface layers. However, although there was also a large increase with Mustard 119, there was also a (smaller) increase in the

uncultivated fallow, resulting in there being no statistically significant difference between Mustard 119 and uncultivated fallow.

Overall, the benefit of incorporating biocidal crops for soil nutrition, compared with leaving the land fallow was not apparent. The statistically significant results with N were heavily influenced by the results at Elveden; at the Knights site the differences in N between the biocide area and the fallow area were far less obvious (Figs 9-16). Thus the characteristics of a particular field (due to environmental factors or management techniques) may have an effect on nutrition as well as pathogen control.

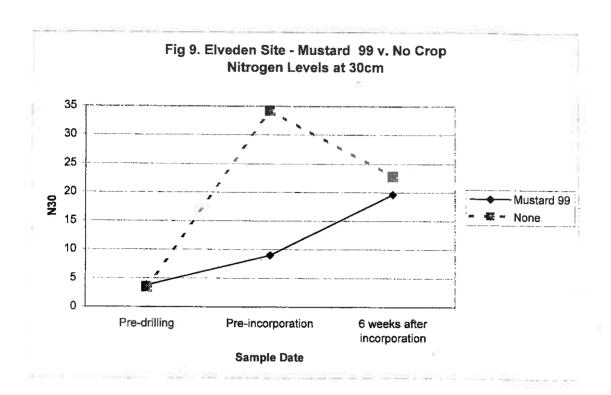
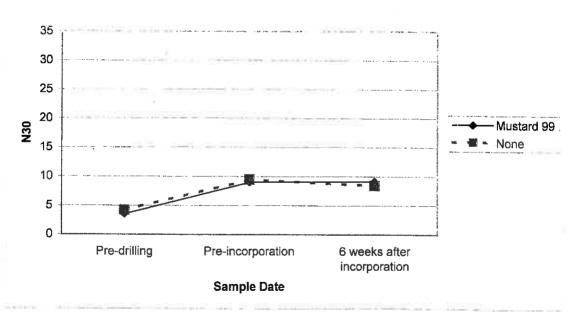


Fig 10. Knights Site - Mustard 99 v. No Crop Nitrogen Levels at 30cm



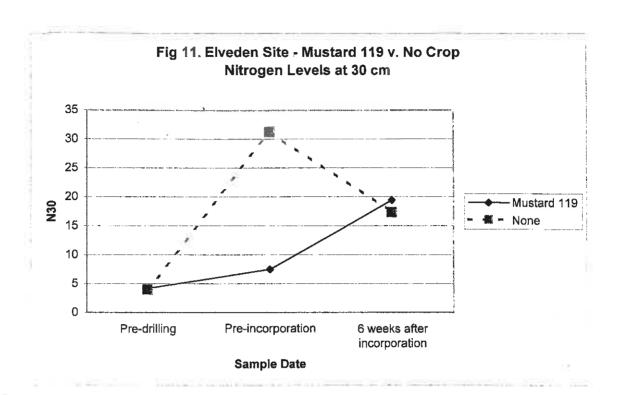
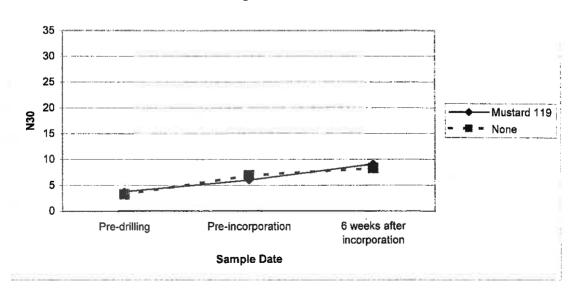


Fig 12. Knights Site - Mustard 119 v. No Crop Nitrogen Levels at 30cm



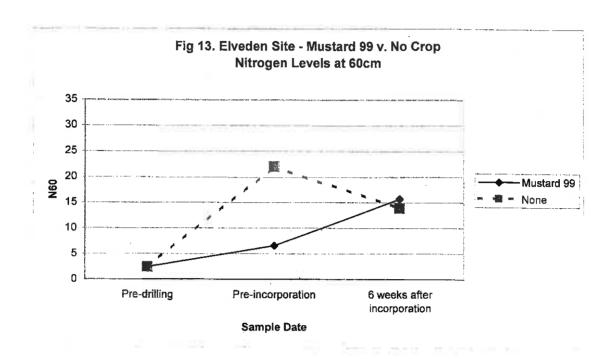
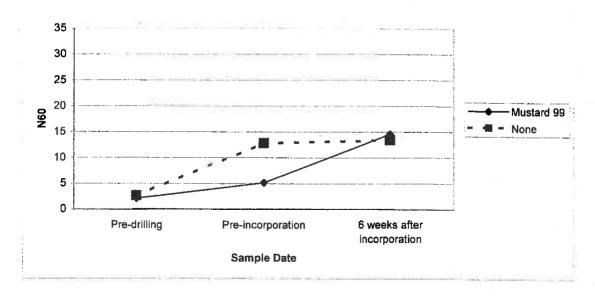


Fig 14. Knights Site - Mustard 99 v. No Crop Nitrogen Levels at 60cm



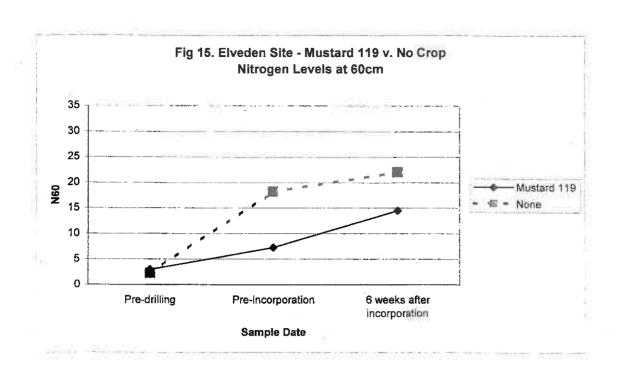
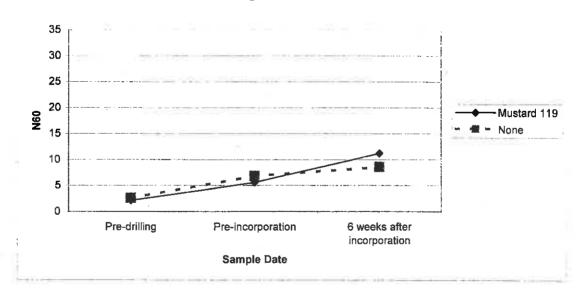


Fig 16. Knights Site - Mustard 119 v. No Crop Nitrogen Levels at 60cm



## Weed Suppression

Field observations recorded that both Mustard 99 and 119 produced total ground cover canopy in three weeks providing good suppression of weeds, including volunteer potatoes, compared to the fallow areas. They were also the dominant plants

up to incorporation and post-incorporation weed suppression remained obvious until the last sampling; weed emergence after Mustards was delayed by seven days and growth was then slower both in terms of weed height and maturity during the sixweek period. These crops therefore offer promise for weed suppression.

#### **Conclusions**

This project showed no significant benefits of an early summer planting and late summer incorporation of the selected biocidal crops compared with uncultivated fallow for nematode and *Pythia* control. However, the project produced some evidence that biocidal crops may assist in the retention of N for the benefit of subsequent cropping and may have a place in the management of N. They may also offer a sustainable method of weed suppression but any possible benefits are likely to be affected by the soil type of the site selected and other environmental factors.

Overwintering a biocidal crop, or drilling and incorporating it close to drilling may provide benefits not seen here, providing tests for phytotoxicity, using cress, indicate no adverse effects are likely. The development of biocidal pellets offers the potential for an improvement in pathogen control, especially for nematodes, as the biofumigants produced would have an effect on a population not increasing on good host root systems. However, this technology would not offer the benefits of Nitrogen management or weed suppression.

It should be acknowledged that such a relatively small investigation such as this can only give an indication to the effects of the selected biocidal crops on the chosen pests and diseases of specific interest to carrot or vegetable growers. Repeated use of these crops with associated records of their benefits may need to be done to illustrate their full potential; in Jersey, brassica crops (but not *B. juncea*) were grown between potato crops for four years but disease assessments generally showed an inconsistent effect. However, many growers are now using these brassicas rather than ryegrass or barley as a break crop and reporting an improvement in tuber quality (R. Collier, personal communication). Most recent work in California using mustard crops in six field trials in processing tomatoes over three years has failed to show benefits for disease control or yield, although environmental benefits, such as the reduction of N leaching seen in

this project, may be achieved (Hartz et al., 2005). Thus further work is needed to clarify the effects of biocidal crops in a sustainable rotation, and their contribution to a long-term improvement in both soil condition and pest and disease control. Such an approach would be in line with the productive use of Defra grants to support a sustainable cropping system for horticultural crops.

#### Technology transfer

The literature review of the biocidal crops Mustard (*Brassica juncea*) and Wild Rocket (*Eruca sativa*) (Appendix 1) has been circulated to interested parties by the HDC. The results of this project have been discussed at a growers' meeting held by the HDC. A short article in the HDC News might be useful for growers if considered to be cost effective.

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# LITERATURE REVIEW OF THE BIOCIDAL CROPS MUSTARD (Brassica juncea) AND WILD ROCKET (Eruca sativa)

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March 2005

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## APPENDIX 1

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## Growers' Summary

- plant-derived chemicals from biofumigant crops offer an additional, sustainable control tool to reduce pathogens and weeds
- a range of biocidal crops are being marketed in the UK, but none offer blanket control of all pathogens, including those of particular interest, *Brassica juncea ISCI 99* and *Eruca sativa NEMAT*
- The effectiveness of many biocidal crops is difficult to predict
- There has been no scientific assessment of the use of biofumigants in horticultural crops in the UK
- Field assessments are required to refine a biofumigation strategy
- Seeding rates are 10kg/ha for ISCI 99 and 8kg/ha for NEMAT. Both will cost approximately £100 per ha. Establishment costs are estimated to be about £75 per ha.
- In the light of developing technology for formulations of biocidal plant products, growers need to consider the advantages of growing such crops solely as a green manure for soil structure or for crop cover for weed control
- The cost:benefit of using biocidal plants as pellets or as a meal will need to be assessed
- After many years research, biofumigants are now used routinely in the USA and parts of Europe

#### Introduction

In order to assess the potential of *Brassica juncea* (available in the UK as ISCI 99) and *Eruca sativa* (available in the UK as NEMAT), growers need to be aware of current research into the use of Brassicaceae crops generally as biofumigants. Many growers are familiar with manufactured products for soil sterilisation, used to reduce the incidence of pests (particularly plant-parasitic nematodes), diseases and weeds. The active ingredients in these commercial products include dazomet and metamsodium, which both release methyl isothiocyanates. The isothiocyanates are a group of volatile compounds that are also produced when plant tissue is damaged and most researchers believe their role in nature is to provide protection against pests and pathogens. They are produced when plant cells are damaged and the glucosinolates they contain react with water and the enzyme myrosinase at neutral pH (also found in plant tissue). However, there are different types of glucosinolates which vary in their reactivity and the quantity of isothiocyanates released. *Brassicaceae* species contain high levels of these glucosinolates and so offer potential as biofumigants for a natural release of isothiocyanates when the plants are chopped and used as a green manure.

Whilst biofumigation is the term generally used to describe the exploitation of the myrosinase-glucosinolates system, it is also used to describe the use of other plantderived chemicals for the control of pathogens. Nematode suppression has occurred following soil incorporation of cyanogenic sudangrass hybrids into soil before cropping with carrots (Widmer & Abawi, 1998), and has been shown to be correlated with the amount of free cyanide released into soil (Widmer & Abawi, 2002). Poultry manures also have potential to suppress nematodes whether through stimulation of antagonistic microbes (Kaplan et al., 1992) or by production of ammonia (Rodriguez-Kabana, 1986). Lucerne soil amendments have also been reported to suppress nematodes (Mankau, 1968; Mankau & Minteer, 1962; Johnson et al., 1967) and have shown potential to reduce plant disease caused by soil-borne fungi (Asirifi et al., 1994; Nam et al., 1988; Okumura, 2000). Sulphur volatiles produced by Allium species also show good potential for pest and disease pathogens (Auger et al., 2004). Amendments with high N contents are generally recognised as being more effective against nematodes than those with lower N contents (Mian & Rodriguez-Kabana, 1982).

Whilst biofumigation seems to be practical and involve little expenditure, efficacy is, in several cases, still far from that which can be obtained with the synthetic compound treatments (Lazzeri et al., 2004). There is no 'blanket' activity against all pests, diseases and weeds by any biofumigant. For example, research in France into the effectiveness of a range of green manures against *Aphanomyces* root rot of pea has not, so far, found a successful candidate (Moussart et al., 2004). Increased research and usage should highlight those crops which have consistently been effective against certain pests and diseases. However, whilst there has been some work on the identity of glucosinolates and their respective efficacy in Europe (Quinsac et al., 2004; Sørensen et al., 2004) the limited amount of research into the complicated reactions that occur in the soil during and after release of the isothiocyanates means that the effectiveness of many biocidal crops will remain difficult to predict.

The quality and quantity of glucosinolates present in cruciferous plants varies according to the genera, species, cultivar and their location in the plant, and thus gives different biofumigants different properties. This has given rise to blends being developed to provide maximum control for particular pest or disease situations. Trials have also been done to investigate the most glucosinolate-productive parts of these plants. For example, in-vitro work in Australia (Bianco et al., 2001) illustrated how root material from a mixture of Brassica napus and B. campestris was more effective against Rhizoctonia fragariae than the shoots from this mixture, suggesting that it might be worthwhile macerating the whole plant, not just the foliage, when incorporating biofumigants into the soil. The mixture used in this in-vitro study also produced 8 times more and a greater variety of isothiocyanates than the use of B. juncea, a popular biofumigant crop, alone. However, this literature study has illustrated that results from many in-vitro tests are contradicted when assessed in field situations. It has taken many years' research in Washington State, U.S.A., to develop a blend of Sinapis alba and Brassica juncea to control major pathogens of potato. namely Meloidogyne chitwoodi (Columbia root-knot nematode) and Verticillium dahliae (potato early dying disease) and weeds (McGuire, 2004a).

Whilst there is increasing knowledge about the characteristics of individual biofumigants there are common traits to be evaluated when considering their use. For example, a key characteristic of the release of the isothiocyanates (either from the enzyme hydrolysis in biofumigants or manufactured chemicals) is that it occurs within a few hours. Indeed, research on biofumigant crops has estimated that degradation may be faster (20 minutes) or slightly slower (7 hours) but the rate of degradation seems to be related to the level and particular types of glucosinolates involved (Aires et al., 2004). Persistence, or the length of time over which isothiocyanates are produced, might also be related to soil pH and moisture levels (Bianco et al., 2001) as well as temperature.

Other factors to be considered, which also apply when using manufactured sterilants, include the production of a fine tilth of soil, a minimal presence of clods, prompt and efficient incorporation and a quick, efficient surface seal. In The Netherlands, where the use of biocidal plants in strawberry, asparagus and woody ornamentals is under investigation, the soil surface is lightly compacted and irrigated after the biofumigant or green manure crop has been incorporated and then covered with a plastic film for 6-10 weeks under warm conditions in the summer. The anaerobic conditions that develop form additional toxic fermentation products but details of the success in controlling pathogens and weeds are not available. In the case of biocidal plants, other factors, such as ensuring a good biomass is produced, and that the plants are not only finely chopped but pulverised and watered before incorporation, have been found to be key elements in achieving maximum isothiocyanate concentration in the soil (Matthiessen, 2004).

An important point that is often overlooked in green manure or biofumigant studies is whether the cultivation of the biocidal crop itself will serve only to increase levels of pathogens, compared to leaving the land fallow. Certainly, Walker (2004), using pot experiments to investigate the control of root-knot nematodes in carrots, found that some green manure cultivars of sorghum and rapeseed he tested resulted in higher densities of *M. javanica* in soil before planting compared to leaving soil fallow. There also appeared to be differences in the effect of particular green manure crops on different populations of the same species of nematode, thus illustrating the importance

of selecting the right cultivar and recording the effects on nematodes at different sites. The susceptibility of some biofumigant cultivars to pest and disease infestation could be overcome by using the most promising candidates, such as lucerne, as a soil amendment in pellet form rather than as a green manure crop. Whilst this would increase costs, the reduced availability of other controls for pest and diseases could make their use a practical proposition. However, the loss of crop cover and biomass would reduce the influence of the crop for weed control and improving soil structure. Certainly the development and marketing of biocidal crops in pellet form is being considered by companies involved in their promotion. Thus whilst green manures may provide other benefits to the soil such as increased levels of organic matter, their use needs to be carefully considered.

Other negative effects of biocidal crops on subsequent cropping also need to be considered carefully. For example, a negative effect of the use of *Brassica napus* (or canola) on the growth of maize has sometimes been reported, and this has been attributed to an effect on vesicular-arbuscular mycorrhizal fungi, thus having a negative effect on uptake of low-mobile nutrients like phosphorous. Work by Pellerin *et al.*, (2004) however, failed to show that the biofumigant had a detrimental effect in this way. Recent research has shown different effects according to the biofumigant crop type on biocontrol agents such as *Trichoderma* spp., used against a range of plant diseases, so information regarding compatibility with such agents could be important (Galletti *et al.*, 2004). Walker (2004) found that carrot emergence was suppressed when amendments were applied 14 days before planting, but not if planting was delayed for at least four weeks.

Buried crop debris has been implicated as a contributory cause of fanging in carrots (Rubatzky *et al.*, 1999) but innovations such as the use of pellets, because of their small size, will be less likely to cause problems. As fanging is one of the symptoms used to measure nematode damage (though it can also be a symptom of disease and herbicide damage) care must be taken not to underestimate the effect of biocidal crops in reducing nematode levels.

Whilst many of the potential benefits have been highlighted and are indeed promoted in the commercial literature, there remains a lack of research into the efficacy and consequences of biofumigation. There are risks associated with the development of enhanced degradation (where the continual use of the same fumigant results in a shift in the soil micro-flora to populations that can break down the fumigant so rapidly that the fumigant is not available to destroy the target organisms) (Warton et al., 2001), potential impacts on associated beneficial organisms and the fate of isothiocyanates and other compounds in the environment (Kirkegaard & Matthiessen, 2004). In addition and in contrast to commercial pesticides, biocidal plant tissues also contain other chemicals and contribute large amounts of organic carbon that may positively or negatively influence the toxicity of isothiocyanates. As yet not enough is known about the complex relationships that occur when using biocidal plants to comment on likely efficacy (Morra, 2004).

Research and development of biofumigants in the field is in its infancy in the UK. As more becomes known about the chemical processes involved in the production, release and effectiveness of isothiocyanates, so it becomes essential to obtain specific information on glucosinolate types, levels and profiles in plant tissues of important cruciferous crops and the benefits of using particular cultivars, and even particular parts of these cultivars i.e. the seeds, roots, foliage, etc. The choice of biocidal plant may also depend on the relative importance of its biofumigant action as well as its benefits in biomass production; the latter will thus not only serve to benefit soil structure but may also increase isothiocyanate production.

Although it is unlikely that biofumigation or the addition of amendments will provide a direct replacement for manufactured sterilants or methyl bromide, its integration with other cultural or chemical methods offers an alternative to improve the sustainability of horticulture in general (Bianco *et al.*, 2001). In this survey of the literature it was interesting to note that some of those involved in organic farming did not view biofumigants as an essential tool, except for use in severe pest and disease outbreaks, such as during the first years of conversion from other farming methods (Micheloni & Conte, 2004). An alternative philosophy, however, might be that

frequent use of biofumigants might help to keep levels of pests and disease down, providing that this approach did not accelerate the biodegradation process.

Such is the confidence in the role that biofumigant crops can play as part of an integrated control programme for pathogen and weed control that there are several research centres developing cultivars specifically for this purpose, such as the Départ. Agronomie et Environnement, ENESAD, Dijon, France, which is working specifically on *B. juncea* genotypes (Merah *et al.*, 2004) and the *Brassicaceae* breeding group at the University of Idaho (Brown *et al.*, 2004), where breeding efforts have been directed towards two biofumigation systems, namely (1) green manure/incorporation cultivars that have high biomass accumulation and high concentrations of specific glucosinolates in the plant tissue, and (2) cultivars with high concentrations of specific glucosinolates in the seed meal, so that the meal can be used as a soil amendment. The latter might require less interruption in cropping schedules but possible phytotoxic effects of cropping close to incorporation need to be investigated.

One Italian company, Cerealtoscana (www.cerealtoscana.it), is promoting the use of biocidal plants under the brand 'Sovesci Bluformula'. It has worked with the Research Institute for Industrial Crops of (or Instituto Sperimentale Colture Industriali - ISCI) Bologna, to develop a selection of green manures, namely three cultivars of *Brassica* juncea (namely ISCI 20, 61 and 99) and one cultivar of Eruca sativa (NEMAT). in These available the UK via Plant Solutions Limited (www.plantsolutionsltd.com). The crops ISCI 99 and NEMAT have been selected for the joint VCS/CSL proposal to investigate the potential of such plants to reduce pests. diseases and weeds and improve soil nutrition.

The use of many different *Brassica* cultivars for biofumigation is cited in the literature because of their well-known properties of the production of glucosinolates (e.g. *Brassica napus* and *Brassica campestris* (Bianco *et al.*, 2001). However, the cultivars currently being offered commercially in the UK include *B. juncea* and *E. sativa* and it is the use of these that have been particularly investigated in this review.

Many of the papers referred herein have not been published in English, and in the time available it has not been possible to obtain complete translations but wherever possible English summaries have been obtained. In addition, there is much information available only as locally produced reports or Conference proceedings, which are also not immediately available to examine in detail.

### Brassica juncea (ISCI 99)

B. juncea is grown in several countries for oil production (e.g. Canada, China, India) and in Burgundy, France where it forms the basis for the famous 'Dijon' mustard used as a table condiment (Lionneton et al., 2004). B. juncea, sometimes known as 'Indian' or 'Brown' mustard, has been highlighted in several research projects as one cultivar having particularly high levels of glucosinolates. In particular it contains the glucosinolate sinigrin (which interestingly, is also responsible for the flavour of Dijon mustard), and work with this group has shown that organic matter and glucosinolate yield is highly dependent on plant type and cultivation time.

Researchers have investigated *B. juncea* but it is not always clear in the literature which cultivar has been used, thus some comments refer to the characteristics of the species in general, although it is clear that cultivars of the same species can have very different properties. *B. juncea* Czern et Coss, for example, has been shown to significantly suppress multiplication of the root-knot nematode *Meloidogyne incognita* on tomato roots and thus increase the crop yield (D'Addabbo *et al.*, 2004), but it would be unsafe to assume that the same effect would be achieved by all other cultivars of this species.

Work in Italy has included the use of the cultivar ISCI 20 (Lazzeri et al., 2003a). It is claimed to be a robust producer of glucosinolates, is adaptable to many soil types and climates and is easy to manage in the field. It is reported to produce up to 138 tonnes dry matter (DM) per ha, which may contain more than 1.6% Nitrogen. Reported variations in DM production are said to be due to differences in cultivation, including a failure to sow at the optimal time. Strawberry crops succeeding *B. juncea* in the rotation have shown no adverse reaction to glucosinolates and are said to have given results comparable those following sterilisation with methyl bromide, but no details are available. ISCI 20 is also being investigated by Applied Plant Research Flower Bulbs, Lisse, in The Netherlands as a control for plant-parasitic nematodes and soil borne fungal diseases (van Bruggen, 2004; van Os, 2004). Results for nematodes were not available, but the cultivar did produce a reduction in the incidence of *Rhizoctonia solani* on lily, resulting in a significant increase in bulb yield compared to other green manure crops. Variable results were obtained in similar work with tulip.

In France, trials have investigated an unknown cultivar of B. juncea amongst a range of potential biofumigants in the control of soil-borne pathogens in vegetable crops. Whilst, unfortunately, the specific effectiveness of B. juncea was not revealed, a general conclusion was that the biofumigants tested were effective against Rhizoctonia solani and R. solani, but not against Phytophthora cactorum (Villeneuve, et al., 2004). The activity against this disease is another example of how in-vitro studies seem to provide little indication as to the effectiveness of biofumigant crops in practice; laboratory work by Dunne et al (2003) suggested that B. juncea was very effective against P. cactorum. However, the laboratory work had demonstrated that there was also significant variation in the sensitivity of the Phytophthora species to the suppressive effects of the biofumigants. Experience gained during the trials work found that success was dependent on many factors, including the plant species used, the quantity of fresh organic matter ploughed in, the soil temperature during the period of coverage with plastic (with lower efficacy at lower temperatures) and the type of plastic used (important in their ability to reduce vapourisation rates, maintain temperature and modify the soil atmosphere).

There are several instances of conflicting information being published concerning the effectiveness of this group of green manure crops against pathogens. In laboratory work using pure isothiocyanates, it was predicted that B. juncea would be one of a group of Brassica plants containing high concentrations of propenyl isothiocyanates most likely to control Fusarium oxysporum isolates obtained from forest tree nurseries in Idaho and Washington (Smolinska et al., 2003). However, at a USDA Forest Service Nursery in Idaho, seedling production was not improved by incorporating brassica green manure crops, compared to the use of dazomet or fallowing, and in some cases large increases of potentially pathogenic Fusarium spp. were recorded (James et al., 2004). Soil Pythium levels were reduced when plastic tarpaulins were used to reduce losses of decomposition products, but this did not result in improved seedling production. The laboratory work had noted that only a fraction of the isothiocyanate potentially available from the glucosinolate within the tissues is actually released and available for pathogen inhibition; this combined with other factors that are necessary to maximise the fumigation effects illustrates the care and preparation necessary when investigating the use of biofumigants.

In Washington State, USA, *B. juncea* and *Sinapis alba* were incorporated into soils in the autumn and planted with potatoes the following spring. No information is available for the effects of *B. juncea* only, but the green manures were considered an effective replacement for manufactured soil fumigants (McGuire, 2004b).

Work in the USA has recently evaluated the herbicidal properties of seed meal of *Brassica* plants in glasshouse tests, including that produced from *B. juncea* under the name 'Pacific Gold'. The results varied according to cultivar, with 'Pacific Gold' having good herbicidal activity on wild oat seeds, but less activity compared to other Brassicaceae on wild mustard and pigweed. It did significantly reduce weed biomass overall. In subsequent field trials in strawberry crops 'Pacific Gold' did not perform as well as other seed meals, but in common with others was responsible for high phytotoxicity on first year strawberry transplants. However, yields from the crops were not significantly lower than those from the standard chemical treatment. Further research will determine efficacy rates and the timing of incorporation for maximum effect and productivity (Brown *et al.*, 2004). No phytotoxicity was reported when a crop of *B. juncea* was incorporated into soil to be cropped some 2-3 months before planting with strawberries (Lazzeri *et al.*, 2003b). In this trial weed control was not necessary, yields of strawberries were not compromised, but the effect on pests and diseases was unclear.

In Southern California, the incorporation of the formulation 'Pacific Gold' or the plant mulch was investigated for nematode, disease and weed control, but it is not clear whether the two types of amendment were used separately or jointly (Daugovish et al., 2004). A reduction of 92% in nematode numbers was achieved, but the growth of sclerotia of Sclerotinia minor (leaf drop of lettuce) was not affected except when used in combination with plastic covers, when a 75% reduction in sclerotial growth compared to the control was achieved. Such work highlights the requirement for a good seal to maximise the effect of the biofumigation. Colony development of Phytophthora cactorum (crown rot of strawberries) from biofumigated plots was inhibited by 90% or more, but intensive growth of Pythium spp. was also observed, leading to suggestions that the lack of growth of P. cactorum may not have been due

to biofurnigation but perhaps more likely a change in the microbiological environment that favoured the development of *Pythium* spp.

'Pacific Gold' seed meal resulted in 100% mortality of vine weevil larvae when incorporated into compost for potting, but resulted in phytotoxic effects on certain nursery tree species in the glasshouse or field. Also in the USA, glasshouse tests have been evaluating the effectiveness of *B. juncea* as chopped residues for weed control, but the results were very variable. Although in laboratory work germination of all weed species was completely inhibited, in the field trials there was no such inhibition in weed seed emergence. This again illustrates that whilst it is important to collect data from laboratory work where the effect of the biofumigants can be studied, it is also important to investigate their effects in the field to provide more lines of enquiry.

Some work has been done on the environmental effects of *B. juncea*. In Italy this has concentrated on investigating laboratory findings that glucosinolates and their hydrolysis products inhibit soil nitrifying bacteria communities, but in the field contradictory results have been obtained, requiring further work to determine the fertiliser value of this green manure crop (Marchetti *et al.*, 2004).

#### Eruca sativa cv. NEMAT

Compared to the work that has been done on *B. juncea*, relatively little information is available specifically for *E. sativa* cv. Nemat, perhaps because it has shown more selective action against pathogens in trials. The genus is part of the *Brassicaceae* and *E. sativa* is more commonly known as garden or salad Rocket, whilst the common name given to the one under investigation here is 'wild Rocket'. The main class of glucosinolates produced by this crop appears to be those containing glucoerucin, in contrast to the sinigrin found mainly in *Brassica* plants.

There is a lack of scientific data for the effect of this crop on nematodes. Hydrated, defatted seed meal (the product remaining after oil has been extracted) of *E. sativa* was tested for its efficacy in controlling *Sclerotinia* species (Marciano *et al.*, 2004). In common with other seed meals it greatly reduced the viability of sclerotia of *S. minor*, but did not affect those of *S. sclerotiorum*. It did have some effect on the antagonistic activity of biocontrol fungi, whereas other seed meals did not, so overall it did not rate highly in selections for disease control. Confirmation of this selective action raises an important point when choosing a biofumigant crop – if biofumigants have selective action then integrating them into control programmes will require specialist advice.

No phytotoxic symptoms were seen in strawberries planted after a crop of *E. sativa* had been incorporated into the soil prior to planting (Lazzeri *et al.*, 2003b). The overwintering crop cover so provided eliminated the need for weed control, and yields of strawberries were better than in untreated plots, but the effect on pests and diseases was unclear.

In Italy laboratory tests using a seed meal formulation of the product resulted in high mortality of wireworms, but insecticidal activity ceased after 2-3 days illustrating typical problems in persistence with brassicas that could cause practical problems (Furlan *et al.*, 2004).

Some work has been done on the fertilising effect of the defatted meal of this crop in soil. The meal is rich in organic nitrogen (4-7% N), phosphorous (2-3% P) and

sulphur (2-3% S). All these elements have to be mineralised to be available to plants. Meal was found to have potential as a good organic fertiliser (Cavani *et al.*, 2004).

#### **Conclusions**

With reform of the Common Agricultural Policy in the EU the basis upon which farm economics operates has been changed quite radically. Whilst the use of plants as whole crop biofumigants may become an economic option there is a need to ensure that truly repeatable results in terms of performance are shown by biofumigants (Askew, 2004).

The promotional literature produced by the commercial companies now marketing biocidal plants tends to present a simplistic picture of their use but evidence for consistent results with pest, disease or weed control is somewhat lacking. For instance, this relatively brief search and report on the available literature concerning *B. juncea* (ISCI 99) and *E. sativa* (Nemat) could obscure the fact that both crops may have a similar effect on levels of plant-parasitic nematodes in the soil. However, both companies have taken an active interest in research projects and in practical trials to gain further knowledge of the effectiveness of these crops in reducing pathogens and weeds. Their accumulated field experience must be utilised to the full when planning to use biocidal crops.

Future challenges for researchers are to identify ways of maximising the release of isothiocyanates from incorporated biofumigants into soil. This might include breeding for higher levels of glucosinolates, improved agronomic practices and understanding the interaction of biofumigation with the soil environment.

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## Results of pathogen analyses for soil samples from prospective trial sites

Field	Total Pythium cfu/g soil	Nematodes Root- lesion	Stubby- root	Needle	Heterodera cysts (juveniles)	Stunts	Spirals
Site E: Chestnuts Court	1,230	56	0	0	5(3)	2	0
Site E: Waterloo	1,485	90	5	0	5	0	0
Site K: Honey Pot 1	11,100	5	6	0	(13)	13	5
Site K: Top Battles	9,360	173	6	1	(1)	11	0

Nematode data Elveden Estates Waterloo

SL Reference Number	Plot number			Number per 200g
20508848	A11	8th June 2005	Root-Lesion	. 70
		_·	Stubby Root	11
			Cyst Male	1
20508849	A12	8th June 2005	Root-Lesion	42
			Stubby Root	5
		_	Stunt	1
20508850	A13	8th June 2005	Root-Lesion	85
			Stunt	5
20508851	A14	8th June 2005	Root-Lesion	55
			Stunt	14
			Stubby Root	1
			Cyst Juvenile	1
20508852	A21	8th June 2005	Root-Lesion	66
20300032	N21	out June 2005	Stubby Root	17
	<del>                                     </del>		Studdy Root	1
20508853	422	041. T 2005		
20300033	A22	8th June 2005	Root-Lesion	50
			Stunt	9
4050051	1.00		Stubby Root	11
20508854	A23	8th June 2005	Root-Lesion	18
<del> </del>	<u>.</u>		Stunt	4
· ·			Stubby Root	4
20508855	A24	8th June 2005	Root-Lesion	101
			Stubby Root	6
			Stunt	3
20508856	A31	8th June 2005	Root-Lesion	83
			Stubby Root	4
			Stunt	2
20508857	A32	8th June 2005	Root-Lesion	63
			Stunt	5
<del></del>			Stubby Root	2
20508858	A33	8th June 2005	Root-Lesion	68
			Stunt	10
			Cyst Male	1
20508859	A34	8th June 2005	Root-Lesion	86
	120.	041041102000	Stubby Root	12
20508860	A41	8th June 2005	Root-Lesion	78
	1111	Odi vane 2003	Stubby Root	2
<u>-</u> _	-		Stunt	2
20508861	A42	8th June 2005	Root-Lesion	146
20508862	A43			
20308802	A43	8th June 2005	Root-Lesion	96
20500060		0.1.7. 000.5	Stubby Root	10
20508863	A44	8th June 2005	Root-Lesion	54
			Stubby Root	34
20508864	A51	8th June 2005	Root-Lesion	90
			Stubby Root	11
	1 1 4 4	8th June 2005	Root-Lesion	77
20508865	A52	our June 2005		
20508865	A52	GH June 2005	Stubby Root	10
20508865 20508866	A52 A53	8th June 2005		<del></del>
			Stubby Root	10
			Stubby Root Root-Lesion	10 97
			Stubby Root Root-Lesion Stunt	10 97 10

			Stunt	1
20508868	A61	8th June 2005	Root-Lesion	87
			Stubby Root	8
- 11			Stunt	1
20508869	A62	8th June 2005	Root-Lesion	133
			Stunt	3
			Stubby Root	3
20508870	A63	8th June 2005	Root-Lesion	72
			Stubby Root	13
		"	Stunt	2
20508871	A64	8th June 2005	Root-Lesion	83
			Stubby Root	3
			Stunt	2

Nematode data Knights Top Battle

<b>CSL Reference Number</b>	Plot number	Sampling Stage (Date)	Common Name	Number per 200g
20508873	B11	8th June 2005	Root-lesion	91
			Stunts	11
<u></u>			Cyst Juvenile	i
20508874	B12	8th June 2005	Root-lesion	73
			Cyst Juvenile	18
<del> </del>			Stunts	10
			Stubby Root	1
20508875	B13	8th June 2005	Root-lesion	91
			Stunts	14
			Cyst Juvenile	8
•			Stubby Root	3
20508876	B14	8th June 2005	Root-lesion	187
		·	Cyst Juvenile	9
	<del></del>	<del></del>	Stubby Root	3
			Stunts	3
20508877	B21	8th June 2005	Root-lesion	73
			Stunts	2
20508878	B22	8th June 2005	Root-lesion	65
	i -		Stunts	2
····			Cyst Juvenile	1
20508879	B23	8th June 2005	Root-lesion	108
			Cyst Juvenile	5
			Stubby Root	3
			Stunts	2
20508880	B24	8th June 2005	Root-lesion	64
			Stunts	1
20508881	B31	8th June 2005	Root-lesion	69
, ···			Stunts	2
<u></u>			Cyst Juvenile	1
20508882	B32	8th June 2005	Root-lesion	103
			Stubby Root	2
<u></u> ,			Stunts	1
20508883	B33	8th June 2005	Root-lesion	44
			Stubby Root	1
20508884	B34	8th June 2005	Root-lesion	69
			Stunts	3
	1		Stubby Root	1
	ļ		Cyst Juvenile	1
20508885	B41	8th June 2005	Root-lesion	49
	Ì		Stunts	11
<del>-</del>	1		Stubby Root	1
	1	· · ·	Cyst Juvenile	1
20508886	B42	8th June 2005	Root-lesion	25
	1		Stunts	8
			Cyst Juvenile	1
20508887	B43	8th June 2005	Root-lesion	72
			Stunts	2
20508888	B44	8th June 2005	Root-lesion	98
			Stunts	2
	1	10	Cyst Juvenile	1
20508889	B51	8th June 2005	Root-lesion	24
20300003	1.22	Our June 2005	Stunts	9
20508890	B52	8th June 2005	Root-lesion	108

			Stunts	14
			Stubby Root	3
20508891	B53	8th June 2005	Root-lesion	139
			Stunts	7
20508892	B54	8th June 2005	Root-lesion	237
			Stunts	4
20508893	B61	8th June 2005	Root-lesion	93
20508894	B62	8th June 2005	Root-lesion	73
		,	Stunts	4
20508895	B63	8th June 2005	Root-lesion	176
			Stunts	5
20508896	B64	8th June 2005	Root-lesion	234
			Stunts	2

Nematode data Elveden Estates Waterloo

SL Reference Number		Sampling Stage (Date)		Number per 200g
20512965	A11	9th August 2005	Root-Lesion	129
			Stubby Root	34
20512966	A12	9th August 2005	Root-Lesion	153
·			Stunts	32
			Stubby Root	13
			Ring	_ 1
20512967	A13	9th August 2005	Root-Lesion	98
			Stunts	18
			Stubby Root	11
20512968	A14	9th August 2005	Root-Lesion	86
			Stunts	45
			Stubby Root	3
20512969	A21	9th August 2005	Root-Lesion	91
		•	Stubby Root	21
			Stunts	4
20512970	A22	9th August 2005	Root-Lesion	145
			Stunts	22
			Stubby Root	2
20512971	A23	9th August 2005	Root-Lesion	31
20312771	1125	Jui Hugust 2005	Stubby Root	3
20512972	A24	9th August 2005	Root-Lesion	111
20312772	NZ-T	Jui August 2005	Stubby Root	4
20512973	A31	9th August 2005	Root-Lesion	
20312973	AJI	9th August 2005		109
			Stubby Root	15
20512974	A32	041- 4	Stunts	2
20312974	A3Z	9th August 2005	Root-Lesion	39
			Stunts	5
20512075	4.00	0.1 4	Stubby Root	2
20512975	A33	9th August 2005	Root-Lesion	55
			Stunts	11
			Stubby Root	3
20512976	A34	9th August 2005	Root-Lesion	42
			Stubby Root	8
20512977	A41	9th August 2005	Root-Lesion	113
<u> </u>			Stunts	9
20512978	A42	9th August 2005	Root-Lesion	72
			Stubby Root	6
20512979	A43	9th August 2005	Root-Lesion	142
			Stubby Root	26
20512980	A44	9th August 2005	Root-Lesion	118
<u> </u>		-	Stubby Root	32
20512981	A51	9th August 2005	Root-Lesion	54
			Stubby Root	2
20512982	A52	9th August 2005	Root-Lesion	61
			Stubby Root	2
20512983	A53	9th August 2005	Root-Lesion	45
	<del>                                     </del>		Stubby Root	6
<del></del>	<del>                                     </del>		Stunts	8
20512984	A54	9th August 2005	Root-Lesion	60
2031270T	AJ4	7 iii August 2005	Stubby Root	2
20512985	A 6 1	Oth Assessed 2005	<del> </del>	
てのろまてみのつ	A61	9th August 2005	Root-Lesion	43

20512986	A62	9th August 2005	Root-Lesion	74
			Stunts	8
20512987	A63	9th August 2005	Root-Lesion	138
			Stubby Root	10
			Stunts	10
20512988	A64	9th August 2005	Root-Lesion	39

Nematode data Knights Top Battle

<b>CSL Reference Number</b>				Number per 200g
20512990	B11	9th August 2005	Root-lesion	136
<u> </u>			Cyst Juveniles	11
			Stunts	2
<u> </u>			Stubby Root	1
20512991	B12	9th August 2005	Root-lesion	111
		-	Cyst Juveniles	27
			Stunts	2
			Stubby Root	1
20512992	B13	9th August 2005	Root-lesion	92
	Ì		Stunts	18
			Cyst Juveniles	16
			Stubby Root	6
20512993	B14	9th August 2005	Root-lesion	143
			Stunts	8
			Cyst Juveniles	
20512994	B21	9th August 2005	Root-lesion	79
			Stunts	4
20512995	B22	9th August 2005	Root-lesion	35
			Cyst Juveniles	19
20512996	B23	9th August 2005	Root-lesion	81
			Cyst Juveniles	134
			Stunts	1
20512997	B24	9th August 2005	Root-lesion	63
			Cyst Juveniles	3
20512998	B31	9th August 2005	Root-lesion	18
200 12770	231	7 LI Tiagast 2000	Stunts	4
20512999	B32	9th August 2005	Root-lesion	60
20012777	232	Zurizuguer 2005	Stunts	10
20513000	B33	9th August 2005	Root-lesion	56
20212000		7 di 11dgabt 2005	Cyst Juveniles	4
•		•	Stunts	4
20513001	B34	9th August 2005	Root-lesion	109
20313001		7th Magast 2005	Cyst Juveniles	17
20513002	B41	9th August 2005	Root-lesion	54
20313002	D-1	7ti August 2005	Stunts	81
· · · · · · · · · · · · · · · · · · ·			Cyst Juveniles	- 1
20513003	B42	9th August 2005	Root-lesion	125
20313003	D72	7tii August 2005	Stunts	54
			Stubby Root	3
<del> </del>	1		Cyst Juveniles	1 .
20513004	B43	9th August 2005	Root-lesion	
ZVJ 13VU <del>1</del>	D43	Jul August 2003	Stunts	142
	<del> </del>			25
20512005	DAA.	0th A 2005	Cyst Juveniles	6
20513005	B44	9th August 2005	Root-lesion	138
20512007	Det	04- 4 12005	Stunts	4
20513006	B51	9th August 2005	Root-lesion	9
00510005	75.50	0.1 4	Stunts	6
20513007	B52	9th August 2005	Root-lesion	28
	<u> </u>		Stunts	3
20513008	B53	9th August 2005	Root-lesion	47
	ļ		Stunts	23
20513009	B54	9th August 2005	Root-lesion	69

			Stunts	4
20513010	B61	9th August 2005	Root-lesion	40
			Stunts	1
20513011	B62	9th August 2005	Root-lesion	31
			Stunts	2
·			Cyst Juveniles	1
20513012	B63	9th August 2005	Root-lesion	72
			Stunts	7
			Cyst Juveniles	3
20513013	B64	9th August 2005	Root-lesion	66
			Cyst Juveniles	2
		1		

Nematode data Elveden Estates Waterloo

CSL Reference Number	Plot number	Sampling Stage (Date)	Common Name	Number per 200g
20516409	A11	21st September 2005	Root-lesion	43
			Stubby Root	10
20516410	A12	21st September 2005	Root-lesion	71
			Stunt	12
20516411	A13	21st September 2005	Root-lesion	155
		2100 2000 1000 2000	Stunt	30
20516412	A14	21st September 2005	Root-lesion	41
	122	21st soptember 2003	Stubby Root	3
			Stunt	4
20516413	A21	21st September 2005	Root-lesion	106
20010110	7121	21st september 2003	Stunt	1
20516414	A22	21st September 2005	Root-lesion	99
20310414	ALL	21st September 2003	Stunt	<del></del>
		·		24
20516415	A23	21at Contamb == 2005	Stubby Root	2
20310413	AZ3	21st September 2005	Root-lesion	54
20516416	424	21-4 5-4 1 2005	Stunt	1
20516416	A24	21st September 2005	Root-lesion	79
20516417			Stunt	4
20516417	A31	21st September 2005	Root-lesion	44
20535410			Stunt	3
20516418	A32	21st September 2005	Root-lesion	82
			Stunt	1
20516419	A33	21st September 2005	Root-lesion	63
			Stunt	2
			<u>P</u> in	1
20516420	A34	21st September 2005	Root-lesion	28
			Stubby Root	1
20516421	A41	21st September 2005	Root-lesion	65
			Stubby Root	4
20516422	A42	21st September 2005	Root-lesion	103
			Stunt	3
20516423	A43	21st September 2005	Root-lesion	80
			Stubby Root	9
20516424	A44	21st September 2005	Root-lesion	76
			Stubby Root	8
20516425	A51	21st September 2005	Root-lesion	70
			Stubby Root	2
20516426	A52	21st September 2005	Root-lesion	84
			Stubby Root	9
20516427	A53	21st September 2005	Root-lesion	58
			Stubby Root	4
20516428	A54	21st September 2005	Root-lesion	16
			Stubby Root	10
			Stunt	1
20516429	A61	21st September 2005	Root-lesion	74
20210127		215t Coptomoct 2003	Stubby Root	8
20516430	A62	21st September 2005	Root-lesion	
20310730	AUZ	21st September 2003		40
			Stubby Root	2
20516431	A 62	21at Contamba 2005	Stunt	2
ZU310431	A63	21st September 2005	Root-lesion	20
00516400	1.71	01.0 . 1 000	Stunt	2
20516432	A64	21st September 2005	Root-lesion	34
<u> </u>		·	Stunt	4

Nematode data	Knights Top Battle

CSL Reference Number	Plot number	Sampling Stage (Date)	Common Name	Number per 200g
20516434	B11	21st September 2005	Root-lesion	54
			Stunt	4
			Cyst Juvenile	1
20516435	B12	21st September 2005	Root-lesion	35
			Stunt	4
			Cyst Juvenile	1
20516436	B13	21st September 2005	Root-lesion	62
			Stunt	40
20516437	B14	21st September 2005	Root-lesion	58
20516438	B21	21st September 2005	Root-lesion	75
20516439	B22	21st September 2005	Root-lesion	28
20516440	B23	21st September 2005	Root-lesion	107
<del></del>		1 1	Cyst Juvenile	14
		1	Stunt	5
20516441	B24	21st September 2005	Root-lesion	108
<del></del>			Stubby Root	1
			Cyst Juvenile	2
20516442	B31	21st September 2005	Root-lesion	28
		2 ist sopionisti 2000	Stubby Root	1
20516443	B32	21st September 2005	Root-lesion	47
	252	21st September 2000	Cyst Juvenile	3
			Stunt	3
20516444	B33	21st September 2005	Root-lesion	68
203,10444		21st September 2003	Cyst Juvenile	27
		· ·	Stunt	2
20516445	B34	21st September 2005	Root-lesion	85
20310443	D3+	21st September 2005	Stubby Root	2
20516446	B41	21st September 2005	Stunt	52
20310440	D41	21st September 2005	Root-lesion	34
			Stubby Root	4
20516447	B42	21st September 2005	Stunt	<del></del>
20310447	D42	21st September 2003	Root-lesion	50
· · · · · · · · · · · · · · · · · · ·		-	Stubby Root	39
20516448	B43	21st September 2005	Root-lesion	3
20310448	D43 ·	21st September 2003		51
	-		Stunt Cyst Juvenile	45
20516449	B44	21st September 2005	Root-lesion	
20310449	D <del>11</del>	21st September 2003		32
			Stunt	5
			Stubby Root	4
20516450	D.51	21st Contact to 2005	Cyst Juvenile	11
20516450	B51	21st September 2005	Root-lesion	46
			Stubby Root	5
<del></del>			Stunt	5
00516451	250	01.0	Cyst Juvenile	2
20516451	B52	21st September 2005	Root-lesion	53
		<u> </u>	Stunt	16
			Stubby Root	1
20516452	B53	21st September 2005	Root-lesion	37
			Stunt	22
			Cyst Juvenile	15
17	_		Stubby Root	1
20516453	B54	21st September 2005	Root-lesion	184
	L			

CSL Reference Number	Plot number	Sampling Stage (Date)	Common Name	Number per 200g
20516454	B61	21st September 2005	Root-lesion	. 37
			Stunt	4
			Stubby Root	1
20516455	B62	21st September 2005	Root-lesion	-59
	_		Cyst Juvenile	12
			Stunt	8
			Stubby Root	2
20516456	B63	21st September 2005	Root-lesion	36
			Stubby Root	7
			Stunt	5
			Cyst Juvenile	1
20516457	B64	21st September 2005	Root-lesion	61
			Stubby Root	
			Cyst Juvenile	1

# Statistical Analysis for Effect of Biocidal Plants on Numbers of Nematodes and *Pythia*

Statistical analysis was by Analysis of Variance (anova) with contrasts, using Genstat 8. There were 3 treatments (Biocide, Cultivation, No cultivation) with 4 replicates of each treatment at each of 2 sites (Elveden, Knights). There were 2 complete experiments conducted at the same time, one with Mustard 99 as the Biocide and one with Mustard 119\*. In the analysis the sites were blocks and the two contrasts were Biocide versus Cultivation and Biocide versus No cultivation. The pythium and nematode counts were transformed to logarithms for the analyses (log(n+1) for stunt and stubby nematodes since these counts contained zeroes), which successfully normalised the data and stabilised the variances, thus meeting the requirements of anova.

At pre-drilling (Count 1) no statistically significant differences were found between the three treatments (biocide, cultivation and no cultivation) for either total pythium, total nematodes, lesion nematodes, stubby nematodes or stunt nematodes, for either Mustard 99 or Mustard 119 (see Tables 1a and 1b).

At pre-incorporation (Count 2) there was a significant difference between treatments for total nematodes and lesion nematodes, due to there being more nematodes in the Mustard 99 and Mustard 119 plots than in the respective Cultivated plots and also, in the case of Mustard 119, in the Uncultivated plots. There were also more stubby nematodes in the Mustard 99 plots, but this difference was not detected with Mustard 119.

At six weeks post-incorporation (Count 3) no differences were detected - all the differences seen at Count 2 had disappeared.

Changes in nematode numbers between Counts 1 and 3 were compared between the treatments (referred to as Diff 1:3 in the Tables) to see if they changed similarly. The values analysed were log(Count1) - log(Count3) which is equivalent to Count1/Count3. No significant differences were detected for pythium or any of the nematodes for either biocides (Tables 1a,b).

Changes in numbers between Counts 2 and 3 were similarly compared between treatments (referred to as Diff 2:3, Table 1c.). Overall, there was a marked drop in total nematode numbers from Count 2 to Count 3 with both Biocides which was not mirrored in the other two treatments. Analyses of the two sites individually confirmed this effect for Knights but not for Elveden. For a simple visualisation see Table 3e for t-tests on the combined sites (don't quote these since the previous approach is better).

The results of the analyses are sumarised in appended Table 1 (log means and probabilities), Table 2 (antilogs of the means and their confidence intervals), and Table 3 (Means and standard errors for each treatment at each site, log and antilog.). Slightly edited (shortened) Genstat output is appended – note cross-reference numbering between the contents of Tables and the output.

Table 1a. Means (log10) and probabilities of differences between treatments for total Pythium, total nematodes and lesion nematodes.

	Mustard 99	Means (logs)	Mustard		e.s.e	Anova	Contrast Must 99	Contrast Must 99
	Total	Cult	99	NoCult	df=20	Treats	vCult	vNoCult
	pythium					р	р	р
1.1.1	Count 1	3.366	3.554	3.521	0.065	0.120	0.055	0.721
1.1.2	Count 2	3.414	3.578	3.565	0.053	0.073	0.039 *	0.861
1.1.3	Count 3	3.485	3.625	3.549	0.052	0.186	0.071	0.310
1.1.4	Diff 1:3	0.119	0.071	0.028	0.081	0.729	0.677	0.710

	Mustard 119	Means (logs)			e.s.e	Anova	Contrast	Contrast
	<b>T</b> ( )			Mustard		<b>-</b> .	Must 119	Must 119
<u> </u>	Total	Cult	NoCult	119	df=20	Treats	vCult	vNoCult
	pythium					р	р	р
1.2.1	Count 1	3.268	3.310	3.296	0.068	0.905	0.770	0.889
1.2.2	Count 2	3.391	3.385	3.446	0.043	0.552	0.378	0.326
1.2.3	Count 3	3.443	3.468	3.531	0.037	0.235	0.102	0.238
1.2.4	Diff 1:3	0.175	0.159	0.235	0.068	0.712	0.541	0.439

	Mustard 99	Means (logs)	Mustard			Anova		Contrast Must 99		Contrast Must 99	
	Total	Cult	99	NoCult	df=20	Treats		vCult		vNoCult	į
	nematodes					р		р		р	
2.1.1	Count 1	1.886	1.974	1.844	0.083	0.300		0.302		0.132	
2.1.2	Count 2	1.799	2.172	1.961	0.076	0.009	**	0.003	**	0.066	
2.1.3	Count 3	1.748	1.842	1.907	0.077	0.365		0.401		0.561	
2.1.4	Diff 1:3	-0.138	-0.132	0.063	0.096	0.265		0.966		0.166	
	Lesion					•					
	nematodes										
2.1.5	Count 1	1.852	1.899	1.787	0.064	0.471		0.611		0.227	
2.1.6	Count 2	1.726	2.065	1.851	0.074	0.014	*	0.004	**	0.054	
2.1.7	Count 3	1.711	1.765	1.879	0.071	0.254		0.594		0.268	
2.1.8	Diff 1:3	-0.142	-0.134	0.093	0.089	0.130		0.950		0.085	

	Mustard 119	Means (logs)	ñ	Mustard		Anova	Contrast Must 119		Contrast Must 119	
	Total	Cult	NoCult	119		Treats	vCult		vNoCult	
	nematodes					р	р		р	•
2.2.1	Count 1	2.062	2.009	1.906	0.074	0.340	0.153		0.338	
2.2.2	Count 2	1.783	1.686	2.146	0.071	<0.001 *	** 0.002	**	<0.001	***
2.2.3	Count 3	1.688	1.850	1.909	0.066	0.072	0.028		0.533	
2.2.4	Diff 1:3	-0.373	-0.159	0.003	0.083	0.016 *	0.005	**	0.184	
	Lesion									
	nematodes									!
2.2.5	Count 1	2.036	1.960	1.838	0.086	0.296	0.121		0.331	
2.2.6	Count 2	1.750	1.608	2.032	0.078	0.003 *	* 0.019	•	<0.001	***
2.2.7	Count 3	1.623	1.745	1.743	0.083	0.509	0.321		0.990	
2.2.8	Diff 1:3	-0.413	-0.215	-0.095	0.093	0.074	0.025	*	0.373	

Table 1b. Means (log10) and probabilities of differences between treatments for stubby and stunt nematodes.

	Mustard 99	Means (logs)	Mustard		e.s.e	Anova	Contrast Must 99		Contrast Must 99	
	Stubby	Cult	99	NoCult	df=20	Treats.	vCult		vNoCult	
	nematodes					р	р		р	
3.1.1	Count 1	0.421	0.458	0.463	0.129	0.969	0.842		0.979	
3.1.2	Count 2	0.405	0.727	0.390	0.100	0.045 *	0.034		0.027	*
3.1.3	Count 3	0.195	0.205	0.097	0.102	0.714	0.940		0.461	
3.1.4	Diff 1:3	-0.227	-0.253	-0.366	0.159	0.807	0.909		0.620	
	Stunt									
	nematodes									
3.1.5	Count 1	0.460	0.769	0.542	0.128	0.232	0.102		0.223	
3.1.6	Count 2	0.600	0.960	0.380	0.254	0.099	0.172		0.035	*
3.1.7	Count 3	0.248	0.789	0.435	0.167	0.090	0.032	*	0.148	
3.1.8	Diff 1:3	-0.212	0.020	-0.107	0.151	0.561	0.289		0.557	

	Mustard 119	Means (logs)		Mustard		Anova	Contrast Must 119	Contrast Must 119	
	Stubby	Cult	NoCult	119	df=20	Treats	vCult	vNoCult	
	nernatodes		7.1			р	р	р	İ
3.2.1	Count 1	0.413	0.462	0.420	0.119	0.953	0.965	0.810	
3.2.2	Count 2	0.218	0.285	0.550	0.187	0.197	0.091	0.171	
3.2.3	Count 3	0.449	0.575	0.582	0.131	0.725	0.481	0.970	ŀ
3.2.4	Diff 1:3	0.036	0.113	0.161	0.140	0.816	0.532	0.810	
	Stunt								ļ
	nematodes								}
3.2.5	Count 1	0.476	0.640	0.433	0.117	0.435	0.796	0.226	l I
3.2.6	Count 2	0.520	0.560	0.850	0.180	0.393	0.214	0.275	
3.2.7	Count 3	0.511	0.459	0.809	0.161	0.275	0.205	0.140	ĺ
3.2.8	Diff 1:3	0.034	-0.181	0.376	0.153	0.055	0.130	0.018	*

Table 1c. Means (log10) and probabilities of differences between log nematode pre-incorporation (Count 2) and six weeks after incorporation (Count 3).

	Mustard 99		iffs (logs) Mustard		e.s.e	Anova		Contrast Must 99		Contrast Must 99	
Diff 2:3		Cult	99	NoCult	df=20	Treats		vCult		vNoCult	
	Both sites Total					Р		p		р	
4.1.1	nems.	-0.051	-0.329	-0.055	0.0795	0.034	*	0.022	•	0.024	*
4.1.2	Lesion	-0.016	-0.300	0.029	0.075	0.011	*	0.014	*	0.006	**
4.1.3	Stunt	-0.349	-0.167	0.052	0.1629	0.243		0.438		0.354	
4.1.4	Stubby	-0.210	-0.522	-0.293	0.1417	0.295		0.135		0.266	
	Elveden Total				df=9						
4.2.1	nems.	-0.107	-0.288	-0.025	0.1308	0.387		0.355		0.189	
4.2.2	Lesion	-0.047	-0.233	-0.002	0.1355	0.47		0.357		0.257	
4.2.3	Stunt	-0.24	-0.29	0.16	0.225	0.347		0.878		0.192	
4.2.4	Stubby	-0.73	-0.68	-0.66	0.241	0.976		0.882		0.952	
	Knights Total				df≕9						
4.3.1	nems.	0.005	-0.371	0.084	0.0986	0.058		0.024	*	0.07	
4.3.2	Lesion	0.016	-0.366	0.059	0.0725	0.005	**	0.005	**	0.002	**
4.3.3	Stunt	-0.46	-0.04	-0.06	0.244	0.426		0.26		0.975	
4.3.4	Stubby	0.314	-0.362	0.075	0.1282	0.014	*	0.005	**	0.039	*

	Mustard			10	-					
	119	Mean	diffs (logs)		e.s.e	Anova		Contrast	Contrast	
D:# 0.3		0	NI-OII	Mustar	46-00	Treatm	l	Must 119	Must 119	
Diff 2:3	<b>.</b>	Cult	NoCult	d 119	df=20	ents		<ul><li>vCult</li></ul>	vNoCult	
	Both sites					р		р	р	
	Total	0.005	0.404	0.007	0.4040	0.005		0.000	0.040	*
4.4.1	nems.	-0.095	0.164		0.1019	0.035	•	0.088	0.012	
4.4.2	Lesion	-0.127	0.137	-0.289	0.118	0.057		0.129	0.019	*
4.4.3	Stunt	-0.01	-0.1	-0.04	0.174	0.928		0.71	0.795	
4.4.4	Stubby	0.231	0.29	0.032	0.1673	0.531		0.806	0.288	
	Elveden				df=9				97	
	Total									
4.5.1	nems.	-0.224	-0.015	-0.161	0.1624	0.661		0.387	0.543	
4.5.2	Lesion	-0.23	-0.05	-0.13	0.174	0.772		0.485	0.758	
4.5.3	Stunt	-0.09	-0.16	-0.1	0.299	0.981		0.858	0.883	
4.5.4	Stubby	-0.08	0.24	-0.29	0.295	0.483		0.473	0.243	
	Knights				df=9					
	Total									
4.6.1	nems.	0.034	0.343	-0.313	0.1078	0.006	**	0.074	0.002	**
4.6.2	Lesion	-0.022	0.327	-0.447	0.1304	0.008	**	0.091	0.002	**
4.6.3	Stunt	0.07	-0.04	0.03	0.21	0.937		0.727	0.832	
4.6.4	Stubby	0.54	0.345	0.349	0.1663	0.654		0.43	0.985	

Table 2a. Antilogs of the means and 95% confidence intervals for total Pythium, total nematodes and lesion nematodes.

Total p	ythium								
	Mustard 99			Cult		ļ	<b>NoCult</b>		
		95%	95%		95%	95%		95%	95%
Count	Mean	low lim	high lim	Mean	low lim	high lim	Mean	low lim	high lim
1	3581.0	2615.6	4902.5	2322.7	1696.6	3180.0	3318.9	2424.3	4543.8
2	3784.4	2938.1	4874.5	2594.2	2014.0	3341.4	3672.8	2851.5	4730.8
3	4217.0	3286.5	5410.8	3054.9	2380.9	3919.8	3540.0	2758.9	4542.1
	Mustard 119			Cult			NoCult		
		95%	95%		95%	95%		95%	95%
Count	Mean	low lim	high lim	Mean	low lim	high lim	Mean	low lim	high lim
1	1977.0	1428.2	2736.7	1853.5	1339.0	2565.8	2041.7	1475.0	2826.3
2	2792.5	2273.6	3429.9	2460.4	2003.2	3021.9	2426.6	1975.7	2980.4
3	3396.3	2850.1	4047.0	2773.3	2327.4	3304.7	2937.6	2465.3	3500.6

									.14
Total n	ematodes						-		
	Mustard 99			Cult			NoCult		
		95%	95%		95%	95%		95%	95%
Count	Mean	low lim	high lim	Mean	low lim	high lim	Mean	low lim	high lim
1	94.2	63.3	140.3	76.9	51.7	114.5	69.8	46.9	104.0
2	148.6	103.0	214.5	63.0	43.6	90.9	91.4	63.3	131.9
3	69.5	47.9	100.7	56.0	38.6	81.1	80.7	55.7	117.0
				10					
Lesion	nematodes		1						
	Mustard 99			Cult			NoCult		
		95%	95%		95%	95%		95%	95%
Count	Mean.	low lim	high lim	Mean	low lim	high lim	Mean	low lim	high lim
1	79.3	58.4	107.6	71.1	52.4	96.6	61.2	45.1	83.2
2	116.1	81.5	165.6	53.2	37.3	75.8	71.0	49.8	101.1
3	58.2	41.4	81.9	51.4	36.5	72.3	75.7	53.8	106.5

Total n	ematodes						<u>=</u>		
	Mustard 119			Cult			NoCult		
		95%	95%		95%	95%		95%	95%
Count	Mean	low lim	high lim	Mean	low lim	high lim	Mean	low lim	high lim
1	80.5	56.3	115.1	115.3	80.7	164.9	102.1	71.4	145.9
2	140.0	99.4	197.0	60.7	43.1	85.4	48.5	34.5	68.3
3	81.1	59.1	111.2	48.8	35.5	66.9	70.8	51.6	97.1
Lesion	nematodes								
	Mustard 119			Cult			NoCult		,
		95%	95%		95%	95%		95%	95%
Count	Mean	low lim	high lim	Mean	low lim	hig <b>h l</b> im	Mean	low lim	high lim
1	68.9	45.5	104.1	108.6	71.8	164.3	91.2	60.3	137.9
2	107.6	74.0	156.5	56.2	38.7	81.8	40.6	27.9	59.0
3	55.3	37.1	82.6	42.0	28.1	62.7	55.6	37.2	83.0

Table 2b. Antilogs of the means and 95% confidence intervals for stubby and stunt nematodes.

Stubby	nematodes								
	Mustard 99		1	Cult			<b>NoCult</b>		
		95%	95%		95%	95%		95%	95%
Count	Mean	low lim	high lim	Mean	low lim	high lim	Mean	low lim	high lìm
1	2.9	1.5	5.3	2.6	1.4	4.9	2.9	1.6	5.4
. 2	5.3	3.3	8.6	2.5	1.6	4.1	2.5	1.5	4.0
3	1.6	1.0	2.6	1.6	1.0	2.6	1.3	8.0	2.0
Stunt n	ematodes								
	Mustard 99			Cult			NoCult		
		95%	95%		95%	95%		95%	95%
Count	Mean	low lim	high lim	Mean	low lim	high lim	Mean	low lim	high lim
1	5.9	3.2	10.9	2.9	1.6	5.3	3.5	1.9	6.4
2	9.1	2.7	30.9	4.0	1.2	13.5	2.4	0.7	8.1
3	6.2	2.8	13.7	1.8	0.8	3.9	2.7	1.2	6.1
					•				
Stubby	nematodes			-					<u> </u>
	Mustard 119			Cult			NoCult		
		95%	95%		95%	95%		95%	95%
Count	Mean		high lim	Mean	low lim	_	Mean		high lim
1	2.6	1.5	4.7	2.6	1.5	4.6	2.9	1.6	5.1
2	3.5	1.4	8.7	1.7	0.7	4.1	1.9	8.0	4.7
3	3.8	2.0	7.2	2.8	1.5	5.3	3.8	2.0	7.0
Stunt n	ematodes								
	Mustard 119	ı		Cult			NoCult		
		95%	95%		95%	95%		95%	95%
Count	Mean	low lim	high lim	Mean	low lim	high lim	Mean	low lim	high lim
1	2.7	1.5	4.8	3.0	1.7	5.3	4.4	2.5	7.7
2	7.1	3.0	16.8	3.3	1.4	7.9	3.6	1.5	8.6
3	6.4	3.0	14.0	3.2	1.5	7.0	2.9	1.3	6.2

Table 3a. Mustard 99 trial: Means and standard errors of the log number of nematodes for each treatment at each site.

Total py	thium								
i Otai py	unum	Mustard	Mustard	Mustard				1	
		991	992	993	Cult1 Cult2 Cult3	NoCult1	NaCulta	NoCult2	
Elveden	Mean		3.355	3.405	3.141 3.243 3.226	3.230	3.478	3.277	
LIVEGEII		1							
	SE	0.049	0.076	0.066	0.093 0.039 0.043	0.101	0.039	0.091	
Knights	Mean	3.737	3.801	3.845	3.591 3.584 3.744	3.812	3.651	3.820	
rangino	SE	0.141	0.109	0.070	0.043 0.043 0.103		0.091	0.071	
Total	OL	0.141	0.100	0.070	0.040 0.040 0.100	0.101	0.031	0.071	
nemato	des								
		Mustard	Mustard	Mustard					
		991	992	993	Cult1 Cult2 Cult3	NoCult1	NoCult2	NoCult3	
Elveden	Mean	1.850	2.185	1.898	1.921 1.825 1.718	1.790	1.971	1.946	
	SE	0.060	0.044	0.134	0.032 0.099 0.099	0.136	0.152	0.078	
								0.0.0	
Knights	Mean	2.098	2.158	1.786	1.851 1.774 1.778	1.898	1.951	1.867	
_	SE	0.070	0.014	0.084	0.076 0.158 0.120	0.059	0.133	0.148	
		'							
Lesion							17		
nematod	des			14					
		Mustard	Mustard	Mustard					
		991	992	993	Cult1 Cult2 Cult3	NoCult1	NoCult2	NoCult3	
Elveden	Mean	1.785	2.055	1.822	1.871 1.748 1.701	1.695	1.914	1.913	
	SE	0.066	0.057	0.134	0.033 0.102 0.101	0.159	0.147	0.066	
					*5				
Knights	Mean	2.013	2.074	1.708	1.833 1.705 1.720	1.879	1.787	1.846	
	SE	0.089	0.044	0.056	0.075 0.163 0.105	0.053	0.085	0.138	
Stubby									
nemato	des	ı				1			
		Mustard	Mustard	Mustard					
		991	992	993	Cult1 Cult2 Cult3				
Elveden		ł .	1.093	0.411	0.573 0.809 0.075		0.780	0.119	
	SE	0.241	0.193	0.254	0.232 0.166 0.075	0.197	0.193	0.119	
Knights	Mean	0.376	0.362	0.000	0.270 0.000 0.314	0.151	0.000	0.075	
	SE	0.144	0.176	0.000	0.099 0.000 0.113	0.151	0.000	0.075	
_									
Stunt	J								
nemato	zes	Muntand	N. de continued	Microsoph				1	
		Mustard 991	Mustard 992	Mustard 993	Cult1 Cult2 Cult3	NoCult1	NoCule	NoCult3	
Elveden	Mean	ļ	1.115	0.826	0.574 0.584 0.345		0.515	0.675	
		•							
	·SE	0.259	0.380	0.319	0.223 0.230 0.131	0.144	0.327	0.259	
بالمارين المارين	N. a. a. a.	0.075	0.707	0.750	0.045.0.040.0.454	0.400	0.0		
Knights	Mean	ł	0.797	0.753	0.345 0.610 0.151		0.250	0.195	
	SE	0.127	0.196	0.331_	0.131 0.219 0.151	0.044	0.166	0.195	

Means and standard errors of log values

Total pythium, total nematodes and lesion nematodes are log10(value) since there were no zeroes Stubby and stunt nematodes are log10(value+1) since there were zeroes

Table 3b. Mustard 119 trial: Means and standard errors of the log number of nematodes for each treatment at each site.

				0.10.						
Total pythium										
	Mustard	Mustard	Mustard			300			1	ľ
	1191	1192	1193	Cult1	Cult2	Cult3	NoCult1	NoCult2	NoCult3	
Elveden Mean			3							
	3.143	3.302	3.458		3.150		3.164	3.218	3.385	
SE	0.102	0.030	0.061	0.174	0.084	0.035	0.085	0.036	0.064	
,										
Knights Mean	2 440	2 500	3.604	2 5 4 0	2 622	3 500	2 456	2 552	2.550	
_	3.449	3.589	l l		3.632		3.456	3.552	3.552	
SE	0.042	0.077	0.073	0.015	0.045	0.034	0.061	0.062	0.027	ľ
Total										
nematodes										
	Mustard	Mustard	Mustard			[				ñ
	1191	1192	1193	Cult1	Cult2	Cult3	NoCult1	NoCult2	NaCulta	
F1 1 14										
Elveden Mean	2.012	2.095	1.934	2.002	1.844	1.620	1.977	1.778	1.762	
SE	0.056	0.073	0.038	0.048	0.137	0.117	0.027	0.012	0.116	
			1			ļ				
Knights Mean	1.799	2.197	1.883	2 424	1.723	1 757	2.040	1.594	1.937	
	1004		<b>I</b>							
SE	0.099	0.032	0.087	0.116	0.090	0.063	0.184	0.163	0.112	
Lesion										
nematodes										
	Mustard	Mustard	Mustard							1
	1191	1192	1193	Cult1	Cideo	Culta	NoCult1	NoCult2	NaCulta	
	5-04									
Elveden Mean	1.943	2.034	1.902			1.576	1.937	1.737	1.684	
SE	0.090	0.063	0.042	0.057	0.126	0.117	0.022	0.030	0.163	
Knights Mean	1.734	2.030	1.584	2 112	1 603	1.670	1.983	1.478	1.805	
_			<b>I</b>							
SE	0.128	0.100 =	0.045	0.118	0.088	0.062	0.213	0.192	0.157	
Stubby										
nematodes										
	Mustard	Mustard	Mustard							
	1191	1192	1193	Cult1	Cult2	Cult3	NoCult1	NoCult2	NoCult3	
Elveden Mean	0.766	0.949	0.663		0.435		0.773	0.569		
	2.3		1						0.804	
SE	0.336	0.350	0.231	0.135	0.261	0.228	0.163	0.092	_ 0.133	
Knights Mean	0.075	0.151	0.500	0.000	0.000	0.540	0.151	0.000	0.345	
	540									
SE	0.075	0.151	0.168	0.000	0.000	0.128	0.150	0.000	0.161	
Stunt					187					
nematodes										
	Mustard	Mustard	Mustard							
	1191	1192	1193	Cult1	Cult2	Cult3	NoCult1	NoCult2	NoCult3	
Elveden Mean	0.119	0.250	0.151			0.413				
	532						-	0.239	0.075	
SE	0.119	0.250	0.151	0.062	0.289	0.147	0.246	0.239	0.075	
Knights Mean	0.747	1.442	1.468	በ ፈጻባ	0.540	0.608	0.945	0.882	0.843	
•	7.5									
\$E	0.158	0.268	0.230	U.1/5	U.128	0.210	0.100	0.174	0.307	

Means and standard errors of log values

Total pythium, total nematodes and lesion nematodes are log10(value) since there were no zeroes Stubby and stunt nematodes are log10(value+1) since there were zeroes

Table 3c. Mustard 99 trial: Antilogs of the Mean log number of nematodes for each treatment at each site, and of the Mean±SE.

Total pythium										
	Mustard 991	Mustard 992	Mustard 993	Cult1	Cult2	Cult3	NoCult1	NoCult2	NoCult3	
Elveden Mean	2349.6	2264.6	2541.0	1383.6	1749.8			3006.1	1892.3	
Mean+SE	2630.3	2697.7	2958.0	1714.0	1914.3	1857.8	2142.9	3288.5	2333.5	
Mean-SE	2098.9	1901.1	2182.7	1116.9	1599.6	1524.1	1345.9	2747.9	1534.6	
Knights Mean	5457.6	6324.1	6998.4	3899.4	3837.1	5546.3	6486.3	4477.1	6606.9	
Mean+SE	7550.9	8128.3	8222.4	4305.3	4236.4	7030.7	8184.6	5520.8	7780.4	
Mean-SE	3944.6	4920.4	5956.6	3531.8	3475.4	4375.2	5140.4	3630.8	5610.5	
Total nematodes						•			,	
	Mustard 991	Mustard 992	Mustard 993	Cult1	Cult2	Cult3	NoCult1	NoCult2	NoCult3	
Elveden Mean	70.8	153.1	79.1	83.4	66.8	52.2	61.7	93.5	88.3	
Mean+SE	81.3	169.4	107.6	89.7	83.9	65.6	84.3	132,7	105.7	
Mean-SE	61.7	138.4	58.1	77.4	53.2	41.6	45.1	65.9	73.8	
Knights Mean	125.3	143.9	61.1	71.0	59.4	60.0	79.1	89.3	73.6	
Mean+SE	147.2	148.6	74.1	84.5	85.5	79.1	90.6	121.3	103.5	
Mean-SE	106.7	139.3	50.4	59.6	41.3	45.5	69.0	65.8	52.4	
Lesion nematodes						,				
	Mustard 991	Mustard 992	Mustard 993	Cult1	Cult2	Cult3	NoCult1	NoCult2	NoCult3	
Elveden Mean	61.0	113.5	66.4	74.3	56.0	50.2	49.5	82.0	81.8	
Mean+SE	71.0	129.4	90.4	80.2	70.8	63.4	71.4	115.1	95.3	
Mean-SE	52.4	99.5	48.8	68.9	44.3	39.8	34.4	58.5	70.3	
Knights Mean	103.0	118.6	51.1	68.1	50.7	52.5	75.7	61.2	70.1	
Mean+SE	126.5	131.2	58.1	80.9	73.8	66.8	85.5	74.5	96.4	
Mean-SE	83.9	107.2	44.9	57.3	34.8	41.2	67.0	50.4	51.1	
Stubby nematodes										
,	Mustard 991	Mustard 992	Mustard 993	Cult1	Cult2	Cult3	NoCult1	NoCult2	NoCult3	
Elveden Mean	2.5	11.4	1.6	2.7	5.4	0.2	5.0	5.0	0.3	
Mean+SE	5.0	18.3	3.6	5.4	8.4	0.4	8.4	8.4	0.7	
Mean-SE	1.0	6.9	0.4	1.2	3.4	0.0	2.8	2.9	0.0	
Knights Mean	1.4	1.3	0.0	0.9	0.0	1.1	0.4	0.0	0.2	
Mean+SE	2.3	2.4	0.0	1.3	0.0	1.7	1.0	0.0	0.4	
Mean-SE	0.7	0.5	0.0	0.5	0.0	0.6	0.0	0.0	0.0	
Stunt nematodes				•						
	Mustard 991	Mustard 992	Mustard 993	Cult1	Cult2	Cult3	NoCult1	NoCult2	NoCult3	
Elveden Mean	2.7	12.0	5.7	2.8	2.8	1.2	3.5	2.3	3.7	
Mean+SE	5.7	30.3	13.0	5.3	5.5	2.0	5.2	6.0	7.6	
Mean-SE	1.0	4.4	2.2	1.2	1.3	0.6	2.2	0.5	1.6	
Knights Mean	8.4	5.3	4.7	1.2	3.1	0.4	1.7	0.8	0.6	
Mean+SE	11.7	8.8	11.1	2.0	5.7	1.0	2.0	1.6	1.4	
Mean-SE	6.0	3.0	1.6	0.6	1.5	0.0	1.4	0.2	0.0	

## Antilogs:

10^value for log10(value).

(10^value)-1 for log10(value+1) Note that this is an approximation

Table 3d. Mustard 119 trial: Antilogs of the Mean log number of nematodes for each treatment at each site, and of the Mean±SE.

Total pyt	hium									
• •		Mustard 1	191 Mustard 1	1192 Mustard 1	193 Cult1	Cult2	Cult3	NoCult1	NoCult2	NoCult
Elveden	Mean	1390.0	2004.	5 2870.8	970.5	1412.	5 2023.0			
	Mean+SE	1757.	2147.8	8 3303.7	1448.	B 1714.	02192.8			
	Mean-SE	1099.	1870.	7 2494.6	650.1	1164.	1 1866.4	1199.5	1520.5	
Knights	Mean	2811.	3881.	5 4017.9	3531.	8 <b>428</b> 5.	5 3801.9	2857.6	3564.5	
_	Mean+SE	3097.4	4 4634.	5 4753.4	3655.	9 4753.	44111.5	3288.5	4111.5	3793.1
	Mean-SE	2552.	7 3250.	9 3396.3	3411.	9 3863.	7 3515.6	2483.1	3090.3	3349.7
Total ner	natodes						-			
		Mustard '	191 Mustard	1192 Mustard 1	193 Cult1	Cult2	Cult3	NoCult1	NoCult2	NoCult
Elveden	Mean	102.8	124.5	85.9	100.5	69.8	41.7	94.8	60.0	57.8
	Mean+SE	116.9	147.2	93.8	112.2	95.7	54.6	100.9	61.7	75.5
	Mean-SE	90.4	105.2	2 78.7	89.9	50.9	31.8	89.1	58.3	44.3
Knights	Mean	63.0	157.4	76.4	132.1	52.8	57.1	109.6	39.3	86.5
	Mean+SE	79.1	169.4	93.3	172.6	65.0	66.1	167.5	57.1	111.9
	Mean-SE	50.1	146.2	2 62.5	101.2	43.0	49.4	71.8	27.0	66.8
Lesion n	ematodes				1					
		111.20111111111111111111111111111111111		1192 Mustard 1	7.5				NoCult2	
Elveden	Mean	87.7	108,1		91.2			86.5	54.6	48.3
	Mean+SE	1			104.0		49.3	91.0	58.5	70.3
	Mean-SE	71.3	93.5		80.0		28.8	82.2	50.9	33.2
Knights	Mean	54.2	107.2		129.4			96.2	30.1	63.8
	Mean+SE	72.8	134.9		169.8			157.0	46.8	91.6
	Mean-SE	40.4	85.1	34.6	98.6	40.3	40.6	58.9	19.3	44.5
Stubby n	ematodes	F 200 1000			بر مامه،		1	المالية الما		
				1192 Mustard 1	ı				NoCult2	
Elveden	Mean	4.8	7.9	3.6	5.7	1.7	1.3	4.9	2.7	5.4
	Mean+SE		18.9		8.1	4.0	2.9	7.6	3.6	7.7
16 1 14	Mean-SE	1.7	3.0	1.7	3.9	0.5	0.3	3.1	2.0	3.7
Knights	Mean	0.2	0.4	2.2	0.0	0.0	2.5	0.4	0.0	1.2
	Mean+SE	1	1.0	3.7	0.0	0.0	3.7	1.0	0.0	2.2
	Mean-SE	0.0	0.0	1.1	0.0	0.0	1.6	0.0	0.0	0.5
Stunt ne	matodes	Westersalls	140434	4400 Minstead 4	سرم امما		040	ير مايدا		
Characters.		1		1192 Mustard 1	ı			1	NoCult2	
Elveden		0.3	0.8	0.4	1.9	2.2	1.6	1.2	0.7	0.2
	Mean+SE	1	2.2	1.0	2.4	5.1.	2.6	2.8	2.0	0.4
Main-ta	Mean-SE	0.0	0.0	0.0	1.5	0.6	0.8	0.2	0.0	0.0
Knights	Mean	4.6	26.7		2.1	2.5	3.1	7.8	6.6	6.0
	Mean+SE	1	50.3		3.6	3.7	5.6	10.1	10.4	13.1
	Mean-SE	2.9	13.9	16.3	1.1	1.6	1.5	6.0	4.1	2.4

## Antilogs:

10^value for log10(value).

(10^value)-1 for log10(value+1) Note that this is an approximation

Table 3e. T-tests on the differences between counts 1 and 3 and between 2 and 3 for log Total nematodes, (sites lumped together).

Trial	Diff	Treat	N	Mean difference	S.E.	t value	df	р	
Mustard 99	1:3	NoCult	8	0.06281	0.08507	0.74	7	0.484	
	1:3	Cult	8	-0.1378	0.1017	-1.36	7	0.217	
	1:3	Mustard 99	8	-0.1319	0.1306	-1.27	7	0.2	
Mustard 99	2:3	NoCult	8	-0.05454	0.06646	-0.82	7	0.439	
	2:3	Cult	8	-0.05112	0.08151	-0.63	7	0.550	
	2:3	Mustard 99	8	-0.3295	0.08377	-3.93	7	0.006	**
Mustard 119	1:3	NoCult	8	-0.1592	0.08157	-1.95	7	0.092	
	1:3	Cult	8	-0.3734	0.08358	-4.47	7	0.003	**
	1:3	Mustard 99	8	0.00312	0.08496	0.04	7	0.972	
Mustard 119	2:3	NoCult	8	0.1639	0.1025	1.60	7	0.154	
	2:3	Cult	8	-0.09483	0.1327	-0.71	7	0.498	
	2:3	Mustard 99	8	-0.2370	0.06428	-3.96	7	0.008	**
		_							

## **Edited Genstat output**

## 1. Analysis of Pythium results

Analysis of Variance with contrasts, Genstat 8.

Three treatments: Biocide, Cultivation, No cultivation.

Four replicates each block. Two blocks: Elveden, Knights

Contrasts: Biocide v Cultivation, Biocide v No cultivation

### 1.1. Mustard 99

## 1.1.1. Total pythium count 1 (transformed to log10).

ANOVA [PRINT=aovtable,information,means,contrast; FACT=32; FPROB=yes; PSE=diff,lsd,means; LSDLEVEL=5] logTotPyth1

Analysis of variance					
******					
Variate: logTotPyth1					
Source of variation	d.f	s.s.	m.s.	v.r.	F pr
Site stratum	1	1.30458	1.30458	38.10	
Site.*Units* stratum		•			
Treat	2	0.16162	0.08081	2.36	0.120 ris
Must99vCult	1	0.14195	0.14195	4.14	0.055 ns
Must99vNocult	1	0.00448	0.00448	0.13	0.721 ns
Residual	20	0.68491	0.03425		
Total	23	2.15111			

#### Tables of contrasts

\*\*\*\*\*\*\*\*\*\*\*\*\*

Variate: logTotPyth1

Must99vCult 0.188, s.e. 0.0925, ss.div. 4.00 Must99vNocult 0.033, s.e. 0.0925, ss.div. 4.00

## Tables of means

Variate: logTotPyth1 Grand mean 3.480

Treat Cult Mustard99 NoCult 3.366 3.554 3.521

## Standard errors of means

Table	Treat
rep.	8
d.f.	20
e.s.e.	0.0654

## Standard errors of differences of means

Table	Treat	
rep.	8	
d.f.	20	

s.e.d. 0.0925

Least significant differences of means (5% level) \_\_\_\_\_\_

Treat rep. 8 d.f. 20

l.s.d. 0.1930

## 1.1.2. Total pythium count 2 (transformed to log10).

145 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff,lsd,means; LSDLEVEL=5] logTotPyth2

Analysis of variance -----

Variate: logTotPyth2

d.f. s.s. m.s. v.r. F pr. 1 0.61648 0.61648 27.75 Source of variation Site stratum

Site.\*Units\* stratum 2 0.13316 0.06658 3.00 0.073 ns 1 0.10784 0.10784 4.85 0.039 \* 1 0.00070 0.00070 0.03 0.861 ns 20 0.44429 0.02221 Treat Must99vCult Must99vNocult

Residual

Total 23 1.19394

Tables of contrasts

------Variate: logTotPyth2

Must99vCult 0.164, s.e. 0.0745, ss.div. 4.00 Must99vNocult 0.013, s.e. 0.0745, ss.div. 4.00

Tables of means -----

Variate: logTotPyth2 Grand mean 3.519

> Treat Cult Mustard99 NoCult 3.414 3.578 3.565

Standard errors of means

Table Treat 8 rep. d.f. 0.0527

Standard errors of differences of means

Treat

. 8 rep. 20 d.f. 0.0745 s.e.d.

Least significant differences of means (5% level)

Table Treat 8 rep. 20 d.f. 0.1555 l.s.d.

## 1.1.3. Total pythium count 3 (transformed to log10).

ANOVA [PRINT=acvtable,information,means,contrast; FACT=32; FPROB=yes; PSE=diff,lsd,means; LSDLEVEL=5] logTotPyth3

Tables of contrasts

Variate: logTotPyth3

Must99vCult 0.140, s.e. 0.0734, ss.div. 4.00 Must99vNocult 0.076, s.e. 0.0734, ss.div. 4.00

Tables of means

Variate: logTotPyth3 Grand mean 3.553

Treat Cult Mustard99 NoCult 3.485 3.625 3.549

Standard errors of means

-----

 Table
 Treat

 rep.
 8

 d.f.
 20

 e.s.e.
 0.0519

Standard errors of differences of means

Table Treat rep. 8 d.f. 20

Least significant differences of means (5% level)

0.0734

Table Treat rep. 8 d.f. 20 l.s.d. 0.1531

## 1.1.4. Difference between Total pythium counts 1 and 3 (log minus log).

157 ANOVA [PRINT=aovtable,information,means,contrast; FACT=32; FPROB=yes; PSE=diff,lsd,means; LSDLEVEL=5] DifflogPyth13

Analysis of variance

Variate: DifflogPyth13

Source of variation	d.f.	S.S.	m.s.	v.r.	F pr.
Site stratum	1	0.00708	0.00708	0.14	
Site.*Units* stratum					
Treat	2	0.03327	0.01664	0.32	0.729 ns
Must99vCult	1	0.00929	0.00929	0.18	0.677 ns
Must99vNocult	1	0.00739	0.00739	0.14	0.710 ns
Residual	20	1.03738 '	0.05187		
Total	23	1.07773			

#### Tables of contrasts

-----

Variate: DifflogPyth13

Must99vCult -0.05, s.e. 0.114, ss.div. 4.00 Must99vNocult 0.04, s.e. 0.114, ss.div. 4.00

#### Tables of means

----

Variate: DifflogPyth13

Grand mean 0.072

Treat Cult Mustard99 NoCult 0.119 0.071 0.028

#### Standard errors of means

Table Treat

rep. 8 d.f. 20 e.s.e. 0.0805

#### Standard errors of differences of means

Table Treat

rep. 8 d.f. 20 s.e.d. 0.1139

#### Least significant differences of means (5% level)

Table Treat rep. 8 d.f. 20 l.s.d. 0.2375

## 1.2. Mustard 119

## 1.2.1. Total pythium count 1 (transformed to log10).

132 ANOVA [PRINT=aovtable,information,means,contrast; FACT=32; FPROB=yes; PSE=diff,lsd,means; LSDLEVEL=5] logTotPyth1

#### Analysis of variance

Variate: logTotPyth1 Source of variation	d.f		m.s.	×	Fnx
Source of Variation		s.s.		v.r.	F pr
Site stratum	1	0.89625	0.89625	24.47	
Site.*Units* stratum					
Treat	2	0.00733	0.00366	0.10	0.905 ns
Must119 vCult	1	0.00322	0.00322	0.09	0.770 ns
Must119vNocult	1	0.00073	0.00073	0.02	0.889 ns
Residual	20	0.73265	0.03663		
Total	23	1.63622			

## Tables of contrasts

~============

Variate: logTotPyth1

Mustl19vCult 0.028, s.e. 0.0957, ss.div. 4.00 Mustl19Nocult -0.014, s.e. 0.0957, ss.div. 4.00

#### Tables of means

-----

Variate: logTotPythl Grand mean 3.291

Treat Cult NoCult Mustard119 3.268 3.310 3.296 Cult

#### Standard errors of means

Table Treat rep. 8

d.f. 20 e.s.e. 0.0677

#### Standard errors of differences of means

Table Treat

rep. 8 d.f. 20 0.0957 s.e.d.

#### Least significant differences of means (5% level)

\_\_\_\_\_\_\_\_\_\_\_

Table Treat rep. 8 d.f. 20 0.1996 l.s.d.

## 1.2.2. Total pythium count 2 (transformed to log10).

114 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff,lsd,means; LSDLEVEL=5] logTotPyth2

#### Analysis of variance -----------

Variate: logTotPvth2

 Source of variation
 d.f.
 s.s.
 m.s.
 v.r.

 Site stratum
 1
 0.81153
 0.81153
 55.43

 v.r. F pr. Site.\*Units\* stratum 2 0.01793 0.00896 0.61 0.552 ns 1 0.01188 0.01188 0.81 0.378 ns 1 0.01485 0.01485 1.01 0.326 ns 0.61 0.552 ns Must119vCult Must119vNocult 0.01464 20 0.29282 Residual 23 1.12228

#### Tables of contrasts

Total

\_\_\_\_\_\_\_

Variate: logTotPyth2

Mustl19vCult 0.055, s.e. 0.0605, ss.div. 4.00 Mustl19vNocult 0.061, s.e. 0.0605, ss.div. 4.00

#### Tables of means

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Variate: logTotPyth2 Grand mean 3.407

NoCult Mustard119 3.385 3.446 Treat Cult

3.391

#### Standard errors of means

\_\_\_\_\_\_

Table rep. d.f. 20 e.s.e. 0.0428

#### Standard errors of differences of means

Table Treat rep. 8 d.f. 20 s.e.d. 0.0605

Least significant differences of means (5% level)

Table Treat rep. 8 d.f 20 l.s.d. 0.1262

## 1.2.3. Total pythium count 3 (transformed to log10).

120 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff,lsd,means; LSDLEVEL=5] logTotPyth3

#### Analysis of variance

\*\*\*\*\*\*\*\*\*\*\*\*\*\*

Variate: logTotPyth3					
Source of variation	d.f.	S.S	m.s.	v.r.	F pr
Site stratum	1	0.22972	0.22972	21.60	
Site.*Units* stratum					
Treat	2	0.03315	0.01657	1.56	0.235 ns
Must119vCult	1	0.03133	0.03133	2.95	0.102 ns
Mustl19vNocult	1	0.01574	0.01574	1.48	0.238 ns
Residual	20	0.21273	0.01064		
Total	23	0.47560			

#### Tables of contrasts ----

Variate: logTotPyth3

Must119vCult 0.088, s.e. 0.0516, ss.div. 4.00 Must119vNocult 0.063, s.e. 0.0516, ss.div. 4.00

#### Tables of means

-----

Variate: logTotPyth3 Grand mean 3.481

NoCult Mustard119 Treat Cult 3.443 3.468 3.531

## Standard errors of means

\_\_\_\_\_\_

Table d.f. 20 e.s.e. 0.0365

## Standard errors of differences of means

Treat rep. 20 8 d.f.

s.e.d.

Least significant differences of means (5% level) 

0.0516

Table Treat rep. 8 d.f. 20 0.1076 l.s.d.

## 1.2.4. Difference between Total pythium counts 1 and 3 (log minus log).

126 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes: PSE=diff, lsd, means: LSDLEVEL=5 DifflogPyth13

#### Analysis of variance

Variate: DifflogPyth13

 
 Source of variation
 d.f.
 s.s.
 m.s.
 v.r.
 F pr.

 Site stratum
 1
 0.21847
 0.21847
 5.85
 Site.\*Units\* stratum 2 0.02584 0.01292 0.35 0.712 ns 1 0.01445 0.01445 0.39 0.541 ns 1 0.02326 0.02326 0.62 0.439 ns 20 0.74697 0.03735 23 0.99128 Treat Must119vCult Mustl19vNocult Residual Total

## Tables of contrasts

\_\_\_\_\_\_

Variate: DifflogPyth13

Must119vCult 0.060, s.e. 0.0966, ss.div. 4.00 Must119vNocult 0.076, s.e. 0.0966, ss.div. 4.00

Tables of means -----

Variate: DifflogPyth13

Grand mean 0.189

Cult NoCult Mustard119 0.175 0.159 0.235

### Standard errors of means

Treat Table rep. 8 d.f 20

e.s.e. 0.0683

### Standard errors of differences of means

Treat rep. 8

20 d.f. 0.0966

Least significant differences of means (5% level)

Table Treat rep.
d.f.
l.s.d. 8 · 20 0.2016

84

## 2. Analysis of Nematode results

Analysis of Variance with contrasts, Genstat 8.

Three treatments: Biocide, Cultivation, No cultivation.

Four replicates each block.
Two blocks: Elveden, Knights

Contrasts: Biocide v Cultivation, Biocide v No cultivation

### 2.1. Mustard 99

## 2.1.1. Total nematode count 1 (transformed to log10).

261 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff, lsd, means; LSDLEVEL=5] logTotNem1

## Analysis of variance

Variate: logTotNem1					
Source of variation	d.f.	s.s.	m.s.	v.r.	F pr
Site stratum	1	0.05472	0.05472	1.99	_
Site.*Units* stratum					
Treat	2	0.07048	0.03524	1.28	0.300 ns
Must99vCult	1	0.03092	0.03092	1.12	0.302 ns
Must99vNoCult	1	0.06768	0.06768	2.46	0.132 ns
Residual	20	0.55015	0.02751		
Total	23	0.67534			

# Tables of contrasts

Variate: logTotNem1

Must99vCult 0.088, s.e. 0.0829, ss.div. 4.00 Must99vNoCult 0.130, s.e. 0.0829, ss.div. 4.00

# Tables of means

Variate: logTotNem1 Grand mean 1.901

Treat Cult Mustard99 NoCult 1.886 1.974 1.844

## Standard errors of means

 Table
 Treat

 rep.
 8

 d.f.
 20

 e.s.e.
 0.0586

## Standard errors of differences of means

 Table
 Treat

 rep.
 8

 d.f.
 20

 s.e.d.
 0.0829

## Least significant differences of means (5% level)

Treat rep. 8 d.f. 20 0.1730

## 2.1.2. Total nematode count 2 (transformed to log10).

267 ANOVA [PRINT=aovtable,information,means,contrast; FACT=32; FPROB=yes; PSE=diff,lsd,means; LSDLEVEL=5] logTotNem2

#### Analysis of variance \*\*\*\*\*\*\*\*\*\*\*\*\*\*\*\*

Variate: logTotNem2

Source of variation d.f. s.s. m.s. v.r. F pr. Site stratum 1 0.00658 0.00658 0.14 Site.\*Units\* stratum 2 0.55723 Treat 0.27861 5.97 0.009 \*\* 1 0.55410 0.55410 11.87 0.003 \*\* 1 0.17693 0.17693 3.79 0.066 ns 20 0.93343 0.04667 Must99vCult Must99vNoCult Residual

23 1.49724 Total

#### Tables of contrasts ------

Variate: logTotNem2

Must99vCult 0.37, s.e. 0.108, ss.div. 4.00 Must99vNoCult 0.21, s.e. 0.108, ss.div. 4.00

#### Tables of means ------

Variate: logTotNem2

Grand mean 1.977

Treat Cult Mustard99 NoCult 1.799 2.172 1.961

#### Standard errors of means

Table Treat rep. d.f. 20 e.s.e. 0.0764

## Standard errors of differences of means

Table Treat 8 rep. 20 d.f. s.e.d. 0.1080

### Least significant differences of means (5% level)

Table Treat 8 20 rep. d.f 0.2253

## 2.1.3. Total nematode count 3 (transformed to log10).

273 ANOVA [PRINT=aovtable,information,means,contrast; FACT=32; FPROB=yes; PSE=diff, 1sd, means; LSDLEVEL=5] logTotNem3

Analysis of variance \_\_\_\_\_

Variate: logTotNem3

d.f. s.s. m.s. 1 0.01148 0.01148 Source of variation v.r. Site stratum 0.24 Site.\*Units\* stratum L 0.10156 0.05078 1.06 0.365 ns 1 0.03521 0.03521 0.74 0.401 ns 1 0.01670 0.01670 0.35 0.561 ns 20 0.95700 0.04785 23 1.07003 Treat Must99vCult Must99vNoCult Residual

20 0.95700 Total 23 1.07003

Tables of contrasts

----------Variate: logTotNem3

Must99vCult 0.09, s.e. 0.109, ss.div. 4.00 Must99vNoCult -0.06, s.e. 0.109, ss.div. 4.00

Tables of means -----

Variate: logTotNem3 Grand mean 1.832

> Treat Cult Mustard99 NoCult 1.748 1.842 1.907

Standard errors of means

- 8 d.f 20 0.0773

Standard errors of differences of means

Table Treat rep. 8 d.f. 20 0.1094 s.e.d.

Least significant differences of means (5% level)

------

Table Treat 8 rep. d.f. 20 l.s.d. 0.2281

### 2.1.4. Difference between Total nematode counts 1 and 3.

279 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff,lsd,means; LSDLEVEL=5] DifflogTotNem13

Analysis of variance 

Variate: DifflogTotNem13

Source of variation d.f. s.s. m.s. Site stratum 1 0.11632 0.11632 v.r. F pr Site.\*Units\* stratum

Treat 2 0.20849 0.10424 1.42 0.265 ns
 Must99vCult 1 0.00014 0.000 0.966 ns
 Must99vNoCult 1 0.15163 0.15163 2.06 0.166 ns
Residual 20 1.46876 0.07344
Total 23 1.79356

Tables of contrasts

\_\_\_\_

Variate: DifflogTotNem13

Must99vCult 0.01, s.e. 0.135, ss.div. 4.00 Must99vNoCult -0.19, s.e. 0.135, ss.div. 4.00

Tables of means

e.s.e.

Variate: DifflogTotNem13

Grand mean -0.069

Treat Cult Mustard99 NoCult -0.138 -0.132 0.063

Standard errors of means

Table Treat rep. 8 d.f. 20

Standard errors of differences of means

\_\_\_\_\_\_

 Table
 Treat

 rep.
 8

 d.f.
 20

 s.e.d.
 0.1355

Least significant differences of means (5% level)

Table Treat rep. 8 d.f. 20 1.s.d. 0.2826

## 2.1.5. Lesion nematode count 1 (transformed to log10).

286 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff, lsd, means; LSDLEVEL=5] logLesion1

Analysis of variance

\*\*\*\*\*\*\*\*\*\*\*\*\*\*\*\* Variate: logLesion1 v.r. F pr. d.f. s.s. m.s. 1 0.09393 0.09393 Source of variation m.s. Site stratum 2.89 2 Site.\*Units\* stratum 2 0.05083 0.02541 0.78 0.471 ns 1 0.00865 0.00865 0.27 0.611 ns 1 0.05034 0.05034 1.55 0.227 ns Treat Must99vCult 1 Must99vNoCult Residual 20 0.64.913 0.03246 Total 23 0.79388

Tables of contrasts
----Variate: logLesion1

Must99vCult 0.046, s.e. 0.0901, ss.div. 4.00 Must99vNoCult 0.112, s.e. 0.0901, ss.div. 4.00

Tables of means

Variate: logLesion1 Grand mean 1.846

Treat Cult Mustard99 NoCult 1.852 1.899 1.787

Standard errors of means

Table Treat rep. 8 d.f. 20 e.s.e. 0.0637

Standard errors of differences of means

------

Table Treat rep. 8 d.f. 20 s.e.d. 0.0901

Least significant differences of means (5% level)

Table Treat
rep. 8
d.f. 20
l.s.d. 0.1879

### 2.1.6. Lesion nematode count 2 (transformed to log10).

292 ANOVA [PRINT=aovtable,information,means,contrast; FACT=32; FPROB=yes; PSE=diff,lsd,means; LSDLEVEL=5] logLesion2

Analysis of variance

Variate: logLesion2

Source of variation d.f. s.s. m.s. v.r. F pr. Site stratum 1 0.01518 0.01518 0.35 0.35 Site.\*Units\* stratum Treat 2 0.46899 0.23449 5.38 0.014 \* 1 0.45828 0.45828 10.51 0.004 \*\* Must99vCult Must99vNoCult 4.20 0.054 1 0.18326 0.18326 0.04362 20 0.87238 Residual 1.35655 23 Total

Tables of contrasts

Variate: logLesion2

Must99vCult 0.34, s.e. 0.104, ss.div. 4.00 Must99vNoCult 0.21, s.e. 0.104, ss.div. 4.00

Tables of means

Variate: logLesion2 Grand mean 1.881

Treat Cult Mustard99 NoCult 1.726 2.065 1.851

#### Standard errors of means

Table Treat rep. d.f. 20 e.s.e. 0.0738

#### Standard errors of differences of means

Table Treat rep. 8 d.f. 20 0.1044 s.e.d.

Least significant differences of means (5% level)

\_\_\_\_\_

Table Treat rep. - 8 20 d.f. 0.2178 l.s.d.

## 2.1.7. Lesion nematode count 3 (transformed to log10).

298 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff,lsd,means; LSDLEVEL=5] logLesion3

#### Analysis of variance

------

Variate: logLesion3

variate: loguesions					
Source of variation	d.f	s.s?	m.s.	v.r.	F pr.
Site stratum	1	0.01728	0.01728	0.43	
Site.*Units* stratum					
Treat	2	0.11890	0.05945	1.47	0.254 ns
Must99vCult	1	0.01184	0.01184	0.29	0.594 ns
Must99vNoCult	1	0.05243	0.05243	1.30	0.268 ns
Residual	20	0.80868	0.04043		
Total	23	0.94486			

#### Tables of contrasts

Variate: logLesion3

Must99vCult 0.05, s.e. 0.101, ss.div. 4.00 Must99vNoCult -0.11, s.e. 0.101, ss.div. 4.00

#### Tables of means

Variate: logLesion3 Grand mean 1.785

Treat Cult Mustard99 NoCult 1.711 1.765 1.879

#### Standard errors of means

Table Treat rep. . 8 d.f. 20 0.0711

Standard errors of differences of means.

 Table
 Treat

 rep.
 8

 d.f.
 20

 s.e.d.
 0.1005

Least significant differences of means (5% level)

Table Treat
rep. 8
d.f. 20
l.s.d. 0.2097

#### 2.1.8. Difference between Lesion nematode counts 1 and 3.

304 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff, lsd, means; LSDLEVEL=5] DifflogLesion13

Analysis of variance

Variate: DifflogLesion13

Source of variation d.f. s.s. m.s. v.r. F pr. Site stratum 1 0.19180 0.19180 3.06 Site.\*Units\* stratum

Treat 2 0.28389 0.14194 2.27 0.130 ns Must99vCult 1 0.00025 0.00025 0.00 0.950 ns Must99vNoCult 1 0.20550 0.20550 3.28 0.085 ns Residual 20 1.25182 0.06259 Total 23 1.72750

Tables of contrasts

Variate: DifflogLesion13

Must99vCult 0.01, s.e. 0.125, ss.div. 4.00 Must99vNoCult -0.23, s.e. 0.125, ss.div. 4.00

Tables of means

Variate: DifflogLesion13

Grand mean -0.061

Treat Cult Mustard99 NoCult -0.142 -0.134 0.093

Standard errors of means

Table

 Table
 Treat

 rep.
 8

 d.f.
 20

 e.s.e.
 0.0885

Standard errors of differences of means

\_\_\_\_\_

 Table
 Treat

 rep.
 8

 d.f.
 20

 s.e.d.
 0.1251

Least significant differences of means (5% level)

Table Treat rep. 8

d.f. 20 1.s.d. 0.2609

#### 2.2. Mustard 119

## 2.2.1. Total nematode count 1 (transformed to log10).

262 ANOVA [PRINT=aovtable,information,means,contrast; FACT=32; FPROB=yes; PSE=diff,lsd,means; LSDLEVEL=5] logTotNem1

## 

# Tables of contrasts

Variate: logTotNem1

Must119vCult -0.16, s.e. 0.105, ss.div. 4.00 Must119vNoCult -0.10, s.e. 0.105, ss.div. 4.00

## Tables of means

Variate: logTotNem1 Grand mean 1.992

Treat Cult NoCult Mustard119 2.062 2.009 1.906

#### Standard errors of means

Table Treat rep. 8 d.f. 20 e.s.e. 0.0744

## Standard errors of differences of means

Table Treat rep. 8 d.f. 20 s.e.d. 0.1053

## Least significant differences of means (5% level)

 Table
 Treat

 rep.
 8

 d.f.
 20

 l.s.d.
 0.2196

### 2.2.2. Total nematode count 2 (transformed to log10).

268 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff,lsd,means; LSDLEVEL=5] logTotNem2

Analysis of variance

-----Variate: logTotNem2

d.f. s.s. m.s. v.r. F pr. 1 0.02757 0.02757 0.68 Source of variation Site stratum Site.\*Units\* stratum 2 0.93985 0.46993 11.59 <.001 \*\*\*
1 0.52561 0.52561 12.97 0.002 \*\*
1 0.84619 0.84619 20.88 <.001 \*\*\*
20 0.81057 0.04053 Mustll9vCult Must119vNoCult Residual

23 1.77800 Total

Tables of contrasts

Variate: logTotNem2

Mustl19vCult 0.36, s.e. 0.101, ss.div. 4.00 Mustl19vNoCult 0.46, s.e. 0.101, ss.div. 4.00

Tables of means ~=========

Variate: logTotNem2

Grand mean 1.872 Treat Cult NoCult Mustard119 1.783 1.686 2.146

Standard errors of means

-----

Table rep. 8 d.f. 20 e.s.e. 0.0712

Standard errors of differences of means

\_\_\_\_\_\_\_

Treat rep. 8 d.f. 20 0.1007

Least significant differences of means (5% level)

Table Treat rep. 8 d.f 20 l.s.d. 0.2100

## 2.2.3. Total nematode count 3 (transformed to log10).

274 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff,lsd,means; LSDLEVEL=5] logTotNem3

Analysis of variance -----

Variate: logTotNem3

Source of variation d.f. s.s. m.s. v.r. F pr.

Site stratum	1	0.04525	0.04525	1.31	
Site.*Units* stratum					
Treat	2	0.20815	0.10407	3.01	0.072 ns
Must119vCult	1	0.19420	0.19420	5.61	0.028 *
Must119vNoCult	1	0.01394	0.01394	0.40	0.533 ns
Residual	20	0.69230	0.03461		
Total	23	0.94570			

Tables of contrasts 

Variate: logTotNem3

Must119vCult 0.220, s.e. 0.0930, ss.div. 4.00 Must119vNoCult 0.059, s.e. 0.0930, ss.div. 4.00

Tables of means

Variate: logTotNem3

Grand mean 1.816

Treat Cult NoCult Mustard119 1.688 1.850 1.909

Standard errors of means

Table Treat rep. 8 d.f. 20 e.s.e. 0.0658

Standard errors of differences of means

\_\_\_\_\_\_

Table Treat 8 rep. d.f. 20 0.0930 s.e.d.

Least significant differences of means (5% level)

Table Treat rep. 8 20 d.f. l.s.d. 0.1940

## 2.2.4. Difference between Total nematode counts 1 and 3.

280 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff, lsd, means; LSDLEVEL=5] DifflogTotNem13

Analysis of variance

Variate: DifflogTotNem13 Source of variation d.f. s.s. m.s. v.r. F pr. Site stratum 1 0.05649 0.05649 1.02 Site.\*Units\* stratum 2 0.57068 0.28534 5.13 0.016 \*
1 0.56709 0.56709 10.20 0.005 \*\*
1 0.10538 0.10538 1.90 0.184 ns
20 1.11158 0.05558 Treat Must119vCult Must119vNoCult Residual 23 Total 1.73875

Tables of contrasts

...................

Variate: DifflogTotNem13

Must119vCult 0.38, s.e. 0.118, ss.div. 4.00 Must119vNoCult 0.16, s.e. 0.118, ss.div. 4.00

Tables of means \_\_\_\_\_

Variate: DifflogTotNem13

Grand mean -0.176

Treat Cult NoCult Mustard119 -0.373 -0.159 0.003

Standard errors of means

Table rep. 8 d.f. 20 e.s.e. 0.0834

Standard errors of differences of means

8 d.f. 20 s.e.d. 0.1179

Least significant differences of means (5% level)

Treat rep. 8 d.f. 20 1.s.d. 0.2459

### 2.2.5. Lesion nematode count 1 (transformed to log10).

286 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff,lsd,means; LSDLEVEL=5] logLesion1

Analysis of variance -----

Variate: logLesion1

S.S. M... d.f. s.s. 1 0.00008 Source of variation v.r. F pr. Site stratum 0.00 Site.\*Units\* stratum Treat 2 0.15846 0.07923 1.33 0,286 ns 2.62 0.121 ns 0.99 0.331 ns Must119vCult 1 0.15573 0.15573 0.05883 Must119vNoCult 1 0.05883 20 1.18748 23 1.34601 Residual 0.05937 Total

Tables of contrasts

\_\_\_\_\_\_

Variate: logLesion1

Must119vCult -0.20, s.e. 0.122, ss.div. 4.00 Mustl19vNoCult -0.12, s.e. 0.122, ss.div. 4.00

Tables of means \_\_\_\_\_\_

Variate: logLesion1

Grand mean 1.945

Treat Cult NoCult Mustard119

2.036 1.960 1.838

#### Standard errors of means

Table Treat rep. 8 d.f. 20 e.s.e. 0.0861

## Standard errors of differences of means

Table Treat rep. 8

d.f. 20 s.e.d. 0.1218

#### Least significant differences of means (5% level)

 Table
 Treat

 rep.
 8

 d.f.
 20

 l.s.d.
 0.2541

## 2.2.6. Lesion nematode count 2 (transformed to log10).

292 ANOVA [PRINT=aovtable,information,means,contrast; FACT=32; FPROB=yes; PSE=diff,lsd, means; LSDLEVEL=5] logLesion2

# Analysis of variance

Variate: logLesion2

Variate: logLesion2
Source of variation d.f. s.s. m.s. v.r. F pr.
Site stratum 1 0.09541 0.09541 1.97
Site.\*Units\* stratum
Treat 2 0.74586 0.37293 7.69 0.003 \*\*
Mustll9vCult 1 0.31700 0.31700 6.53 0.019 \*
Mustll9vNoCult 1 0.72021 0.72021 14.84 <.001 \*\*\*
Residual 20 0.97047 0.04852
Total 23 1.81174

Tables of contrasts

Variate: logLesion2

Mustl19vCult 0.28, s.e. 0.110, ss.div. 4.00 Mustl19vNoCult 0.42, s.e. 0.110, ss.div. 4.00

Tables of means

Variate: logLesion2 Grand mean 1.797

Treat Cult NoCult Mustard119
1.750 1.608 2.032

#### Standard errors of means

Table Treat rep. 8 d.f. 20 e.s.e. 0.0779

## Standard errors of differences of means

Table Treat rep. 8 d.f. 20 s.e.d. 0.1101

Least significant differences of means (5% level)

Table Treat rep. 8 d.f. 20 l.s.d. 0.2297

## 2.2.7. Lesion nematode count 3 (transformed to log10).

298 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff, lsd, means; LSDLEVEL=5] logLesion3

## Analysis of variance

\*\*\*\*\*\*\*\*\*\*\*\*\*\*

Variate: logLesion3
Source of variation d.f. s.s. m.s. v.r. F pr.
Site stratum 1 0.00715 0.00715 0.13
Site.\*Units\* stratum
Treat 2 0.07785 0.03892 0.70 0.509 ns
Mustll9vCult 1 0.05765 0.05765 1.04 0.321 ns
Mustll9vNoCult 1 0.00001 0.00001 0.00 0.990 ns
Residual 20 1.11330 0.05567
Total 23 1.19830

Tables of contrasts

Variate: logLesion3

Mustl19vCult 0.12, s.e. 0.118, ss.div. 4.00 Mustl19vNoCult 0.00, s.e. 0.118, ss.div. 4.00

#### Tables of means

Variate: logLesion3
Grand mean 1.704

Treat Cult NoCult Mustard 119 1.623 1.745 1.743

#### Standard errors of means

Table Trea

rep. 8
d.f. 20
e.s.e. 0.0834

## Standard errors of differences of means

------

 Table
 Treat

 rep.
 8

 d.f.
 20

 s.e.d.
 0.1180

Least significant differences of means (5% level)

Table Treat

rep. 8 d.f. 20 l.s.d. 0.2461

#### 2.2.8. Difference between Lesion nematode counts 1 and 3.

304 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff, lsd, means; LSDLEVEL=5] DifflogLesion13

Analysis of variance

Variate: Diffloglesion13

variate: DiffiogLesion	13				
Source of variation	d.f	SS	m.s.	v.r.	F pr.
Site stratum	1	0.00575	0.00575	0.08	
Site.*Units* stratum					
Treat	2	C.41097	0.20548	2.98	0.074
Must119vCult	1	0.40289	0.40289	5.83	0.025
Must119vNoCult	1	0.05737	0.05737	0.83	0.373
Residual	20	1.38113	0.06906		
.Total	23	1.79785			

Tables of contrasts

----

Variate: DifflogLesion13

Mustl19vCult 0.32, s.e. 0.131, ss.div. 4.00

Mustl19vNoCult 0.12, s.e. 0.131, ss.div. 4.00

Tables of means

Variate: DifflogLesion13

Grand mean -0.241

Treat Cult NoCult Mustard119 -0.413 -0.215 -0.095

Standard errors of means

 Table
 Treat

 rep.
 8

 d.f.
 20

 e.s.e.
 0.0929

Standard errors of differences of means

-----

Table Treat rep. 8 d.f. 20 s.e.d. 0.1314

Least significant differences of means (5% level)

\_\_\_\_\_\_

Table Treat rep. 8 d.f. 20 1.s.d. 0.2741

end of Mustard 119 v. Nematodes

start of Mustard 99 v. Stubby and Stunt

## 3.1.1. Stubby nematode count 1 (transformed to log10).

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311 "General Analysis of Variance."
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312 BLOCK Site

313 TREATMENTS COMP(Treat;2;Cont)

315 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff, means | logStubby1

## Analysis of variance

=======================================	
Variate: logStubbyl	
Source of variation	d.f.
Site stratum	1
Site.*Units* stratum	
M	_

Site.*Units* stratum					
Treat	2	0.0083	0.0041	0.03	0.969
Must99vCult	1	0.0054	0.0054	0.04	0.842
Must99vNoCult	1	0.0001	0.0001	0.00	0.979
Residual	20	2.6708	0.1335		
Total	23	3.4720			

s.s. m.s. v.r. F pr. 0.7929 0.7929 5.94

## Tables of contrasts

Variate: logStubby1 Must99vCult 0.04, s.e. 0.183, ss.div. 4.00 Must99vNoCult 0.00, s.e. 0.183, ss.div. 4.00

## Tables of means

Variate: logStubby1 Grand mean 0.447

Treat Cult Mustard99 NoCult 0.421 0.458 0.463

#### Standard errors of means

------

Table	Treat
rep.	8
d.f.	20
e.s.e	0.1292

### Standard errors of differences of means

Table		Treat
rep.		8
d.f.	70	20
s.e.d.		0.1827

## 3.1.2. Stubby nematode count 2 (transformed to log10).

322 ANOVA [PRINT=aovtable,information,means,contrast; FACT=32; FPROB=yes; PSE=diff, means]logStubby2

#### Analysis of variance

CHURROSCHONGSCHUR Variate: logStubby2

Source of variation	d.f.	5.5	m.s.	v.r.	F pr
Site stratum	1	3.59012	3.59012	44.82	-

Site.\*Units\* stratum

2 0.58145 0.29073 3.63 0.045 1 0.41636 0.41636 5.20 0.034 Treat Must99vCult

1 0.45496 0.45496 5.68 0.027 Must99vNoCult

20 1.60209 0.08010 23 5.77366 Residual

Total

Tables of contrasts \_\_\_\_

Variate: logStubby2

Must99vCult 0.32, s.e. 0.142, ss.div. 4.00 Must99vNoCult 0.34, s.e. 0.142, ss.div. 4.00

Tables of means -----

Variate: logStubby2

Grand mean 0.507

Treat Cult Mustard99 NoCult

0.405 0.727 0.390

Standard errors of means

Table Treat rep. Я d.f. 20 e.s.e. 0.1001

Standard errors of differences of means

Table Treat 8 rep. d.f. 20 s.e.d. 0.1415

### 3.1.3. Stubby nematode count 2 (transformed to log10).

329 ANOVA [PRINT=aovtable,information,means,contrast; FACT=32; FPROB=yes; PSE=diff, means]logStubby3

Analysis of variance \_\_\_\_\_

Variate: logStubby3

Source of variation d.f. s.s. m.s. v.r. F pr. Site stratum 1 0.03120 0.03120 0.38

Site.\*Units\* stratum

2 0.05674 0.02837 0.34 0.714 Treat 0.00047 0.01 0.940 0.00047 Must99vCult

1 1 0.04680 0.57 0.461 0.08273 Must99vNoCult 0.04680

20 1.65450 23 1.74244 Residual 1.65450 Total

Tables of contrasts

\*\*\*\*\*\*\*\*\*\*\*\* Variate: logStubby3

Must99vCult 0.01, s.e. 0.144, ss\_div. 4.00

Must99vNoCult 0.11, s.e. 0.144, ss.div. 4.00

Tables of means -----

Variate: logStubby3

Grand mean 0.166

Treat Cult Mustard99 NoCult

0.195 0.205 0.097

#### Standard errors of means

Table Trea rep. d.f. 2

d.f. 20 e.s.e. 0.1017

## Standard errors of differences of means

\_\_\_\_\_\_

 Table
 Treat

 rep.
 8

 d.f.
 20

 s.e.d.
 0.1438

## 3.1.4. Difference Stubby nematode counts 1 and 3.

336 ANOVA [PRINT=aovtable,information,means,contrast; FACT=32; FPROB=yes; PSE=diff,means]DifflogStubby13

# Analysis of variance

Variate: DifflogStubby13

Source of variation d.f. s.s. m.s. Site stratum 1 0.5096 0.5096 v.r. F pr. 2.53 Site.\*Units\* stratum Treat 2 0.0873 0.22 0.807 0.01 0.909 0.0437 0.0027 0.01 0.909 0.0511 0.25 0.620 Must99vCult 1 0.0027 1 0.0027 1 0.0511 20 4.0336 23 4.6305 Must99vNoCult 0.0511 0.2017 Residual Total

Tables of contrasts

-----

Variate: DifflogStubby13

#### Tables of means

Variate: DifflogStubby13

Grand mean -0.282

Treat Cult Mustard99 NoCult -0.227 -0.253 -0.366

#### Standard errors of means

\_\_\_\_\_\_

 Table
 Treat

 rep.
 8

 d.f.
 20

 e.s.e.
 0.1588

### Standard errors of differences of means

 Table
 Treat

 rep.
 8

 d.f.
 20

 s.e.d.
 0.2245

## 3.1.5. Stunt nematode count 1 (transformed to log10).

342 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff,means]logStunt1

#### Analysis of variance

Variate: logStunt1 Source of variation d.f. s.s. m.s. v.r. F pr. Site stratum 1 0.0008 0.0008 0.01 Site.\*Units\* stratum 2 0.4116 0.2058 1.57 0.232 1 0.3835 0.3835 2.93 0.102 1 0.2069 0.2069 1.58 0.223 20 2.6146 0.1307 23 3.0270 Treat Must99vCult Must99vNoCult Residual

#### Tables of contrasts

Total

------

Variate: logStunt1

Must99vCult 0.31, s.e. 0.181, ss.div. 4.00 Must99vNoCult 0.23, s.e. 0.181, ss.div. 4.00

#### Tables of means

Variate: logStunt1 Grand mean 0.590

> Cult Mustard99 NoCult 0.460 0.769 0.542

#### Standard errors of means

Treat

8 rep. d.f. 20 0.1278 e.s.e.

#### Standard errors of differences of means

Table Treat 8 rep. d.f. 20 0.1808 s.e.d.

### 3.1.6. Stunt nematode count 2 (transformed to log10).

349 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff,means]logStunt2

#### Analysis of variance

\*\*\*\*\*\*\*\*\*\*\*\*\* Variate: logStunt2 d.f. s.s. m.s. 1 0.2069 0.2069 v.r. F pr. Source of variation Site stratum 0.80 Site.\*Units\* stratum 2 1.3428 0.6714 2.61 0.099 1 0.5161 0.5161 2.00 0.172 1 1.3148 1.3148 5.10 0.035 20 5.1518 0.2576 23 6.7015 Treat Must99vCult Must99vNoCult Residual Total

#### Tables of contrasts

-----

Variate: logStunt2

Must99vCult 0.36, s.e. 0.254, ss.div. 4.00 Must99vNoCult 0.57, s.e. 0.254, ss.div. 4.00

Tables of means ~==#=======

Variate: logStunt2 Grand mean 0.65

> Treat Cult Mustard99 NoCult 0.60 0.96

Standard errors of means

\_\_\_\_\_

Treat 8 d.f. 20 e.s.e. 0.179

Standard errors of differences of means

Table Treat rep. 8

d.f. 20 s.e.d. 0.254

## 3.1.7. Stunt nematode count 3 (transformed to log10).

356 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff, means)logStunt3

Analysis of variance ......

Variate: logStunt3

d.f. 5.2 1 0.3731 Source of variation m.s. v.r. F pr. Site stratum 0.3731 1.68 Site.\*Units\* stratum Treat 2 1.2109 0.6054 2.72 0.090 5.28 0.032 2.27 0.148 Must99vCult 1 1.1733 1.1733 Must99vNoCult 1 0.5033 0.5033 2.27 0.148 4.4440 Residual 20 0.2222

Total 23 6.0280

Tables of contrasts

\_\_\_\_\_ Variate: logStunt3

Must99vCult 0.54, s.e. 0.236, ss.div. 4.00 Must99vNoCult 0.35, s.e. 0.236, ss.div. 4.00

Tables of means

HENDRESS HERE Variate: logStunt3

Grand mean 0.491

Treat Cult Mustard99 NoCult. 0.248 0.789 0.435

Standard errors of means

Table Treat 8 rep.

d.f. 20 e.s.e. 0.1667

Standard errors of differences of means

Table Treat rep. 8

d.f. 20 s.e.d. 0.2357

## 3.1.8. Difference Stunt nematode counts 1 and 3.

363 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff, means] DifflogStunt13

Analysis of variance

Variate: DifflogStunt13

Source of variation d.f. s.s. m.s. v.r. F pr. Site stratum 1 0.3384 0.3384 1.87 Site.\*Units\* stratum 2 0.2159 0.1079 0.60 0.561 Must99vCult 1 0.2152 0.2152 1.19 0.289

Must99vCult 1 0.0648 0.0648 0.36 0.557
Residual 20 3.6247 0.1812

Total 23 4.1790

Tables of contrasts

Variate: DifflogStunt13

Must99vCult 0.23, s.e. 0.213, ss.div. 4.00 Must99vNoCult 0.13, s.e. 0.213, ss.div. 4.00

Tables of means

Variate: DifflogStunt13

Grand mean -0.100

Treat Cult Mustard99 NoCult
-0.212 0.020 -0.107

Standard errors of means

\_\_\_\_\_\_

 Table
 Treat

 rep.
 8

 d.f.
 20

 e.s.e.
 0.1505

Standard errors of differences of  $\ensuremath{\mathsf{means}}$ 

Table Treat rep. 8 d.f. 20 s.e.d. 0.2129

end of Mustard 99 stubby stunt start of Mustard 119 stubby stunt

# 3.2.1. Stubby nematode count 1 (transformed to log10).

314 ANOVA [PRINT=aovtable,information,means,contrast; FACT=32; FPROB=yes; PSE=diff,means]logStubbyl

## Analysis of variance

d.f.	S.S.	m.s.	v.r.	F pr.
1	3.0492	3.0492	26.86	_
2	0.0109	0.0055	0.05	0.953
1	0.0002	0.0002	0.00	0.965
1	0.0068	0.0068	0.06	0.810
20	2.2705	0.1135		
23	5.3307			
	2 1 1 20	1 3.0492 2 0.0109 1 0.0002 1 0.0068 20 2.2705	1 3.0492 3.0492 2 0.0109 0.0055 1 0.0002 0.0002 1 0.0068 0.0068 20 2.2705 0.1135	1 3.0492 3.0492 26.86 2 0.0109 0.0055 0.05 1 0.0002 0.0002 0.00 1 0.0068 0.0068 0.06 20 2.2705 0.1135

#### Tables of contrasts

Variate: logStubby1

Mustl19vCult 0.01, s.e. 0.168, ss.div. 4.00 Mustl19vNoCult -0.04, s.e. 0.168, ss.div. 4.00

# Tables of means

Variate: logStubby1 Grand mean 0.432

Treat Cult NoCult Mustard119 0.413 0.462 0.420

#### Standard errors of means

Table Treat rep. 8 d.f. 20

# Standard errors of differences of means

\_\_\_\_\_\_

 Table
 Treat

 rep.
 8

 d.f.
 20

 s.e.d.
 0.1685

# 3.2.2. Stubby nematode count 2 (transformed to log10).

321 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff, means]logStubby2

#### Analysis of variance

Variate: logStubby2					
Source of variation	d.f.	s.s.	m.s.	<b>v r</b>	F pr.
Site stratum	1	2.1658	2.1658	15.51	-
Site.*Units* stratum					
Treat	2	0.4934	0.2467	1.77	0.197
Must119vCult	1	0.4411	0.4411	3.16	0.091
Must119vNoCult	1	0.2811	0.2811	2.01	0.171
Residual	20	2.7926	0.1396		
Total	23	5.4518			

Tables of contrasts

------

Variate: logStubby2

Mustl19vCult 0.33, s.e. 0.187, ss.div. 4.00 Must119vNoCult 0.27, s.e. 0.187, ss.div. 4.00

Tables of means

-----Variate: logStubby2

Grand mean 0.351

Treat Cult NoCult Mustard119 0.218 0.285 0.550

Standard errors of means

Treat rep. 8

d.f. 20 e.s.e. 0.1321

Standard errors of differences of means

Table Treat rep. 8 d.f. 20 s.e.d. 0.1868

# 3.2.3. Stubby nematode count 3 (transformed to log10).

327 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff, means]logStubby3

Analysis of variance \_\_\_\_\_\_\_

Variate: logStubby3

d.f: s.s. m.s. 1 0.1296 0.1296 Source of variation v.r. F pr. Site stratum 0.95

Site.\*Units\* stratum

0.0448 0.33 0.725 0.0896 2 Treat Must119vCult

1 0.0707 0.0707 0.52 0.481 1 0.0002 0.0002 0.00 0.970 20 2.7367 0.1368 23 2.9558 Mustll9vNoCult

Residual

Total

Tables of contrasts \_\_\_\_

Variate: logStubby3

Mustl19vCult 0.13, s.e. 0.185, ss.div. 4.00 Mustl19vNoCult 0.01, s.e. 0.185, ss.div. 4.00

Tables of means THE RESIDENCE OF THE PARTY OF THE RESIDENCE

Variate: logStubby3

Grand mean 0.535

Treat Cult NoCult Mustard119 0.449 0.575 0.582

Standard errors of means

Table Treat rep. 8 d.f. 20 e.s.e. 0.1308

Standard errors of differences of means

------

 Table
 Treat

 rep.
 8

 d.f.
 20

 s.e.d.
 0.1850

# 3.2.4. Difference Stubby nematode counts 1 and 3.

333 ANOVA [PRINT=aovtable,information,means,contrast; FACT=32; FPROB=yes; PSE=diff,means]DifflogStubby13

Analysis of variance

Variate: DifflogStubby13

Variate: DifflogStubby13
Source of variation d.f. s.s. m.s. v.r. F pr.
Site stratum 1 1.9216 12.32
Site.\*Units\* stratum

Treat 2 0.0642 0.0321 0.21 0.816
Mustl19vCult 1 0.0630 0.0630 0.40 0.532
Mustl19vNoCult 1 0.0092 0.0092 0.06 0.810
Residual 20 3.1190 0.1559
Total 23 5.1048

Tables of contrasts

Variate: DifflogStubby13

Mustll9vCult 0.13, s.e. 0.197, ss.div. 4.00 Mustll9vNoCult 0.05, s.e. 0.197, ss.div. 4.00

Tables of means

Variate: DifflogStubby13

Grand mean 0.103

Treat Cult NoCult Mustard119
0.036 0.113 0.161

Standard errors of means

\_\_\_\_\_

 Table
 Treat

 rep.
 8

 d.f.
 20

 e.s.e.
 0.1396

Standard errors of differences of means

Table Treat rep. 8 d.f. 20 s.e.d. 0.1975

### 3.2.5. Stunt nematode count 1 (transformed to log10).

340 ANOVA [PRINT=aovtable,information,means,contrast; FACT=32; FPROB=yes; PSE=diff,means]logStunt1

# Analysis of variance \_\_\_\_\_ Variate: logStunt1 Source of variation d.f. s.s. m.s. v.r. F pr. Site stratum 1 1.0597 1.0597 9.65 Site.\*Units\* stratum 2 0.1906 0.0953 0.87 0.435 1 0.0075 0.0075 0.07 0.796 1 0.1714 0.1714 1.56 0.226 20 2.1968 0.1098 23 3.4472 Treat Must119vCult Must119vNoCult Residual Total

#### Tables of contrasts

Variate: logStunt1

Mustl19vCult -0.04, s.e. 0.166, ss.div. 4.00 Must119vNoCult -0.21, s.e. 0.166, ss.div. 4.00

## Tables of means

-----------

Variate: logStunt1 Grand mean 0.517

> Treat Cult NoCult Mustard 119 0.476 0.640 0.433

#### Standard errors of means

Table	Treat
rep.	8
d.f.	20
e.s.e.	0.1172

## Standard errors of differences of means

Table	Treat
rep.	8
d.f.	20
s.e.d.	0.1657

# 3.2.6. Stunt nematode count 2 (transformed to log10).

347 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff,means]logStunt2

## Analysis of variance

\_\_\_\_\_

Variate: logStunt2					
Source of variation	d.f.	s.s.	m.s.	v.r.	F pr
Site stratum	1	2.3456	2.3456	9.05	
Site.*Units* stratum					
Treat	2	0.5072	0.2536	0.98	0.393
Must119vCult	1	0.4271	0.4271	1.65	0.214
Must119vNoCult	1	0.3271	0.3271	1.26	0.275
Residual	20	5.1817	0.2591		
Total	23	8.0345			

Tables of contrasts

Variate: logStunt2

Must119vCult 0.33, s.e. 0.255, ss.div. 4.00 Must119vNoCult 0.29, s.e. 0.255, ss.div. 4.00

Tables of means -----

Variate: logStunt2

Grand mean 0.64

Treat Cult 0.52 NoCult Mustard119 0.56 0.85

Standard errors of means

\_\_\_\_\_\_

Table Treat rep. d.f. 20 e.s.e. 0.180

Standard errors of differences of means

Table Treat rep. 8 d.f 20 0.255 s.e.d.

3.2.7. Stunt nematode count 3 (transformed to log10).

354 ANOVA [PRINT=aovtable, information, means, contrast; FACT=32; FPROB=yes; PSE=diff, means]logStunt3

Analysis of variance

\_\_\_\_\_\_

Variate: logStunt3

Source of variation	d.f.	S.S.	m.s.	v.r.	F pr.
Site stratum	1	3.4642	3.4642	16.66	-
Site.*Units* stratum					
Treat	2	0.5726	0.2863	1.38	0.275
Must119vCult	1	0.3571	0.3571	1.72	0.205
Must119vNoCult	1	0.4912	0.4912	2.36	0.140
Residual	20	4.1597	0.2080		
Total	23	8.1965			

Tables of contrasts

Variate: logStunt3

Mustl19vCult 0.30, s.e. 0.228, ss.div. 4.00 Mustl19vNoCult 0.35, s.e. 0.228, ss.div. 4.00

Tables of means

\_\_\_\_\_\_\_\_\_\_

Variate: logStunt3 Grand mean 0.593

> Cult NoCult Mustard119 0.511 0.459 0.809

Standard errors of means

Table Treat 8 rep.

d.f. 20 e.s.e. 0.1612

Standard errors of differences of means

Table Treat rep. 8 d.f. 20

d.f. s.e.d.

#### 3.2.8. Difference Stunt nematode counts 1 and 3.

0.2280

360 ANOVA [PRINT=aovtable,information,means,contrast; FACT=32; FPROB=yes; PSE=diff,means]DifflogStunt13

## Analysis of variance

Variate: DifflogStunt13

Source of variation d.f. s.s. m.s. v.r. F pr. Site stratum 1 0.6919 0.6919 3.69

Site.\*Units\* stratum

Treat 2 1.2643 0.6321 3.37 0.055

Must119vCult 1 0.4682 0.4682 2.50 0.130

Must119vNoCult 1 1.2428 1.2428 6.63 0.018

Posidual 20 3.7462 0.1873

Residual 20 3.7462 0.1873

Total 23 5.7024

# Tables of contrasts

Variate: DifflogStunt13

Mustl19vCult 0.34, s.e. 0.216, ss.div. 4.00 Mustl19vNoCult 0.56, s.e. 0.216, ss.div. 4.00

# Tables of means

ARRESCHARICHE

Variate: DifflogStunt13
Grand mean 0.076

Treat Cult NoCult Mustard119 0.034 -0.181 0.376

#### Standard errors of means

\_\_\_\_

 Table
 Treat

 rep.
 8

 d.f.
 20

 e.s.e.
 0.1530

# Standard errors of differences of means

Table Treat rep. 8 d.f. 20 s.e.d. 0.2164

# end of Mustard 119 stubby stunt

# 4. Cultivated v Mustard 99/Mustard 119, difference between counts 2 and 3.

# 4.1. Difference between log nematode counts 2 and 3 for Mustard 99 (both sites).

# 4.1.1. Difference between log total nematodes counts 2 and 3, Mustard 99

335 "General Analysis of Variance."

336 BLOCK Site

337 TREATMENTS COMP(Treat;2;Cont)

338 COVARIATE "No Covariate"

339 ANOVA []difflogTot23

### Analysis of variance

Variate: difflogTot23

·					
Source of variation	d.f.	s.s.	m.s.	v.r.	F pr
Site stratum	1	0.00068	0.00068	0.01	
Site.*Units* stratum					
Treat	2	0.40823	0.20411	4.04	0.034
Must99vCult	1	0.30994	0.30994	6.13	0.022
Must99vNocult	1	0.30236	0.30236	5.98	0.024
Residual	20	1.01163	0.05058		
Total	23	1.42053			

#### Tables of contrasts

-----

Variate: difflogTot23

Must99vCult -0.28, s.e. 0.112, ss.div. 4.00 Must99vNocult -0.27, s.e. 0.112, ss.div. 4.00

# Tables of means

-----

Variate: difflogTot23 Grand mean -0.145

Treat Cult Mustard99 NoCult
-0.051 -0.329 -0.055

### Standard errors of means

Table Treat
rep. 8
d.f. 20
e.s.e. 0.0795

### Standard errors of differences of means

------

Table Treat rep. 8

d.f. 20 s.e.d. 0.1125

## 4.1.2. Difference between log Lesion nematodes counts 2 and 3, Mustard 99

341 "General Analysis of Variance."

345 ANOVA []difflogLesion23

Analysis of Variance

Variate: difflogLesion23

Source of variation d.f. 5.5. m.s. v.r. F pr. 0.00007 0.00007 0:00 Site stratum Site.\*Units\* stratum 2 0.50830 0.25415 5.64 0.011 Treat Must99vCult 1 0.32282 0.32282 7.17 0.014 Must99vNocult 1 0.43173 0.43173 9.58 0.006 20 0.90095 0.04505 Residual Total 23 1.40931

Tables of contrasts

\_\_\_\_\_

Variate: difflogLesion23

Must99vCult -0.28, s.e. 0.106, ss.div. 4.00 Must99vNocult -0.33, s.e. 0.106, ss.div. 4.00

Tables of means

-----

Variate: difflogLesion23

Grand mean -0.096

Treat Cult Mustard99 NoCult
-0.016 -0.300 0.029

Standard errors of means

Table Treat
rep. 8
d.f. 20
e.s.e. 0.0750

Standard errors of differences of means

\_\_\_\_\_

Table Treat
rep. 8
d.f. 20
s.e.d. 0.1061

## 4.1.3. Difference between log Stunt nematodes counts 2 and 3, Mustard 99

347 "General Analysis of Variance."

351 ANOVA []difflogStunt23

Analysis of variance

Variate: difflogStunt23

Source of variation d.f. s.s. m.s. v.r. F pr.

Site stratum	1	0.0243	0.0243	0.11	
Site.*Units* stratum					
Treat	2	0.6449	0.3225	1.52	0.243
Must99vCult	1	0.1331	0.1331	0.63	0.438
Must99vNocult	1	0.1911	0.1911	0.90	0.354
Residual	20	4.2473	0.2124		
Total	23	4.9166			10

#### Tables of contrasts

.

Variate: difflogStunt23

Must99vCult 0.18, s.e. 0.230, ss.div. 4.00 Must99vNocult -0.22, s.e. 0.230, ss.div. 4.00

#### Tables of means

Variate: difflogStunt23

Grand mean -0.154

Treat Cult Mustard99 NoCult -0.349 -0.167 0.052

#### Standard errors of means

Table Treat rep. d.f. 20

e.s.e. 0.1629

## Standard errors of differences of means

......

Table Treat rep. 8 d.f. 20 0.2304 s.e.d.

# 4.1.4. Difference between log Stubby nematodes counts 2 and 3, Mustard 99

353 "Géneral Analysis of Variance."

357 ANOVA []difflogStubby23

Analysis of variance

Variate: difflogStubby23

 
 Source of variation
 d.f.
 s.s.
 m.s.
 v.r.
 F pr.

 Site stratum
 1
 2.9520
 2.9520
 18.38
 Site.\*Units\* stratum

2 0.4173 0.2087 1.30 0.295 1 0.3887 0.3887 2.42 0.135 Must99vCult 1 0.2099 20 3.2126 23 6.5819 0.2099 1.31 0.266 Must99Nocult 0.1606

Residual Total 23

Tables of contrasts

\*\*\*\*\*\*\*\*\*\*\*

Variate: difflogStubby23

Must99vCult -0.31, s.e. 0.200, ss.div. 4.00 Must99vNocult -0.23, s.e. 0.200, ss.div. 4.00

Tables of means

\_\_\_\_\_\_

Variate: difflogStubby23

Grand mean -0.342

Treat Cult Mustard99 NoCult
-0.210 -0.522 -0.293

#### Standard errors of means

\_\_\_\_\_

Table Treat
rep. 8
d.f. 20
e.s.e. 0.1417

#### Standard errors of differences of means

Table Treat
rep. 8
d.f. 20
s.e.d. 0.2004

# 4.2. Difference between nematodes counts 2 and 3, Mustard 99, Elveden

# 4.2.1. Difference log Total nematodes counts 2 and 3, Mustard 99, Elveden

372 "General Analysis of Variance."

376 ANOVA []difflogTot23

Analysis of variance

\*\*\*\*\*

Variate: difflogTot23

 Source of variation
 d.f.
 s.s.
 m.s.
 v.r.
 F pr.

 Treat
 2
 0.14462
 0.07231
 1.06
 0.387

 Must99vCult
 1
 0.06522
 0.06522
 0.95
 0.355

 Must99vNocult
 1
 0.13818
 0.13818
 2.02
 0.189

Residual 9 0.61637 0.06849

Total 11 0.76100

# Tables of contrasts

Variate: difflogTot23

Must99vCult -0.18, s.e. 0.185, ss.div. 2.00 Must99vNocult -0.26, s.e. 0.185, ss.div. 2.00

Tables of means

\_\_\_\_

Variate: difflogTot23 Grand mean -0.140

Treat Cult Mustard99 NoCult
-0.107 -0.288 -0.025

#### Standard errors of means

rep. d.f. 9 e.s.e. 0.1308

#### Standard errors of differences of means

\_\_\_\_\_

Table Treat rep. 4 d.f. 9 s.e.d. 0.1850

# 4.2.2. Difference log Lesion nematodes counts 2 and 3, Mustard 99, Elveden

378 "General Analysis of Variance."

382 ANOVA []difflogLesion23

Analysis of variance

-----

Variate: difflogLesion23

Source of variation d.f. s.s. m.s. v.r. F pr. 2 0.12061 0.06031 0.82 0.470 Treat Must99vCult 1 0.06935 0.06935 0.94 0.357 Must99vNocult 1 0.10742 0.10742 1.46 0.257

Residual 9 0.66103 0.07345

Total 11 0.78164

#### Tables of contrasts

\_\_\_\_\_

Variate: difflogLesion23

Must99vCult -0.19, s.e. 0.192, ss.div. 2.00 Must99vNocult -0.23, s.e. 0.192, ss.div. 2.00

# Tables of means

\_\_\_\_\_

Variate: difflogLesion23

Grand mean -0.094

Treat Cult Mustard99 NoCult -0.047 -0.233 -0.002

### Standard errors of means

Table 4 rep.

9 d.f.

#### Standard errors of differences of means

\_\_\_\_\_\_\_

Table Treat rep.

d.f. 9 s.e.d. 0.1916

## 4.2.3. Difference log Stunt nematodes counts 2 and 3, Mustard 99, Elveden

384 "General Analysis of Variance."

388 ANOVA []difflogStunt23

Analysis of variance

Variate: difflogStunt23

Source of variation d.f. 5.5 m.s. v.r F pr. 2 0.4829 1 0.0051 1 0.4023 Treat 2 0.2415 1.19 0.347 Must99vCult 0.0051 0.03 0.878 0.4023 1.99 0.192 Must99vNocult Residual 9 1.8189 0.2021 2.3018 Total 11

Tables of contrasts

Variate: difflogStunt23

Must99vCult -0.05, s.e. 0.318, ss.div. 2.00 Must99vNocult -0.45, s.e. 0.318, ss.div. 2.00

Tables of means

\*\*\*\*\*\*

Variate: difflogStunt23

Grand mean -0.12

Treat Cult Mustard99 NoCult
-0.24 -0.29 0.16

Standard errors of means

\_\_\_\_\_

 Table
 Treat

 rep.
 4

 d.f.
 9

 e.s.e.
 0.225

Standard errors of differences of means

-----

 Table
 Treat

 rep.
 4

 d.f.
 9

 s.e.d.
 0.318

# 4.2.4. Difference log Stubby nematodes counts 2 and 3, Mustard 99, Elveden

390 "General Analysis of Variance."

394 ANOVA []difflogStubby23

Analysis of variance

Variate: difflogStubby23

Source of variation d.f. s.s. m.s. v.r. F pr. Treat 2 0.0114 0.0057 0.02 0.976 Must99vCult 1 0.0054 0.0054 0.02 0.882

```
Must99vNocult 1 0.0009 0.0009 0.00 0.952
Residual 9 2.0876 0.2320
Total 11 2.0989
```

#### Tables of contrasts

-----

Variate: difflogStubby23

Must99vCult 0.05, s.e. 0.341, ss.div. 2.00 Must99vNocult -0.02, s.e. 0.341, ss.div. 2.00

#### Tables of means

-----

Variate: difflogStubby23

Grand mean -0.69

Treat Cult Mustard99 NoCult
-0.73 -0.68 -0.66

#### Standard errors of means

 Table
 Treat

 rep.
 4

 d.f.
 9

 e.s.e.
 0.241

#### Standard errors of differences of means

-----

 Table
 Treat

 rep.
 4

 d.f.
 9

 s.e.d.
 0.341

# 4.3. Difference between log nematodes counts 2 and 3 for Mustard 99, Knights

# 4.3.1. Difference log Total nematodes counts 2 and 3 for Mustard 99, Knights

409 "General Analysis of Variance."

413 ANOVA []difflogTot23

Analysis of variance

Variate: difflogTot23

Source of variation	d.f.	s.s.	m.s.	v.r.	F pr.
Treat	2	0.30906	0.15453	3.98	0.058
Must99vCult	1	0.28295	0.28295	7.28	0.024
Must99vNocult	1	0.16476	0.16476	4.24	0.070
Residual	9	0.34980	0.03887		
Total	11	0.65886			

#### Tables of contrasts

Variate: difflogTot23

Must99vCult -0.38, s.e. 0.139, ss.div. 2.00 Must99vNocult -0.29, s.e. 0.139, ss.div. 2.00

#### Tables of means

Variate: difflogTot23 Grand mean ~0.150

Treat Cult Mustard99 NoCult
0.005 -0.371 -0.084

#### Standard errors of means

\_\_\_\_\_

 Table
 Treat

 rep.
 4

 d.f.
 9

 e.s.e.
 0.0986

### Standard errors of differences of means

 Table
 Treat

 rep.
 4

 d.f.
 9

 s.e.d.
 0.1394

# 4.3.2. Difference log Lesion nematodes counts 2 and 3 for Mustard 99, Knights

415 "General Analysis of Variance."

419 ANOVA []difflogLesion23

Analysis of variance

-----

Variate: difflogLesion23

 Source of variation
 d.f.
 s.s.
 m.s.
 v.r.
 F pr.

 Treat
 2
 0.43821
 0.21910
 10.41
 0.005

 Must99vCult
 1
 0.29179
 0.29179
 13.87
 0.005

 Must99vNocult
 1
 0.36176
 0.36176
 17.19
 0.002

 Residual
 9
 0.18939
 0.02104

Total 11 0.62760

Tables of contrasts

Variate: difflogLesion23

Must99vCult -0.38, s.e 0.103, ss.div. 2.00 Must99vNocult -0.43, s.e. 0.103, ss.div. 2.00

Tables of means

-----

Variate: difflogLesion23

Grand mean -0.097

Treat Cult Mustard99 NoCult 0.016 -0.366 0.059

#### Standard errors of means

------

Table Treat
rep. 4
d.f. 9
e.s.e. 0.0725

#### Standard errors of differences of means

\_\_\_\_\_\_

Table Treat
rep. 4
d.f. 9
s.e.d. 0.1026

# 4.3.3. Difference log Stunt nematodes counts 2 and 3 for Mustard 99, Knights

421 "General Analysis of Variance."

425 ANOVA []difflogStunt23

Analysis of variance

Variate: difflogStunt23

Source of variation d.f. s.s. m.s. v.r. F pr. Treat 2 0.4475 0.2237 0.94 0.426 Must99vCult 1 0.3448 0.3448 1.45 0.260 Must99vNocult 1 0.0003 0.0003 0.00 0.975 Residual 9 2.1429 0.2381 Total 11 2.5904

# Tables of contrasts

Variate: difflogStunt23

Must99vCult 0.42, s.e. 0.345, ss.div. 2.00 Must99vNocult 0.01, s.e. 0.345, ss.div. 2.00

Tables of means

Variate: difflogStunt23

Grand mean -0.19

Treat Cult Mustard99 NoCult
-0.46 -0.04 -0.06

#### Standard errors of means

-----

 Table
 Treat

 rep.
 4

 d.f.
 9

 e.s.e.
 0.244

#### Standard errors of differences of means

Table	Treat
rep.	4
d.f.	9
s.e.d.	0.345

# 4.3.4. Difference log Stubby nematodes counts 2 and 3 for Mustard 99, Knights

427 "General Analysis of Variance."

431 ANOVA []difflogStubby23

Analysis of variance

\_\_\_\_\_

Variate: difflogStubby23

Source of variation	d.f.	s.s.	m.s.	v.r.	F pr.
Treat	2	0.93916	0.46958	7.14	0.014
Must99vCult	1	0.91289	0.91289	13.88	0.005
Must99vNocult	1	0.38202	0.38202	5.81	0.039
Residual	9	0.59181	0.06576		
Total	11	1.53096			

# Tables of contrasts

Variate: difflogStubby23

Must99vCult -0.68, s.e. 0.181, ss.div. 2.00 Must99vNocult -0.44, s.e. 0.181, ss.div. 2.00

# Tables of means

Variate: difflogStubby23

Grand mean 0.009

Treat Cult Mustard99 NoCult
0.314 -0.362 0.075

#### Standard errors of means

Table Treat
rep. 4
d.f. 9
e.s.e. 0.1282

#### Standard errors of differences of means

Table Treat

rep. 4
d.f. 9
s.e.d. 0.1813

### 4.4. Difference between log nematodes counts 2 and 3 for Mustard 119

# 4.4.1. Difference between log Total nematode counts 2 and 3 for Mustard 119

- -3 C:/ajpNowNew/aaShocklandNov2005/Analysis1/ajpMustard 1191.GSH"
- 307 CALCULATE [SEED=18467] difflogStunt23=logStunt3-logStunt2
- 310 CALCULATE [SEED=18467] difflogStubby23=logStubby3-logStubby2
- 332 "General Analysis of Variance."
- 333 BLOCK Site
- 334 TREATMENTS COMP(Treat; 2; Cont)
- 336 ANOVA []difflogTot23

Analysis of variance

------

Variate:	difflogTot23
variace.	difficult 23

Source of variation	d.f.	s.s.	m.s.	v.r.	F pr.
Site stratum	1	0.14347	0.14347	1.73	
Site.*Units* stratum		39			
Treat	2	0.66100	0.33050	3.98	0.035
Must119vCult	1	0.26780	0.26780	3.22	0.088
Must119vNocult	1	0.64287	0.64287	7.73	0.012
Residual	20	1.66230	0.08311		
Total	23	2.46677			

#### Tables of contrasts

#### ------

Variate: difflogTot23

Must119vCult 0.26, s.e. 0.144, ss.div. 4.00 Must119vNocult 0.40, s.e. 0.144, ss.div. 4.00

### Tables of means

\*\*\*\*\*\*\*\*\*\*\*

Variate: difflogTot23 Grand mean -0.056

> Treat Cult NoCult Mustard119 -0.095 0.164 -0.237

#### Standard errors of means

Table Treat 8 d.f. 20 e.s.e. 0.1019

#### Standard errors of differences of means

Table Treat rep. 8 d.f. 20 s.e.d. 0.1441

# 4.4.2. Difference between log Lesion nematodes counts 2 and 3 for Mustard **119**

- 344 "General Analysis of Variance."
- 345 BLOCK Site
- 348 ANOVA [ difflogLesion23

#### Analysis of variance

Variate: difflogLesion23

Source of variation d.f. s.s. m.s. v.r. F pr. Site stratum 1 0.0503 0.0503 0.45 Site.\*Units\* stratum

Treat 2 0.7395 0.3697 3.32 0.057 Mustl19vCult 1 0.2796 0.2796 2.51 0.129 Mustl19vNocult 1 0.7253 0.7253 6.51 0.019 Residual 20 2.2294 0.1115

Total 23 3.0192

#### Tables of contrasts

\*\*\*\*\*\*\*\*\*\*\*

Variate: difflogLesion23

Mustl19vCult 0.26, s.e. 0.167, ss.div. 4.00 Mustl19vNocult 0.43, s.e. 0.167, ss.div. 4.00

Tables of means

Variate: difflogLesion23

Grand mean ~0.093

Treat Cult NoCult Mustard119
-0.127 0.137 -0.289

#### Standard errors of means

 Table
 Treat

 rep.
 8

 d.f.
 20

 e.s.e.
 0.1180

#### Standard errors of differences of means

\_\_\_\_\_\_

 Table
 Treat

 rep.
 8

 d.f.
 20

 s.e.d.
 0.1669

# 4.4.3. Difference between log Stunt nematodes counts 2 and 3 for Mustard 119

350 "General Analysis of Variance."

351 BLOCK Site

354 ANOVA []difflogStunt23

Analysis of variance

Variate: difflogStunt23

Source of variation d.f. s.s. m.s. v.r. F pr. Site stratum 1 0.1087 0.1087 0.45

Site.\*Units\* stratum

Treat 2 0.0360 0.0180 0.07 0.928

Must119vCult	1	0.0342	0.0342	0.14	0.710
Must119vNocult	1	0.0166	0.0166	0.07	0.795
Residual	20	4.8185	0.2409		
Total	23	4.9632			

Tables of contrasts

----------

Variate: difflogStunt23

Mustl19vCult -0.09, s.e. 0.245, ss.div. 4.00 Mustl19vNocult -0.06, s.e. 0.245, ss.div. 4.00

Tables of means

-----

Variate: difflogStunt23

Grand mean -0.05

Treat Cult NoCult Mustard119
-0.01 -0.10 -0.04

Standard errors of means

Table Treat rep. 8 d.f. 20

20 e.s.e. 0.174

Standard errors of differences of means

\_\_\_\_\_

Table Treat
rep. 8
d.f. 20
s.e.d. 0.245

# 4.4.4. Difference between log Stubby nematodes counts 2 and 3 for Mustard 119

356 "General Analysis of Variance."

357 BLOCK Site

360 ANOVA []difflogStubby23

Analysis of variance

Variate: difflogStubby23

 Source of variation
 d.f.
 s.s.
 m.s.
 v.r.
 F pr.

 Site stratum
 1
 1.2359
 1.2359
 5.52

Site.\*Units\* stratum

Treat 2 0.2927 0.1464 0.65 0.531
Mustl19vCult 1 0.0139 0.0139 0.06 0.806
Mustl19vNocult 1 0.2665 0.2665 1.19 0.288

Residual 20 4.4779 0.2239 Total 23 6.0065

Tables of contrasts

\*\*\*\*\*\*\*\*\*\*\*\*\*\*\*

Variate: difflogStubby23

Mustl19vCult 0.06, s.e. 0.237, ss.div. 4.00 Mustl19vNocult 0.26, s.e. 0.237, ss.div. 4.00

Tables of means

Variate: difflogStubby23

Grand mean 0.184

Treat Cult NoCult Mustard119
0.231 0.290 0.032

#### Standard errors of means

\_\_\_\_\_

Table	Treat
rep.	8
d.f.	20
e.s.e.	0.1673

#### Standard errors of differences of means

Table Treat
rep. 8
d.f. 20
s.e.d. 0.2366

# 4.5.1. Difference log Total nematodes counts 2 and 3 for Mustard 119, Elveden

375 "General Analysis of Variance."

379 ANOVA []difflogTot23

Analysis of variance

Variate: difflogTot23

 Source of variation
 d.f.
 s.s.
 m.s.
 v.r.
 F pr.

 Treat
 2
 0.0916
 0.0458
 0.43
 0.661

 Must119vCult
 1
 0.0871
 0.0871
 0.83
 0.387

 Must119vNocult
 1
 0.0422
 0.0422
 0.40
 0.543

Residual 9 0.9499 0.1055

Total 11 1.0415

# Tables of contrasts

Variate: difflogTot23

Mustl19vCult 0.21, s.e. 0.230, ss.div. 2.00 Mustl19vNocult 0.15, s.e. 0.230, ss.div. 2.00

Tables of means

Variate: difflogTot23 Grand mean -0.133

Treat Cult NoCult Mustard119

-0.224 -0.015 -0.161

#### Standard errors of means

------

Table rep. 4 d.f. 9 e.s.e. 0.1624

#### Standard errors of differences of means

\_\_\_\_\_\_

Table Treat rep. 4 d.f. 9 s.e.d. 0.2297

# 4.5.2. Difference log Lesion nematodes counts 2 and 3 for Mustard 119, Elveden

381 "General Analysis of Variance."

385 ANOVA []difflogLesion23

Analysis of variance -----

Variate: difflogLesion23

Variate: diffiogresion25

Source of variation d.f. s.s. m.s. v.r. F pr.

Treat 2 0.0647 0.0324 0.27 0.772

Must119vCult 1 0.0644 0.0644 0.53 0.485

Must119vNocult 1 0.0122 0.0122 0.10 0.758 Residual 9 1.0920 0.1213

Total 11 1.1568

Tables of contrasts ------

Variate: difflogLesion23

Must119vCult 0.18, s.e. 0.246, ss.div. 2.00 Must119vNocult 0.08, s.e. 0.246, ss.div. 2.00

Tables of means

\_\_\_\_\_\_

Variate: difflogLesion23

Grand mean -0.14

Treat Cult NoCult Mustard119 -0.23 -0.05 -0.13

Standard errors of means

\_\_\_\_\_

Table Treat rep. d.f. 9 0.174 e.s.e.

Standard errors of differences of means

-----

Table Treat rep. d.f. 9 s.e.d. 0.246

## 4.5.3. Difference log Stunt nematodes counts 2 and 3 for Mustard 119, Elveden

387 "General Analysis of Variance."

391 ANOVA []difflogStunt23

Analysis of variance ------

Variate: difflogStunt23

s.s. Source of variation d.f. m.s. v.r. F pr. 0.0137 0.0069 0.02 0.981 2 Treat Mustll9vCult 1 0.0121 0.0121 0.03 0.858 Must119vNocult 0.0081 0.0081 0.02 0.883 1 3.2227 Residual 9 0.3581 Total 11 3.2364

Tables of contrasts

\*\*\*\*\*\*

Variate: difflogStunt23

Must119vCult -0.08, s.e. 0.423, ss.div. 2.00 Must119vNocult -0.06, s.e. 0.423, ss.div. 2.00

Tables of means

-----

Variate: difflogStunt23

Grand mean -0.12

Treat Cult NoCult Mustard119 -0.09 -0.16 -0.10

Standard errors of means 

Table Treat rep. d.f. 9 0.299 e.s.e.

Standard errors of differences of means

\_\_\_\_\_

Table Treat rep. 4 d.f. 9 s.e.d. 0.423

# 4.5.4. Difference log Stubby nematodes counts 2 and 3 for Mustard 119, Elveden

393 "General Analysis of Variance."

397 ANOVA []difflogStubby23

Analysis of variance

\_\_\_\_\_

Variate: difflogStubby23

Source of variation	d.f.	S.S	m.s.	v.r.	F pr.
Treat	2	0.5495	0.2748	0.79	0.483
Must119vCult	1	0.1953	0.1953	0.56	0.473
Must119vNocult	1	0.5422	0.5422	1.56	0.243
Residual	9	3.1265	0.3474		
Total	11	3.6760			

#### Tables of contrasts

-----

Variate: difflogStubby23

Mustl19vCult 0.31, s.e. 0.417, ss.div. 2.00 Mustl19vNocult 0.52, s.e. 0.417, ss.div. 2.00

Tables of means

Variate: difflogStubby23

Grand mean -0.04

Treat Cult NoCult Mustard119
-0.08 0.24 -0.29

# Standard errors of means

Table Treat rep. 4 d.f. 9

Standard errors of differences of means

0.295

\_\_\_\_\_\_\_\_

 Table
 Treat

 rep.
 4

 d.f.
 9

 s.e.d.
 0.417

\*\*\*\*\*\*\*\*\*NOW RESTRICTED TO KNIGHTS, still MUSTARD 119\*

# 4.6.1. Difference log Total nematodes counts 2 and 3 for Mustard 119, Knights

412 "General Analysis of Variance."

416 ANOVA []difflogTot23

Analysis of variance

Variate: difflogTot23

Source of variation d.f. s.s. m.s. v.r. F pr. 2 Treat 0.86323 0.43162 9.28 0.006 Must119vCult 1 0.19066 0.19066 4.10 0.074 1 0.86221 0.86221 18.54 0.002 Must119vNocult Residual 9 0.41858 0.04651 Total 11 1.28181

Tables of contrasts

----

Variate: difflogTot23

Must119vCult 0.31, s.e. 0.152, ss.div. 2.00 Must119vNocult 0.66, s.e. 0.152, ss.div. 2.00

Tables of means

------

Variate: difflogTot23
Grand mean 0.021

Treat Cult NoCult Mustard119
0.034 0.343 -0.313

#### Standard errors of means

 Table
 Treat

 rep.
 4

 d.f.
 9

 e.s.e.
 0.1078

#### Standard errors of differences of means

 Table
 Treat

 rep.
 4

 d.f.
 9

 s.e.d.
 0.1525

# 4.6.2. Difference log Lesion nematodes counts 2 and 3 for Mustard 119, Knights

418 "General Analysis of Variance."

422 ANOVA []difflogLesion23

Analysis of variance

Variate: difflogLesion23

 Source of variation
 d.f.
 s.s.
 m.s.
 v.r.
 F pr.

 Treat
 2
 1.20027
 0.60013
 8.83
 0.008

 Mustl19vCult
 1
 0.24406
 0.24406
 3.59
 0.091

 Mustl19vNocult
 1
 1.19654
 1.19654
 17.60
 0.002

Residual 9 0.61182 0.06798

Total 11 1.81209

### Tables of contrasts

Variate: difflogLesion23

Mustl19vCult 0.35, s.e. 0.184, ss.div. 2.00 Mustl19vNocult 0.77, s.e. 0.184, ss.div. 2.00

Tables of means

Variate: difflogLesion23

Grand mean -0.047

Treat Cult NoCult Mustard119
-0.022 0.327 -0.447

#### Standard errors of means

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Table Treat
rep. 4
d.f. 9
e.s.e. 0.1304

### Standard errors of differences of means

\_\_\_\_\_

Table Treat
rep. 4
d.f. 9
s.e.d. 0.1844

# 4.6.3. Difference log Stunt nematodes counts 2 and 3 for Mustard 119, Elveden

424 "General Analysis of Variance."

428 ANOVA []difflogStunt23

Analysis of variance

Variate: difflogStunt23

 Source of variation
 d.f.
 s.s.
 m.s.
 v.r.
 F pr.

 Treat
 2
 0.0234
 0.0117
 0.07
 0.937

 Mustl19vCult
 1
 0.0230
 0.0230
 0.13
 0.727

 Mustl19vNocult
 1
 0.0085
 0.0085
 0.05
 0.832

 Residual
 9
 1.5947
 0.1772

Total 11 1.6181

#### Tables of contrasts

-----

Variate: difflogStunt23

Must119vCult -0.11, s.e. 0.298, ss.div. 2.00 Must119vNocult -0.07, s.e. 0.298, ss.div. 2.00

# Tables of means

\_\_\_\_\_

Variate: difflogStunt23

Grand mean 0.02

Treat Cult NoCult Mustard119 0.07 -0.04 0.03

#### Standard errors of means

Table Treat rep. 4 d.f. 9 e.s.e. 0.210

#### Standard errors of differences of means

-----

Table Treat

rep. 4
d.f. 9
s.e.d. 0.298

# 4.6.4. Difference log Stubby nematodes counts 2 and 3 for Mustard 119, Elveden

430 "General Analysis of Variance."

434 ANOVA []difflogStubby23

Analysis of variance

----

Variate: difflogStubby23

Source of variation d.f. s.s.
Treat 2 0.0987 5.5. m.s. v.r. F pr. 0.0493 0.45 0.654 Must119vCult 1 0.0757 0.0757 0.68 0.430 0.0000 0.00 0.985 Mustl19vNocult 1 0.0000 Residual 9 0.9959 0.1107

Total 11 1.0946

#### Tables of contrasts

\_\_\_\_\_

Variate: difflogStubby23

Must119vCult -0.19, s.e. 0.235, ss.div. 2.00 Must119vNocult 0.00, s.e. 0.235, ss.div. 2.00

Tables of means

Variate: difflogStubby23

Grand mean 0.411

Treat Cult NoCult Mustard119 0.540 0.345 0.349

#### Standard errors of means

Table Treat
rep. 4
d.f. 9
e.s.e. 0.1663

#### Standard errors of differences of means

\_\_\_\_\_

 Table
 Treat

 rep.
 4

 d.f.
 9

 s.e.d.
 0.2352

AJP.

# Statistical analysis of soil nutrient data, including N

All analyses were performed using Genstat 8.

Means (n=4) and their individual standard errors are tabulated in Table 1 for each treatment at each site.

NB. June = pre-drilling, Aug = pre-incorporation Sep = six weeks post-incorporation

Table 1. HDC Soil analyses - means and standard errors

1.a. Nitrogen at 30 and 60cm

Group n	neans (N=4)	N30June	N60June	N30Aug	N60Aug	N30Sep	N60Sep
Elveden	Mustard 99	3.78	2.43	9.00	6.57	19.62	15.68
Elveden	None	3.50	2.42	34.23	22.06	22.85	13.87
Knights	Mustard 99	3.52	2.14	9.00	5.18	9.17	14.60
Knights	None	4.18	2.68	9.47	12.81	8.44	13.45
Elveden	Mustard 119	4.16	2.99	7.48	7.27	19.44	14.52
Elveden	None	3.98	2.27	31,21	18.30	17.32	22.07
Knights	Mustard 119	3.81	2.13	6.01	5.59	9.11	11.20
Knights	None	3.28	2.59	6.93	6.84	8.29	8.56
Standard	d errors	N30June	N60June	N30Aug	N60Aug	N30Sep	N60Sep
Elveden	Mustard 99	0.83	0.34	0.36	0.97	2.00	2.96
Elveden	None	0.51	0.26	2.56	1.17	4.03	4.18
Knights	Mustard 99	0.45	0.23	1.69	0.66	0.91	0.74
Knights	None	0.92	0.43	0.39	2.81	0.29	3.14
Elveden	Mustard 119	0.15	0.24	0.16	0.94	3.20	1.76
Elveden	None	0.50	0.43	1.40	3.78	2.83	5.34
Knights	Mustard 119	0.48	0.22	1.27	1.71	0.95	1.34
Knights	None	0.24	0.29	0.57	1.89	0.30	0.43

1.b. Phosphorus and potassium

1.b. Phosphorus and potassium										
Group n	neans (N=4)	PJune	PAug	PSep	KJune	KAug	KSep			
Elveden	Mustard 99	99.00	113.00	87.50	175.50	145.00	156.75			
Elveden	None	92.25	106.50	79.75	162.25	155.75	131.75			
Knights	Mustard 99	44.50	53.00	38.00	87.50	81.00	88.00			
Knights	None	43.75	54.00	36.00	79.25	81.75	82.75			
El∨eden	Mustard 119	76.25	93.75	77.25	141.00	130.75	147.25			
Elveden	None	79.25	91.00	72.00	157.50	147.50	132.75			
Knights	Mustard 119	43.75	52.50	41.00	74.50	102.00	86.75			
Knights	None	46.50	57.25	40.00	111.25	80.25	92.25			
Standard errors		PJune	PAug	PSep	KJune	KAug	KSep			
Elveden	Mustard 99	4.18	8.22	5.91	10.91	9.68	13.57			
Elveden	None	1.44	1.32	4.91	14.10	6.60	10.26			
Knights	Mustard 99	1.94	1.08	0.91	0.50	5.34	. 11.11			
Kn ights_	None	0.85	1.41	1.08	6.65	7.16	1.44			
Elveden	Mustard 119	2.39	2.50	2.06	3.51	9.82	5.38			
El∨eden	None	3.42	3.74	1.58	18.75	13.22	14.50			
Knights	Mustard 119	1.65	1.55	1.08	2.10	20.55	4.44			
Kn ights	None	2.22	1.11	1.08	12.57	5.12	7.81			

1.c. Magnesium and pH

Group m	eans (N=4)	MgJune	MgAug	MgSep	pHJune	pHAug	pHSep
					•		
Elveden	Mustard 99	91.50	87.25	83.50	7.20	7.29	6.65
Elveden	None	82.00	80.50	79.00	7.20	7.11	6.74
Knights	Mustard 99	44.50	42.25	35.75	7.80	7.74	7.50
Knights	None	42.50	47.00	37.75	7.69	7.74	7.53
Elveden	Mustard 119	85.25	91.50	84.00	7.33	7.35	6.91
Elveden	None	92.75	87.00	84.00	7.38	7.03	6.93
Knights	Mustard 119	48.75	53.75	45.25	7.48	6.98	7.08
Knights	None	49.75	55.25	48.00	7.46	6.99	7.06
Standard errors		MgJune	MgAug	MgSep	pHJune	pHAug	pHSep
Elveden	Mustard 99	4.33	6.79	4.77	0.04	0.09	0.03
Elveden	None	4.18	2.99	1.22	0.04	0.08	0.06
Knights	Mustard 99	3.66	1.03	1.49	0.04	0.01	0.00
Knights	None	0.96	3.11	0.85	0.04	0.02	0.08
Elveden	Mustard 119	7.22	3.07	3.29	0.03	0.05	0.04
Elveden	None	2.25	1.08	4.56	0.02	0.03	0.07
Knights	Mustard 119	3.42	4.59	3.75	0.13	0.19	0.21
Knights	None	3.71	2.84	5.51	0.17	0.17	0.23

To compare the difference between the August and September soil analyses, statistical analysis was by Analysis of variance (Anova) with Mustard 99/Mustard 119 as the two treatments and the two sites as blocks. The Mustard 99 and Mustard 119 trials were analysed separately.

Analyses were performed on both the change between August and September and the proportionate change. Examination of the residuals in the analyses of proportionate values showed no sign of non-normality and hence transformation was not necessary. A summary of the outcome is given in Table 2.

Table 2. Comparisons between Crop and Nocults (Anova, Genstat8)

Variate is the change between August and September, either as percentage or actual Percentage (e.g. %23K) calculated as (Sep-Aug)/Aug\*100
Actual (e.g. diff23K) calculated as Sep-Aug
Difference between the changes calculated as Mustard 99-NoCult

2.a. Mustard 99

Variate	Mean o Mustard	hange						
	99	NoCult	SE	difference	SE diff	F (df=1,13)	р	
%23K	7.8	-6.3	4.6	14.1	6.5	4.73	0.049	*
%23Mg	-9.5	-10.0	3.1	0.5	4.3	0.01	0.924	
%23N30	64.0	-21.0	17.1	85.0	24.2	12.59	0.004	**
%23N60	169.0	-10.0	19.9	179.0	28.1	40.85	<0.001	***
%23P	-25.3	-29.2	2.2	3.9	3.1	1.58	0.231	
%23pH	-5.89	-4.00	0.65	-1.89	0.92	4.22	0.061	

	Mustard							
2	99	NoCult	SE	difference	SE diff	F (df=1,13)	р	
diff23K	9.4	-11.5	5.5	20.9	7.7	7.31	0.018	
diff23Mg	-5.1	-5.4	2.0	0.3	2.9	0.01	0.932	
diff23N30	5.4	-6.2	2.7	11.6	3.9	8.94	0.01	**
diff23N60	9.3	-3.8	2.0	13.1	2.9	20.37	<0.001	***
diff23P	-20.2	-22.4	2.7	2.2	3.7	0.32	0.58	
diff23pH	-0.44	-0.29	0.05	-0.14	0.07	4.14	0.063	

# 2.b. Mustard

Mean change Mustard							
119	NoCult	SE	difference	SE diff	F (df=1,13)	р	
6.1	3.4	10.4	2.7	14.6	0.03	0.855	
-12.0	-8.6	2.9	-3.4	4.1	0.66	0.433	
116.0	-11.0	23.4	127.0	33.0	17.73	0.002	**
131.0	60.0	37.6	71.0	53.1	1.79	0.204	
-19.6	-25.4	1.7	5.8	2.4	6.09	0.028	*
-2.25	-0.20	0.85	-2.05	1.21	2.89	0.113	
0.6	-1.4	10.9	2.0	15.4	0.02	0.899	
-8.0	-5.1	2.0	-2.9	2.8	1.05	0.324	
7.5	-6.3	2.8	13.8	4.0	12.17	0.004	**
6.4	2.8	2.6	3.6	3.7	0.98	0.341	
-14.0	-18.1	1.6	4.1	2.3	3.21	0.096	
-0.17	-0.01	0.06	-0.16	0.09	3.21	0.097	
	Mustard 119 6.1 -12.0 116.0 131.0 -19.6 -2.25 0.6 -8.0 7.5 6.4 -14.0	Mustard 119 NoCult 6.1 3.4 -12.0 -8.6 116.0 -11.0 131.0 60.0 -19.6 -25.4 -2.25 -0.20  0.6 -1.4 -8.0 -5.1 7.5 -6.3 6.4 2.8 -14.0 -18.1	Mustard 119 NoCult SE 6.1 3.4 10.4 -12.0 -8.6 2.9 116.0 -11.0 23.4 131.0 60.0 37.6 -19.6 -25.4 1.7 -2.25 -0.20 0.85  0.6 -1.4 10.9 -8.0 -5.1 2.0 7.5 -6.3 2.8 6.4 2.8 2.6 -14.0 -18.1 1.6	Mustard         SE         difference           6.1         3.4         10.4         2.7           -12.0         -8.6         2.9         -3.4           116.0         -11.0         23.4         127.0           131.0         60.0         37.6         71.0           -19.6         -25.4         1.7         5.8           -2.25         -0.20         0.85         -2.05           0.6         -1.4         10.9         2.0           -8.0         -5.1         2.0         -2.9           7.5         -6.3         2.8         13.8           6.4         2.8         2.6         3.6           -14.0         -18.1         1.6         4.1	Mustard         SE         difference         SE diff           6.1         3.4         10.4         2.7         14.6           -12.0         -8.6         2.9         -3.4         4.1           116.0         -11.0         23.4         127.0         33.0           131.0         60.0         37.6         71.0         53.1           -19.6         -25.4         1.7         5.8         2.4           -2.25         -0.20         0.85         -2.05         1.21           0.6         -1.4         10.9         2.0         15.4           -8.0         -5.1         2.0         -2.9         2.8           7.5         -6.3         2.8         13.8         4.0           6.4         2.8         2.6         3.6         3.7           -14.0         -18.1         1.6         4.1         2.3	Mustard         SE         difference         SE diff         F (df=1,13)           6.1         3.4         10.4         2.7         14.6         0.03           -12.0         -8.6         2.9         -3.4         4.1         0.66           116.0         -11.0         23.4         127.0         33.0         17.73           131.0         60.0         37.6         71.0         53.1         1.79           -19.6         -25.4         1.7         5.8         2.4         6.09           -2.25         -0.20         0.85         -2.05         1.21         2.89           0.6         -1.4         10.9         2.0         15.4         0.02           -8.0         -5.1         2.0         -2.9         2.8         1.05           7.5         -6.3         2.8         13.8         4.0         12.17           6.4         2.8         2.6         3.6         3.7         0.98           -14.0         -18.1         1.6         4.1         2.3         3.21	Mustard         119         NoCult         SE         difference         SE diff         F (df=1,13)         p           6.1         3.4         10.4         2.7         14.6         0.03         0.855           -12.0         -8.6         2.9         -3.4         4.1         0.66         0.433           116.0         -11.0         23.4         127.0         33.0         17.73         0.002           131.0         60.0         37.6         71.0         53.1         1.79         0.204           -19.6         -25.4         1.7         5.8         2.4         6.09         0.028           -2.25         -0.20         0.85         -2.05         1.21         2.89         0.113           0.6         -1.4         10.9         2.0         15.4         0.02         0.899           -8.0         -5.1         2.0         -2.9         2.8         1.05         0.324           7.5         -6.3         2.8         13.8         4.0         12.17         0.004           6.4         2.8         2.6         3.6         3.7         0.98         0.341           -14.0         -18.1         1.6         4.1

## K

There was a significant difference between Mustard 99 and NoCult in the change in K between August and September (7.8% increase for Mustard 99 but 6.3% drop for NoCult) but this did not occur in the Mustard 119 trial.

#### Mg

No statistically significant differences - similar approximate 10% decrease for Mustard 99 and Nocult and also Mustard 119 and NoCult.

#### N30

Large increase with both Mustard 99 and Mustard 119, versus small loss with NoCult. All comparisons statistically significant.

#### **N60**

With Mustard 99 there was a statistically significant difference between the large increase with Mustard 99 and the small loss with NoCult (this difference was more pronounced than that with N30). However, although there was also a large increase with Mustard 119, there was also a (smaller) increase with NoCult, resulting in there being no statistically significant difference between Mustard 119 and NoCult.

Р

No statistically significant difference with Mustard 99, but with Mustard 119 the percentage change (%23P) differed but not the actual change (diff23P).

### pH.

No statistically significant differences with either Mustard 99 or Mustard 119.

The initial values in June were compared by Anova (Genstat) to see if they differed between the Crop and NoCult plots (Table 3).

Table 3. Comparisons between Crop and NoCult initial June values (Anova)

Difference calculated as Crop-NoCult

3.a. Mustard 99

Variate	Means						
	Mustard 99	NoCult	SE	difference	SE diff	F (df=1,13)	р
N30June	3.7	3.8	0.5	-0.2	0.7	0.08	0.785
N60June	2.3	2.6	0.2	-0.3	0.3	0.70	0.418
PJune	71.8	68.0	1.8	3.8	2.5	2.25	0.157
KJune	131.5	120.8	6.5	10.7	9.2	1.37	0.262
MgJune	68.0	62.2	2.5	5.8	3.6	2.59	0.131
pHJune	7.50	7.44	0.03	0.06.	0.04	1.95	0.186

#### 3.b. Mustard 119

Variate	Means Mustard							
	119	NoCult	SE	difference	SE diff	F (df=1,13)	р	
N30June	4.0	3.6	0.3	0.4	0.4	0.95	0.349	
N60June	2.6	2.4	0.2	0.1	0.3	0.15	0.703	
PJune	,60.0	62.9	1.7	-2.9	2.4	1.43	0.254	
KJune	107.8	134.4	8.0	-26.6	11.4	5.48	0.036	
MgJune	67.0	71.2	3.2	-4.2	4.5	0.91	0.358	
pHJune	7.40	7.42	0.07	-0.02	0.11	0.03	0.862	

There were no statistically significant differences between Crop and NoCult except for K in the Mustard 119 trial, where the mean level was higher in the NoCult plots than in the Mustard 119 plots.

The difference between N at 30 and 60cm was compared between treatments at each time point (Table 4). The difference was calculated as N60cm-N30cm (i.e. was not proportional).

Table 4. Comparisons between N at 30 and 60cm

Difference calculated as N60-N30

Variate	Mean o	lifference						
	Crop	NoCult	SE	difference	SE diff	F (df=1,13	3) p	
Mustard 99							, '	
diffNJune	-1.4	-1.3	0.4	-0.1	0.5	0.02	0.890	
diffNAug	-3.1	-4.4	2.1	1.3	3.0	0.19	0.670	
diffNSep	0.7	-2.0	2.1	2.7	3.0	0.83	0.379	
Mustard 119								
diffNJune	-1.2	-1.4	0.3	0.2	0.5	0.21	0.651	
diffNAug	-6.5	-0.3	1.9	-6.2	2.6	5.51	0.035	*
diffNSep	2.5	-1.4	1.9	3.9	2.7	2.13	0.168	

There was no statistical difference between the treatments (Crop,NoCult) in the difference in N between 30 and 60cm at any time with Mustard 99, nor in June and September with Mustard 119, but there was a difference (just) with Mustard 119 in August (difference greater in Crop than NoCult).

Table 5. Effect of treatment (Biocide or NoCult) on the change in nutrients between June and August.

August v	_June							
	Biocide	NoCult	S.E.			F		
Variate	Aug-June	Aug-June	Aug-June	Difference	S.E. difference	(df=1,13)	р	
Mustard 9	9 Mustard 99	NoCult						
%12K	-12.2	8.0	4.66	-13	6.59	3.87	0.071	n.s
%12Mg	-3.6	4.7	5.1	-8.3	7.21	1.33	0.270	n.s
%12N30	166 📖	545	95.4	-379	134.9	7.88	0.015	*
%12N60	173	603	68.4	-430	96.7	19.8	<.001	***
%12P	16.8	19.5	2.86	<b>-2</b> .7	4.04	0.47	0.505	n.s.
%12pH	0.21	-0.28	0.634	0.49	0.896	0.30	0.592	n.s.
diff12K	-18.5	-2	5.88	-16.5	8.31	3.94	0.069	n.s.
diff12Mg	-3.2	1.5	2.57	-4.7	3.64	1.70	0.215	n.s.
diff12N30	5.4	18	2.74	-12.6	3.87	10.7	0.006	**
diff12N60	3.59	14.88	1,315	-11.29	1.86	36.83	<.001	***
diff12P	11.2	12.2	2.3	-1	3.25	0.09	0.764	n.s.
diff12pH	0.012	-0.019	0.0463	0.031	0.0655	0.23	0.641	n.s.
Mustard 119	Mustard 119	NoCult		_	<del></del>	. <u>.</u>	·	
%12K	16.4	-15.5	13.19	31.9	18.65	2.93	0.111	n.s.
%12Mg	10.7	2.9	6.09	7.8	8.61	0.83	0.378	n.s.
%12N30	68	417	67.7	-349	95.8	13.29	0.003	**
%12N60	177	520	141	-343	199.4	2.96	0.109	n.s.
%12P	21.7	19.3	2.66	2.4	3.76	0.41	0.531	n.s.
%12pH	3.19	-5.55	0.854	2.36	1.207	3.82	0.073	n.s.
diff12K	8.6	-20.5	11.69	29.1	16.53	3.11	0.102	n.s.

Variate	Biocide Aug-June	NoCult Aug-June	S.E. Aug-June	Difference S	S.E. difference	F e (df=1,13)	р	
diff12Mg	5.6	-0.1	3.81	5.7	5.39	1.14	0.305	n.s.
diff12N30	2.8	15.4	2.3	-12.6	3.25	15.23	0.002	**
diff12N60	3.9	10.1	2.02	-6.2	2.86	4.80	0.047	*
diff12P	13.12	11.25	1.412	1.87	1.997	0.88	0.365	n:s.
diff12pH	-0.237	-0.413	0.0624	0.176	0.0883	3.93	0.069	n.s.

Conclusion: The change in nutrients between June and August did not differ between Biocide and NoCult for K, Mg, P or PH (all p>0.05) but did differ for N at both 30cm and 60cm with the change in level being greater in the NoCult plots than in the Biocide plots.

Table 6. Effect of treatment (Biocide or NoCult) on the change in nutrients between June and September.

September	v June							•
	Biocide							
	Sept-	NoCult	S.E.		S.E.	F		•
Variate	June	Sept-June	Sept-June	Difference	difference	(df=1,13)	p	
Mustard								
99	Mustard							
99	99	NoCult						
%13K	-5.1	-6.1	5.6	1	7.92	0.02	0.902	n.s.
%13 <b>M</b> g	-13	-7.1	4.03	-5.9	5.7	1.09	0.316	n.s.
%13N30	355	387	103.4	-32	146.2	0.05	0.832	n.s.
%13N60	577	454	77.7	123 ·	109.8	1.25	0.284	n.s.
%13P	-13.1	-15.7	2.1	2.6	2.96	0.80	0.388	n.s.
%13pH	-5.74	-4.26	0.462	-1.48	0.654	5.09	0.042	*
diff13K	-9.1	-13.5	5.59	4.4	7.91	0.31	0.590	n.s.
diff13Mg	-8.4	-3.9	2.18	-4.5	3.09	2,12	0.169	n.s.
diff13N30	10.7	11.8	1.91	-1.1	2.7	0.15	0.703	n.s.
diff13N60	12.9	11.1	1.99	1.8	2.81	0.39	0.545	n.s.
diff13P	-9	-10.1	1.59	1.1	2.24	0.25	0.624	n.s.
diff13pH	-0.425	-0.312	0.0355	-0.113	0.0502	5.01	0.043	*
			. 1165					
Mustard	N. II							
119	Mustard 119	NoCult	30					
%13K	10.6	-14.8	4.92	25.4	6.95	13.32	0.003	**
%13Mg	-2.7	-6.6	5.7	3.9	8.06	0.24	0.634	n.s.
%13N30	258	264	51.9	-6	73.4	0.01	0.935	n.s.
%13N60	421	574	123.1	-153	174.1	0.77	0.397	n.s.
%13P	-2.3	-11.2	1.97	8.9	2.79	10.31	0.007	**
%13pH	-5.52	-5.76	0.647	0.24	0.914	0.07	0.797	n.s.
diff13K	9.2	-21.9	6.86	31.1	9.7	10.31	0.007	**
diff13Mg	-2.4	-5.2	3.81	2.8	5.39	0.28	0.603	n.s.
diff13N30	10.3	9.2	1.55	1.1	2.19	0.26	0.620	n.s.
diff13N60	10.3	12.9	2.23	-2.6	3.15	0.67	0.427	n.s.
diff13P	-0.9	-6.9	1.46	6	2.06	8.48	0.012	*
diff13pH	-0.406	-0.425	0.0453	0.019	0.064	0.09	0.774	n.s.

#### Conclusion:

**Mustard 99**: No effect with K, Mg, N30, N60 or P but significant difference with pH – slightly smaller change (decrease) in NoCult than in Biocide.

**Mustard 119**: No effect with Mg, N30, N60, or pH but significant difference with K (increase with Mustard 119 but decrease with NoCult) and P (greater decrease with NoCult than with Mustard 119).

Table 7. Effect of treatment (Biocide or NoCult) on the level of nutrients in

September.

depterriber.								
	Biocide	NoCult			S.E.	F		
Variate _	September	September	S.E.	Difference	difference	(df=1,13)	р	
Mustard 99	Mustard 99	NoCult						
N30Sep	14.4	15.6	1.61	-1.2	2.28	0.30	0.5930	n.s.
N60Sep	15.1	13.7	2.06	1.4	2.91	0.26	0.6200	n.s.
PSep	62.8	57.9	2.71	4.9	3.84	1.62	0.2260	n.s.
KSep	122.4	107.2	7.18	15.2	10.16	2.22	0.1600	n.s.
MgSep	59.6	58.4	1.88	1.2	2.66	0.22	0.6470	n.s.
pHSep	7.075	7.131	0.0344	-0.056	0.0487	1.33	0.2690	n.s.
		·						
	Biocide	NoCult			S.E.	F	_	
Variate	September	September	S.E.	Difference	difference	(df=1,13)	р	
Mustard 119	Mustard 119	NoCult						
N30Sep	14.3	12.8	1.49	1.5	2.11	0.48	0.4990	n.s.
N60Sep	12.9	15.3	2.21	-2.4	3.12	0.62	0.4450	n.s.
PSep	59.12	<sub>,</sub> 56	1.104	3.12	1.562	4.00	0.0670	n.s.
KSep	117	112.5	6.38	4.5	9.03	0.25	0.6270	n.s.
MgSep	64.6	66	2.97	-1.4	4.21	0.11	0.7490	n.s.
pHSep	6.994	6.994	0.1096	0 -	0.155_	0.00	1.0000	n.s.

Conclusion: No significant differences detected.

Table 8. Effect of cultivation (Cult or NoCult) on the level of Nematodes (and

Pythium) at Count 2.

ryinium) at	Count Z.								
Variate	Cult Count2	NoCult Count2	S.E.	Diffèrence	S.E. difference	F (df=1,13)	р		
Count2									
Mustard 99	Cult	NoCult			<del></del>				
logLesion2	1.726	1.851	0.0875	-0.125	0.1237	1.01	0.333	n.s	
logStubby2	0.405	0.39	0.0865	0.015	0.1223	0.01	0.907	n.s	
logStunt2	0.6	0.38	0.167	0.22	0.236	0.82	0.381	n.s	
logTotNem2	1.799	1.961	0.0934	-0.162	0.1321	1.50	0.242	n.s	
logTotPyth2	3.414	3.565	0.0422	-0.151	0.0596	6.42	0.025	*	
Variate	Cult Count2	NoCult Count2	S.E.	Difference	S.E. difference	F (df=1,13)	р		
Count2									
Mustard 119	Cult	NoCult	<del>,</del>						
logLesion2	1.75	1.608	0.0854	0.142	0.1207	1.40	0.258	n.s	
logStubby2	0.218	0.285	0.0948	-0.067	0.1341	0.25	0.626	n.s	

Variate	Cult Count2	NoCult Count2	S.E.	Difference	S.E. e difference	F (df=1,13)	р	
logStunt2	0.52	0.56	0.158	-0.04	0.224	0.03	0.858	n.s
logTotNem2	1.783	1.686	0.0789	0.097	0.1116	0.76	0.398	n.s
logTotPyth2	3.391	3.385	0.0428	0.006	0.0606	0.01	0.917	n.s

Conclusion: No significant effect on any of the nematode groups.

Small and just significant effect of cultivation on Pythium in Mustard 99 part of the trial but not in the Mustard 119 part.